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BIOTECH HORIZONS: TRANSLATIONAL RESEARCH IN HUMAN HEALTH

DEPARTMENT OF BIOTECHNOLOGY
SCHOOL OF LIFE SCIENCES
SWAMI VIVEKANANDA UNIVERSITY

- ✓ Antimicrobial resistance
- ✓ Biomedical Research
- ✓ Therapeutic Potential
and Mechanistic Insights

**EDITOR: DR. SEMANTI
GHOSH**

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Biotech Horizons: Translational Research in Human Health

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Declaration by the Editor

I hereby declare that the book titled “**Biotech Horizons: Translational Research in Human Health**” has been edited under my supervision. I have carefully reviewed the manuscript for clarity, coherence, structure, and language, and ensured that it meets the required academic and publishing standards.

To the best of my knowledge, the content of this book is original and has not been published elsewhere in the same form. All sources used by the author(s) have been duly acknowledged, and any necessary permissions have been obtained where applicable.

I confirm that this book is suitable for publication and dissemination.

Acknowledgments

First and foremost, I would like to praise and thank God, the Almighty, who has granted countless blessing, knowledge, and opportunity to accomplish the book project work. Thanks to all the authors of the various chapters for their contributions. It had been a bit of a long process from the initial outlines to developing the full chapters and then revising them in the light of reviewer's comments. I sincerely acknowledge the author's willingness to go through this process. I also acknowledge the work and knowledge of the members of our review panels, many of which had to be done at short notice. I also thank the publishing team for their professional support throughout the editorial process. My appreciation extends to colleagues and mentors in the field of Biotechnology whose guidance and discussions have enriched this work. Finally, I acknowledge the support of Swami Vivekananda University and all my colleagues for their encouragement and patience during the preparation of this book.

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Preface

The field of biotechnology has emerged as a transformative force in advancing human health, enabling the translation of fundamental biological discoveries into meaningful clinical and therapeutic applications. *Biotech Horizons: Translational Research in Human Health* has been carefully curated to reflect this evolution, bringing together diverse perspectives that highlight the role of translational research in bridging the gap between laboratory science and patient care.

As editor, it has been my objective to assemble contributions that address both the scientific depth and practical relevance of contemporary biotechnological research. The chapters in this volume explore key areas such as molecular and cellular mechanisms of disease, diagnostic and therapeutic innovations, regenerative medicine, genomics, and emerging biotechnological tools that are shaping the future of healthcare. Each contribution emphasizes translational potential, underscoring pathways through which research findings can be effectively applied to improve human health outcomes.

This book brings together the work of researchers, clinicians, and academicians whose collective expertise reflects the interdisciplinary nature of translational biotechnology. The contributors have not only presented current research and reviews but have also critically examined challenges related to clinical validation, ethical considerations, regulatory frameworks, and accessibility of biotechnological advancements.

Biotech Horizons is intended to serve as a valuable resource for postgraduate students, researchers, healthcare professionals, and policymakers seeking insight into the dynamic landscape of translational research. By fostering dialogue across disciplines and encouraging innovation grounded in scientific rigor and social responsibility, this book aspires to contribute meaningfully to the advancement of human health.

Barrackpore, Kolkata.

Semanti Ghosh

About the Editor

Dr. Semanti Ghosh, currently working as Assistant Professor in the Department of Biotechnology,



Swami Vivekananda University, Barrackpore, Kolkata. Dr. Ghosh did her PhD in Biochemistry in 2018 from University of Kalyani in the field of Computational Biology. Then Dr. Ghosh joined Crystallography & Molecular Biology Division at Saha Institute of Nuclear Physics, Kolkata. As a postdoc. Dr. Ghosh received prestigious DBT Research Associateship fellowship of Govt. of India from July, 2018 to June, 2021. Dr. Ghosh worked in the field of protein-protein/DNA/ligand interactions and

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Introduction of correspondence authors



Dr. Pritha Pal is an Assistant Professor in the Department of Microbiology at Swami Vivekananda University, Barrackpore, Kolkata, where she also serves as Head of the department. She holds a Ph.D. and has been teaching and researching since 2020. Her research interests span cancer biology, environmental toxicology (metal remediation), and microbial biotechnology. Dr. Pal has authored numerous scientific publications and contributed to studies on microbial impacts and metal resistance.



Dr. Priyankar Pal is a reproductive toxicologist with over six years of research experience using rodent models, focusing on environmentally induced male reproductive disorders and their amelioration through nutritional antioxidants. He earned his Ph.D. in 2024 with a thesis on fluoride-induced reproductive toxicity and the protective roles of vitamins C and E. His research expertise includes animal handling, histology, histomorphometry, biochemical assays, and molecular techniques such as PCR, ELISA, FACS, and immunofluorescence. Dr. Pal has authored more than eight peer-reviewed research articles and contributed extensively to book chapters as first author, co-author, and corresponding author. He has over three years of teaching experience in Zoology and Biotechnology and is currently serving as an Assistant Professor of Biotechnology. His research interests span reproductive toxicology, oxidative stress, immunotoxicology, and molecular mechanisms of environmental toxicants.



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Dr Bidisha Ghosh is presently serving as an Assistant Professor in the Department of Biotechnology in the School of Life Sciences, in the prestigious Swami Vivekananda University, Barrackpore. She has completed her PhD in Biophysics from Jadavpur University in 2024. Just after her Masters in Biotechnology from the Department of Life Science & Biotechnology, Jadavpur University in 2012. Dr Ghosh served as a Research Trainee in Herbicare Healthcare Bioherbal Research foundation where she continued in the role till 2017. Dr Ghosh has been working with nano materials and her thesis work entails synthesis of varied kinds of nanomaterials and exploring their potential in waste water management along with their biological applications. She has contributed to 37 Peer Reviewed Journals, 48 Book Chapters and 01 Filed Patent (National). She has attended various National and International Conferences and has won several awards in the same. Dr Ghosh has worked as a CO-PI in the ICSSR Vision Viksit Bharat 2047 Collaborative Research Project (funded by Govt. of India) entitled “Post Covid Era and the Zero Generation: Vulnerability, Resilience and Adaptive Capacity in Ageing Communities of Select Urban Spaces of North-east India”. Dr Ghosh is a member of Society of Conservation Biology.



Dr. Debaprasad Koner earned his Ph.D. in Zoology from North-Eastern Hill University, Shillong focusing on the protective role of nitric oxide during stress response. He served as Research Associate-II in a SERB-funded project on hypoxia adaptation.

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Dr. Suranjana Sarkar holds a Bachelor's, Master's, and Ph.D. degree in Microbiology. Her doctoral research was centered on Drug Discovery and Rational Drug Design. Her research is fundamentally grounded in microbiology, with emphasis on microbial pathogenesis, antimicrobial strategies, and the development of therapeutics targeting infectious diseases. She has published in peer-reviewed journals, and her ongoing research interests focus on combinational antimicrobial systems, comparative evaluation of natural and synthetic therapeutics, and systems biology–driven drug–target interaction and safety profiling along with ML-assisted quantum chemical computational based drug designing. Dr. Sarkar is committed to advancing microbiology-driven therapeutic innovation and mentoring emerging researchers in interdisciplinary life sciences. At Swami Vivekananda University, she teaches both Undergraduate and Postgraduate courses in Biotechnology and Microbiology, delivering core and applied subjects including *General Microbiology*, *Microbial Genetics*, *Biochemistry*, *Quality Assurance and Quality Control*, and *Medical Coding & Scribing*. She integrates foundational microbiology with translational and industry-oriented perspectives in her teaching. She is committed to student mentorship, providing personalized academic guidance and professional development support to foster students' holistic growth and research excellence. She effectively balances research and academics, aiming excellence in both scientific inquiry and student learning.



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Dr. Subhasis Sarkar, an Assistant Professor in Microbiology of Swami Vivekananda University, has completed his Ph.D in Biotechnology from University of Calcutta in 2014. After completing this journey, he served the department of Molecular biology & Bioinformatics, Tripura University (A Central University) as Guest Faculty, from September, 2014 till May, 2015. Thereafter, he contributed as Senior Project Fellow at the Institute of Environmental Studies and Wetland Management till 2016. Later on, he served as Assistant Professor in Microbiology in Kingston College of Science together with has served as Guest Faculty in Panihati College and West Bengal State University in various department including Food & Nutrition and Microbiology. In the post PhD journey of 12 Years, he has guided UG, PG and PhD students in different relevant domains of Microbiology and Biotechnology. Till date he has contributed in various in 46 articles in Book Chapters, 51 articles in Peer Reviewed Journals and 01 patent (National). Right Now, his work has been appreciated as evidenced in Google scholar record of 2108 citations with H index 12 and i-10 index 13. During presenting his work in National and International conferences, he has been awarded twice with 2nd Prize (Speaker) in National level Platform. He has been recognized by a couple of prestigious editorial board as reviewer including 3-Biotech, Frontiers in Pharmacology etc. He has been serving as a member of American Society for Microbiology, Society for Conservation Biology (USA) etc.



Dr. Priyajit Banerjee serving as an Assistant Professor in the Department of Biotechnology, School of Life Sciences, Swami Vivekananda University, West Bengal, India. He is involved in teaching advanced biotechnology courses such as Drug Discovery, Protein Engineering, Biophysical Chemistry, and API formulation etc. Dr. Banerjee's doctoral research at the UNESCO–Regional Centre for Biotechnology centered on the structural and functional characterization of bacterial regulatory proteins. His work unravelled key mechanisms underlying motility-to-biofilm transitions in *Pseudomonas aeruginosa* and generated high-impact structural insights, including multiple Protein Data Bank depositions. He is recipient of international award such as David Blow Studentship Bursary, Carl Storm International Diversity Fellowship. He is also recipient of DST-SERB National Postdoctoral Fellowship. His lab currently focusing on Drug Discovery, Protein Engineering, and Mechanistic Toxicological Assessment of compounds integrating both computational and experimental approaches. His work exploring natural and synthetic bioactive molecules, engineers' proteins and enzymes with therapeutic relevance, and evaluates safety through toxicity profiling in biological models. Dr. Banerjee has contributed substantially to the field through publications in leading journals, including Science Advances, Molecular Aspects of Medicine, EBioMedicine, Frontiers in Immunology etc. Beyond research, he is deeply dedicated to teaching and mentorship, inspiring the next generation of life sciences students and researchers at Swami Vivekananda University to pursue transformative scientific discoveries.

Chapter 1:

Bioaccumulation of Arsenic in Potatoes in India: Health Risks to Humans

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Abstract

In India, arsenic contamination of soils and irrigation water poses a serious threat to public health and agriculture, especially in the Ganga-Brahmaputra-Meghna River basin. Tissue-specific deposition is seen in potatoes cultivated in polluted areas; the highest concentration of arsenic is found in the roots, which are followed by the stems, leaves, and tubers. Phosphate transporters allow arsenate (As(V)) to enter roots, where it is transformed into arsenite (As(III)), which disrupts enzyme functions and releases reactive oxygen species (ROS), leading to DNA damage, oxidative stress, and lipid peroxidation. The seriousness of arsenic exposure throughout the food chain is highlighted by the fact that eating contaminated potatoes contributes to long-term health issues in humans, including skin lesions, cancer, cardiovascular disease, and neurological disorders. This review looks at how arsenic enters the food chain, how it transmits from the environment to plants to humans, and the health risks to humans.

Keywords: *Arsenic Contamination; food chain; potatoes; reactive oxygen species;*

Introduction

Arsenic is a toxic heavy metal which have significant role in public health as it is carcinogenic. The main causes of soil contamination with extremely high levels of arsenic are a strong dependency on the use of inorganic fertilisers and pesticides that contain arsenic residues, and the use of arsenic-concentrated groundwater for crop irrigation. Other than that, different anthropogenic activities lead to subsequent accumulation of such hazardous arsenic metalloids into edible plants tissue [1].

Globally, around 66% of the global extracted groundwater focusses on the severity of arsenic poisoning in Asia, notably in South and Southeast Asia. In India arsenic contamination prominently found in aquifers of Ganga-Brahmaputra-Meghna River basin includes West Bengal, Assam, Bihar, Jharkhand, Uttar Pradesh, and other afflicted areas [3]. AAS investigated

and assessed the influence of As-contaminated irrigation water on alkaline soils, as well as the result of As absorption by potatoes and other food crops (onions, cauliflower, eggplant and rice). AAS revealed that the As concentration in irrigation water ranged from <0.005 to 1.014 mg/L and in soils from 6.1 to 16.7 mg/kg. Research has shown the concentration of As in different parts of the crops appeared to be in the sequence of roots > shoots > leaves > edible parts, with the maximum amount of As observed in potato roots [2].

Furthermore, the consumption edible crops grown in highly concentrated arsenic-polluted soils enhances its bio-magnification in several components of the food chain, posing major health hazards as well as threats to global food and nutritional security [1]. The epidemiological studies demonstrated that long exposure to arsenic can cause chronic illness such as, cancer, melanosis, hyperkeratosis, restrictive lung disease, peripheral vascular disease (which is also known as black foot disease), gangrene, diabetes mellitus, hypertension, ischemic heart disease. Studies shows that arsenic promotes cancer rather than initiate it. Due to its widespread and long-term impact WHO classified the arsenic contamination as “the largest mass poisoning in human history” [4].

Tissue-Specific Accumulation of Arsenic in Potato: Arsenic accumulation in potato plant is not uniform across tissue. Research was done to evaluate the arsenic accumulation in potato under a contaminated zone of Eastern India that shows the concentration of arsenic in root (55%) is higher than other part of the plant. At various phases of plant growth, roots have greater arsenic levels than stems, leaves, and tubers. Arsenic levels in stems and leaves were 2.5 times lower than those in roots. Stems (23%) had a greater percentage of arsenic than leaves (19%). Potato tubers (3%) had a significantly lower concentration of this potentially dangerous metalloid than other plant parts. Arsenic buildup in potato plant components was shown to be lower in the edible part compared to other parts. Arsenic dispersion in plants often occurs in the following order: root > shoot [5].

Movement of Arsenic: Arsenic occurs in 200 mineral forms, including arsenates (60%), sulphides and sulfosalts (20%), and tiny quantities of arsenide, arsenates, oxides, silicates, and As in their natural form. Arsenic released into the environment through two primary pathways: natural processes and industrial activity. Natural processes like weathering, hydrothermal ore deposits, volcanic eruptions, geothermal activities, forest fires, wind-blown dust, and sea salt spray can release arsenic into the environment, which can travel long distances as suspended particulates or gaseous forms in water or air. Industrial processes emit significant amounts of As and other trace metals into the environment, contaminating soils, waters, and air worldwide. As cycling occurs because of interactions between natural water, bedrocks, sediments, and soils, as well as local atmospheric deposition. Weathering and leaching of geological formations and

mining wastes cause high As concentrations in natural water sources in several areas [6]. Plants take up the As, which accumulates in the edible sections present in the soil. As(V) is the most prevalent As species in aerobic soils, and it easily enters plant roots via phosphate (Pi) transporters because its oxyanion chemical structure is physically similar to that of Pi. So far, a number of characterised high-affinity Pi transporters have been classified in the Pi transporter 1 (Pht1) family, some of which are also involved in As (V) transport in plants. As (III) is the most common type of As in anaerobic conditions like submerged soils. Significant progress has been made in discovering and characterising the proteins that control As(III) absorption in plants. Plant aquaporins, membrane channels that transport water and tiny neutral molecules, have been demonstrated to be particularly important in the transport of metalloids, including As(III) [7]. These arsenics accumulate inside the tissue of edible part of potato plant. Consumption of the edible part of potatoes introduce arsenic in human body.

Contaminated Groundwater



Soil Contamination (through leaching & irrigation)



Potato Plant Uptake

|----- Arsenate [As(V)] → via phosphate transporter → converted to As(III)

ROS generation → DNA damage, lipid peroxide, enzyme inactivation

|___ Arsenite [As (III)] → via aquaporins → ROS generation → DNA damage, lipid peroxidation, enzyme

Inactivation



Edible Part (Potato Tuners) - lower As
than other tissues but still present



Human Consumption- Longer period



Chronic Health Effects:

- Skin lesions & pigmentation changes
- Multiple cancers (skin, bladder, lung, kidney, liver, prostate)
- Cardiovascular diseases
- Neurological disorders
- Diabetes and systemic organ damage

Fig1: Transmission of Arsenic from Groundwater to Potato to Food Chain and its effect on human health

Mechanisms of Arsenic Toxicity in Plant: Arsenic is not a nutrient and has been linked to harmful effects on plants. Arsenic types and toxicity varies by species. As(V) is readily absorbed by plant roots through phosphate metabolism, but As(III) interacts with sulfhydryl groups, impairing catalytic activities. Plants absorb As(V), which is then converted to As(III) by the reductase enzyme. However, the harmful effects of As(V) in plants may be attributed to its conversion into As(III). Exposure to As induces lipid peroxidation, leading to plant death [8]. Many studies have found that As availability in the soil might impair the morphological and physio-biochemical functioning of plants, lowering yields from agriculture. For example, plants exposed to As exhibited discolouration, lignification, and plasmolysis of root cells, resulting in reduced plant development. ROS production is a frequent response to both abiotic and biotic stressors. ROS overproduction impairs plant health under stress by interfering with a variety of physiological processes such as lipid metabolism, DNA, photosynthesis, respiration, enzyme deactivation, and growth retardation. Several investigations have shown that As(III) and As(V) generate ROS, including superoxide ($O_2^{\bullet-}$), hydroxyl radical ($OH^{\bullet}$), and H_2O_2 . As(III) is more harmful to plant development and produces more $O_2^{\bullet-}$ than As(V), which generates more H_2O_2 . Although root cells perceive As first, the formation of ROS in leaves began well before As buildup in the leaf tissues, suggesting that root cells convey As toxicity to leaves, most likely via H_2O_2 . Under aerobic conditions, As(V) is the primary form that enters plant roots via phosphate transporters and is converted within the cell to As(III), the primary cause of ROS formation. The conversion of As(V) to As(III) is both enzymatic and nonenzymatic. Arsenate reductase (glutaredoxin) mediates an enzymatic process in which GSH functions as an electron donor. This reduction is followed by a methylation process, which yields MMA, DMA, TETRA, and TMAO, as well as arsenocholine, arsenobetaine, and arseno-sugars. These methylation compounds react immediately with molecular oxygen, generating ROS. ROS-induced lipid peroxidation is primarily measured by the presence of malondialdehyde (MDA), a major byproduct of lipid peroxidation, as well as membrane leakage. Overproduction of ROS increases polyunsaturated fatty acids (PUFA) and decreases saturated fatty acids in membrane lipids and fluidity, leading to membrane leakage. ROS also has an effect on enzyme and protein structure and activity by oxidising side chains, cross-linking them, and causing backbone fragmentation. ROS formation under As-stress also modifies nitrogenase base, nucleotide deletion, affects protein-DNA interaction, and may cause DNA breaks [9]. As(V) can be transported across cellular membranes by phosphate transporters, causing phosphate supply imbalances. It can compete with phosphate in phosphorylation processes, forming unstable and short-lived As(V) adducts. For example, the synthesis and quick autohydrolysis of As(V)-ADP initiates a futile cycle that separates

photophosphorylation and oxidative phosphorylation, reducing cells' capacity to make ATP and carry out normal metabolism. As(III) is a dithiol reactive chemical that binds to and may inactivate enzymes with closely spaced cysteine residues or dithiol cofactors. Arsenic exposure normally causes the production of reactive oxygen species, which can result in the synthesis of antioxidant metabolites and a variety of enzymes involved in antioxidant defence. As exposure has an influence on oxidative carbon metabolism, amino acid and protein interactions, as well as nitrogen and sulphur assimilation pathways [10].

Disease Prevalence: Approximately 200 million people worldwide are vulnerable to arsenic poisoning through groundwater. This has resulted in a variety of health problems in the population, including skin illnesses, anaemia, bronchitis, gastrointestinal issues, hormonal imbalance, and cancer. A study has been done in Bihar that shows Acute arsenic poisoning causes vomiting, diarrhoea, and stomach discomfort, followed by extremity paraesthesia, muscular cramps, and, in the most severe instances, death. Long-term exposure to high amounts of inorganic arsenic causes pigmentation changes in the skin, followed by skin lesions and hard patches on the palms and soles of the feet. Long-term exposure to high amounts of inorganic arsenic can also cause peripheral neuropathy, renal system effects, gastrointestinal symptoms, diabetes, high blood pressure, conjunctivitis, an enlarged liver, bone marrow depression, erythrocyte destruction, and cardiovascular disease. Arsenic can cause cancer in the skin, bladder, lungs, kidney, liver, and prostate [11]. Among the multiple genotoxic effects of arsenic in humans, chromosomal aberration and increased frequency of micronuclei in diverse cell types have been identified as noteworthy. Several plausible mechanisms have been proposed to explain DNA damage caused by persistent arsenic exposure. The findings of the West Bengal study imply that arsenic-induced illness manifestation in humans may be caused by a lack of DNA repair ability, disruption of methylation of the promoter area of the p53 and p16 genes, and genomic methylation modification. P53 polymorphism has been linked to an increased risk of arsenic-induced keratosis. In one study, single-nucleotide polymorphisms of purine nucleoside phosphorylase, which are important in the control of arsenic metabolism, were found to increase the prevalence of arsenicosis [12].

Conclusion

The bioaccumulation of arsenic (As) in the plants grown under contaminated soils is one of the major risks to crop productivity and human health worldwide, especially in India where As- rich groundwater is widely used for irrigation. This is an important route of arsenic exposure in humans as consumption rates show that numerous people are at risk due to the relatively

moderate levels of individual exposure. Although potato tubers contain less concentration of As than root, leaves and stem, their high consumption rates make them important route of chronic exposure in humans. The uptake and accumulation of Arsenic is physiologically complex processes. Arsenate (As(V)) enters the plant through Pi transporters and arsenite (As (III)) transported through aquaporins. After the absorption, As(V) is reduced to As(III) which interacts with the sulfhydryl groups, disrupt enzymatic functions and also trigger oxidative stress via excessive production of reactive oxygen species (ROS). These mechanisms lead to DNA damage, lipid peroxidation, chromosomal aberration, impaired metabolic pathways, ultimately hampering the plant growth and yield.

High consumption of contaminated potatoes for longer period of time can cause different disease in human such as, skin disorders, cancers, cardiovascular disease, diabetes, neurological disorders and lung diseases. This risk is amplified in rural communities reliant on locally produced crops. Effective mitigation measure should be taken. Initiating the use of Arsenic-safe water for irrigation, explore different soil remediation strategies, and aware public about Arsenic contamination. To keep an eye on pollution levels, enforce safety rules, and come up with long-term farming methods, lawmakers, scientists, and public health officials need to work together. To stop the cycle of toxins moving from the environment to people, to ensure food security, and protect public health, it is important to deal with arsenic pollution.

Reference:

1. Das, C.K. *et al.* (2023). Bioaccumulation of Arsenic in Different Crop Plants and Its Remediation Using Molecular Breeding Tools and Biotechnological Interventions. In: Kumar, N., Kumar, S. (eds) Arsenic Toxicity Remediation: Biotechnological Approaches. Environmental Science and Engineering. Springer, Cham. https://doi.org/10.1007/978-3-031-37561-3_9.
2. Stasinos, S., Nasopoulou, C., Tsikrika, C., & Zabetakis, I. (2014). The bioaccumulation and physiological effects of heavy metals in carrots, onions, and potatoes and dietary implications for Cr and Ni: a review. *Journal of food science*, 79(5), R765–R780. <https://doi.org/10.1111/1750-3841.12433>.
3. Marghade, D., Mehta, G., Shelare, S., Jadhav, G., & Nikam, K. C. (2023). Arsenic contamination in Indian groundwater: from origin to mitigation approaches for a sustainable future. *Water*, 15(23), 4125.

4. Bhattacharya, P., Samal, A. C., Majumdar, J., & Santra, S. C. (2010). Arsenic contamination in rice, wheat, pulses, and vegetables: a study in an arsenic affected area of West Bengal, India. *Water, Air, & Soil Pollution*, 213(1), 3-13.
5. Rajib Kundu, R. K., Aparajita Majumder, A. M., & Sukanta Pal, S. P. (2012). Evaluation of potato cultivars against arsenic accumulation under an arsenic-contaminated zone of eastern India.
6. Nriagu, J. O., Bhattacharya, P., Mukherjee, A. B., Bundschuh, J., Zevenhoven, R., & Loeppert, R. H. (2007). Arsenic in soil and groundwater: an overview. *Trace Metals and other Contaminants in the Environment*, 9, 3-60.
7. Li, N., Wang, J., & Song, W. Y. (2016). Arsenic uptake and translocation in plants. *Plant and Cell Physiology*, 57(1), 4-13.
8. Farooq, M. A., Islam, F., Ali, B., Najeeb, U., Mao, B., Gill, R. A., ... & Zhou, W. (2016). Arsenic toxicity in plants: cellular and molecular mechanisms of its transport and metabolism. *Environmental and Experimental Botany*, 132, 42-52.
9. Nahar, Kamrun & Rhaman, Mohammad & Parvin, Khursheda & Bardhan, Kirti & Marques, Deyvid & García-Caparrós, Pedro & Hasanuzzaman, Mirza. (2022). Arsenic-Induced Oxidative Stress and Antioxidant Defense in Plants. *Stresses*. 2. 179-209. 10.3390/stresses2020013.
10. Finnegan, P. M., & Chen, W. (2012). *Arsenic toxicity: the effects on plant metabolism*. *Front Physiol* 3: 182.
11. Rahman, M. S., Kumar, A., Kumar, R., Ali, M., Ghosh, A. K., & Singh, S. K. (2019). Comparative Quantification Study of Arsenic in the Groundwater and Biological Samples of Simri Village of Buxar District, Bihar, India. *Indian journal of occupational and environmental medicine*, 23(3), 126–132. https://doi.org/10.4103/ijoom.IJOEM_240_18.
12. Guha Mazumder, D., & Dasgupta, U. B. (2011). Chronic arsenic toxicity: studies in West Bengal, India. *The Kaohsiung journal of medical sciences*, 27(9), 360–370. <https://doi.org/10.1016/j.kjms.2011.05.003>.

Chapter 2:

From Inheritance to Complexity: A New Perspective on X-Linked Diseases

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Abstract

The greatest number of immune-related genes, which are essential for immunological control and autoimmunity are found on the X chromosome. Due to factors such as X chromosome inactivation (XCI), duplication, or monosomy, there is increasing evidence of X-linked but autosomal gene abnormalities, primarily in women. Turner syndrome, which has a single X chromosome and is crucial for immunological homeostasis, is one of the rare X-linked immune abnormalities. The function of X-linked miRNAs in autoimmune susceptibility is almost being examined. The intricacy is shown by a number of ailments. Only Alport syndrome (COL4A5) showed evidence of a significant genotype-phenotype association; that is, truncating mutations around the gene 5' end were linked to a lower age at End-Stage Kidney Disease (ESKD) and a higher incidence of sensory impairments. Males with mosaic present with fever-epilepsy and a new cellular interference mechanism, whereas females have X-linked PCDH19 mutations. The primarily neurological condition known as Rett syndrome (MECP2 mutations) includes lipid metabolism and raises the possibility that MECP2 plays a wider function. Fabry disease, which is characterized by a deficiency of α -galactosidase A due to XCI, varies significantly in females and causes systemic issues. While genetic counseling presents challenges, enzyme replacement therapy (ERT) shows promise. Together, these conditions challenge conventional notions of heredity and have an impact on precision medicine by demonstrating how the X chromosome is essential for immunological, neurological, and metabolic control.

Keywords: X chromosome, Autoimmunity, Alport syndrome, PCDH19, RETT syndrome, Fabry Disease

INTRODUCTION

Sexual dualism where organisms are clearly male or female. Generally, in humans, women are homogametic (XX) and men are heterogametic (XY). At the time of reproduction, eggs carry an X, while sperm carry X or Y, determining the gender of next generation. Recently, rapid advances in human genetics through genome-wide association studies (GWAS) and research on X chromosome abnormalities which have revealed strong links between X-linked gene defects and autoimmune diseases (AID) [Selmi C, 2010]. It focuses on how X-linked gene defects - abnormal gene dosage, are progressively linked to autoimmunity. These findings are modifying our understanding of AID and pointing to new disease pathways [Invernizzi P, 2009].

X-linked Alport syndrome (XLAS) is a monogenic inherited kidney disorder which is caused by mutations in the COL4A5 gene, that accounts for 80% of cases [Gregory MC, 2007]. As the result, the symptoms are seen - kidney damage, loss of hearing, and abnormalities of eye. Due to more than of 440 COL4A5 mutations, XLAS is shown a wide clinical and genetic variability. Age of renal failure onset and hearing loss are distinct among individuals. Genetic testing is important for diagnosis and prognosis. A U.S. study is shown that 681 people from 175 families provided fundamental insights into genotype-phenotype correlations, helping better understanding and management of XLAS [Bekherinia MR, 2010].

In 2008, mutations in PCDH19 were first discovered in large families with women patients producing variable degrees of academic abnormalities and contraction. Earlier this condition was identified as a woman-only mental retardation and contraction that is now named as early infantile epileptic encephalopathy 9 (EIEE9), Juberg-Hellman Syndrome, that was spread by asymptomatic males, which indicate an atypical X-linked inheritance pattern with involvement of females [Dibbens et al., 2008]. PCDH19-related EIEE9 found to be more relevant than initially expected based on their family observations; clinically PCDH19 has happened as the second most significant gene following SCN1A. Maximum patients with mutation in PCDH19 are irregular cases which come from families that have a small number of affected female patients, hindering the inheritance pattern identification. However, assembling of recurrent cases within short intervals are now identified as important features that guide the diagnosis towards PCDH19 [Depienne C, 2012].

Rett syndrome (RTT) which is a neurological disorder caused due to the mutations in the X-linked MECP2 gene, a transcriptional regulator. Generally, it occurs in 1: 10000-15000 live female births [Burd L, 1991]. However, after studying under the light of central nervous system, evidence from mouse models and patients shows that MeCP2 also affects the peripheral systems,

that leads to various phenotypes. Recently studies shows that metabolic dysfunction, especially in lipid metabolism, may rise to RTT symptoms. Understanding MeCP2's various roles all over the body is important for developing effective treatments for this complex incurable disorder. [Stephanie M. Kyle, 2018]

Fabry Disease (FD) is a rare X-linked lysosomal storage disorder due to the deficiency of α -galactosidase A [Brady RO, 1967]. It leads to the glycosphingolipid accumulation in the various body tissues. Hemizygous males are shown few symptoms i.e., pain, renal failure, cardio issues, loss of hearing and strokes. [Sweeley CC, 1963] On the other hand, heterozygous females have variable symptoms due to XCI. FD shortens life expectancy by 20 years in untreated men and 10 in women. Ongoing research inspects oral therapies and upgrades genetic counseling strategies.

This article contains the compiled and cumulative information about the sex-linked inheritance which have not been discussed earlier.

X LINKED INHERITANCE

An X-linked mutation or disorder is caused by where a gene located on the X chromosome that one of the two sex chromosomes in humans. Since males have one X and one Y chromosome (XY), a single altered gene on the X chromosome can cause the condition because there is no second X to provide a normal copy. Females have two X chromosomes (XX), so if one carries the altered gene, the other X often compensates making them carriers who may have mild or no symptoms. X-linked conditions can be recessive or dominant and they often affect males more severely because they have only one X chromosome.

X- LINKED DISORDERS & AUTOIMMUNITY

Most of the genes in the X chromosome are immune related, focusing on AID research. 10% population is affected by AIDs; female predominance is mainly found [Eaton WW, 2007]. Many immunodeficiencies are X linked, related with autoimmune manifestations. X chromosome monosomy or deletions leads to Turner syndrome [Ranke MB, 2001], increase autoimmunity. Moreover, effect of gene dosage like duplication or XCI—is found in women who are related with AID. Researchers are working on the X-linked miRNAs involvement to autoimmunity. X chromosome changes can be a factor in susceptibility of autoimmune and help tells us about its higher chances in female. It is found that the X chromosome has higher amount of miRNA. About 10% of 800 miRNA is found in the human genome in contrast to autosome whereas Y chromosome has no miRNA [Guo X, 2009]. There is a hypothesis that mechanisms are

associated with X chromosome affect X-linked genes as well as X-linked miRNA [Pinheiro I, 2011]. To equalize gene expression level in males and females, XCI has developed. Several X chromosome genes are known to be immune to silencing. When these genes are implicated in immunity, females may have higher quantities of functional immune proteins, which could be directly linked to variations in immunological responses between the sexes. Furthermore, XCI may not be random, and the degree of mosaicism in females may vary depending on how much one of the paternal X chromosomes is selectively inactivated. When taken as a whole, these anomalies may lead to variations in immunological gene expression patterns among females as well as variations in X chromosome gene expression levels between the sexes.

X- LINKED ALPORT SYNDROME (XLAS)

Alport syndrome, nephropathy, hereditary disease followed by failure of kidney, sensorineural hearing loss and changes in eyes can be found [Flinter FA, 1988]. Type IV collagen gene, which is a constituent of base membrane gets mutated and cause this disease [Barker DF, 1990]. There are six α (IV)-chains which are genetically different, corresponding to this gene are situated jointly on X chromosome. Every α (IV) chain have a NC1- domain at C terminal, that is collagenous and contains a repeat of Gly-X-Y forming triple helical structure, and a 7S-domain at N terminal. The $\alpha 1$ & $\alpha 2$ chains are found in basement membranes, the $\alpha 3$, $\alpha 4$ & $\alpha 5$ (IV) chains are distributed little and they are expressed in the inner ear, glomerulus & eye [Hudson BG, 1993]. COL4A5 mutation change expression of $\alpha 5$ (IV)-chain, present at Xq22 [Gregory MC, 2007]. In males end stage renal disease (ESRD) varies by the 2nd and 3rd decade. Although, cases that are not severe up to the 5th or 6th decade ESRD is delayed. Patients have been described to have a wide variety of ophthalmological disorders including maculopathy and cataracts [Kashtan CE, 2000]. There are better-confirmed techniques for screening patient mutations, and genetic testing is likely to provide even further useful prognostic information [Gubler MC, 2007]. The study of XLAS, however, grappling with many families has proven very difficult due to the medical and genetic multitude and the relationship of the phenotype and the mutation of the gene. It's been studied in a European populace [Gross O, 2002]; however, the US population is lacking absolutely.

Clinical Overview

The onset of ESRD varies greatly depending on the type of mutation. Missense mutations showed the latest median onset (37 years), but large and small deletions showed the earliest (22 years). Types of mutation related significantly with ocular changes that were more frequent in

large deletions, splice sites and truncating mutations but less in the missense & small deletions. People with missense mutations demonstrated a significantly lower risk of hearing loss—up to 20 times less—compared to those carrying truncating, splice site, or deletion mutations. The presence of truncating or similar mutations was linked with a markedly increased likelihood of experiencing hearing impairment. Mutations close to the 5' end were highly associated with deafness, vision disorders, and an earlier onset of ESRD (HR 0.766 per 1000-bp shift toward the 3' end; $P < 0.0001$).

Mutation Position on the Basis of Domain Structure of Collagen-5 (IV) Chain Protein

The classification of XLAS mutations was based upon the affected structural domains of the collagen α -5 (IV) chain; thus, a full correlation between phenotype and mutation position was investigated. The patients with mutations in the signal peptide region represented the earliest cases of the disease, at the age of 22 years. This was followed by patients with mutations in the collagenous domain at 29 & finally in the NC1 domain at 36. Narrowing down further within the collagenous domain: the 5' region showed an earlier age of onset for ESRD than the 3' region. *Glycine-X-Y Mutations-* Among patients with Gly-X-Y mutations in the collagenous domain, ESRD appeared to occur somewhat earlier (median 33 years) than in patients with other missense mutations in the same domain ($P = 0.033$). However, there was no significant relationship between mutation position and age at ESRD for Gly-X-Y mutations, nor were there any apparent differences observed in the frequency of other clinical phenotypes for these mutations compared to non-Gly-X-Y mutations.

Concise Methods

Participants & Clinical Criteria- In the study, clinical and genetic data were analyzed for 681 men in generally unaffected families with XLAS from across 175 families with COL4A5 mutations who participated in the University of Utah Alport Syndrome study. Clinical diagnosis was by either family medical history or kidney biopsy findings consistent with AS. Informed consent and approval from Institutional Review Board (IRB) were obtained.

Mutation Analysis- COL4A5 mutations were determined with an array of methodologies including restriction site mapping, RT-PCR, DGGE, RNase protection, allele-specific probes [Barker DF, 1996], direct DNA sequencing, multiplex genomic PCR-SSCP [Barker DF, 2001]. In every family, all these methodologies were used.

Mutation Categories- COL4A5 variations were classified as truncating, missense, splice-site (donor/acceptor), large deletions, or small deletions. All were recorded at the cDNA/protein

levels and numbered using the reference transcript ENST00000361603. Nucleotide location & protein domains—signal peptide (bp 203–280), NC2 (281–322), collagenous (323–4570), and NC1 (4583–5257) were used to correlate positions, as identified by Ensembl, with phenotype. Gly-X-Y substitutions were examined separately.

Phenotype Information- The next step involved obtaining clinical histories between 2005 and 2007 via structured telephone interviews, with the questionnaires addressing various data from ESRD status, age at diagnosis, hypertension, proteinuria and hematuria, transplant history to ocular changes such as lens malformation, cataract, maculopathy, as well as hearing problems. This extensive US study investigated XLAS's progression and how genes link to its traits. Most (91%) had microscopic hematuria, and nearly half (49%) experienced visible hematuria. Proteinuria was present in 85%, with 54% having hypertension, and 60% developing ESRD, typically around 37 years of age. Ocular issues arose in 30%, while a notable 67% reported experiencing hearing loss [Jais JP et al., 2000]. The study revealed clear genotype-phenotype correlations, with missense mutations associated with the mildest phenotype and latest ESRD onset and truncating, splice-site, and deletion mutations causing more severe manifestations. Missense mutation types and positions were strong prognostic factors for XLAS when taking family correlation into account. Accordingly, this allows findings to reinforce genetic testing as an important prognostic and management tool in XLAS, suggesting that it has a predictive value even more than renal or skin biopsy at certain times.

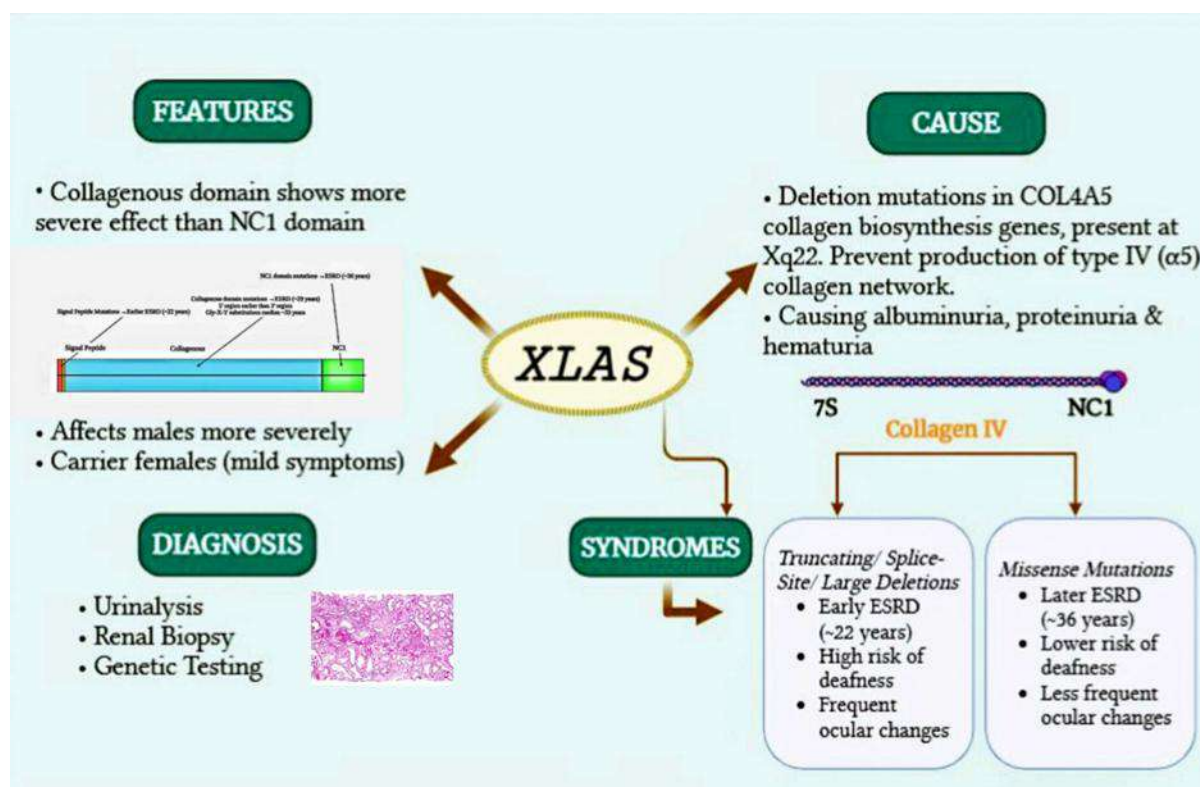


Figure 1: X- Linked Alport Syndrome

PCDH19- RELATED INFANTILE EPILEPTIC ENCEPHALOPATHY

Protocadherin (PCDH), is the largest segment of the cadherin superfamily, here the transmembrane proteins included in calcium-dependent cell adhesion, especially in the brain. They contain six or more EC domains necessary for cell–cell interactions [Morishita and Yagi, 2007] and play important roles in neurodevelopment, including synaptic plasticity and neuronal migration [Frank and Kemler, 2002]. PCDH19, a $\delta 2$ -subclass non-clustered PCDH located on chromosome Xq22.3, encodes 1148-amino-acid protein. Here the first exon breaks all 6 EC domains [Depienne C, 2012]. PCDH19 shows different, tightly controlled expression in the development of brain, particularly in the subventricular zone, intermediate zone, subplate, and cortex. $\delta 2$ -PCDHs aid in calcium-dependent adhesion and may provide to neuronal network formation [Biswas et al, 2010].

Pathogenic Mutation

Over 60 different kinds of mutations have been recognized in the PCDH19 gene, as well as nonsense, frameshift, splice site, missense mutations, and gene deletions. All over 50% of these mutations result in premature termination codons (PTCs), produced by nonsense mutations (18.3%), small insertions/deletions (25.8%) and splice site changes (3.2%). When mutations are

found throughout the gene, exon 2 remains genuine. Missense mutations, composing 46.2%, and this occupied to exon 1 and affect protected remainders in the extracellular domain. Gene deletions are less common (6.5%) but may be insufficiently. The Asn340Ser missense mutation is the most regularly detect variant [Depienne C, 2011].

Polymorphisms & Variants of Unknown Significance

The PCDH19 gene has a limited number of known polymorphisms. The most common natural variations are synonymous substitutions found in exon 1. Rare, nonpathogenic missense polymorphisms—concentrated in the intracellular domain of PCDH19 (exon 6) have been occasionally detected in patients as well as in ethnically matched controls. It indicates that the intracellular domain is capable of accommodating certain missense alterations, in distinctness to the extracellular domain. A missense variant located in exon 3, which was discovered in a patient with intellectual disability, while its pathogenic implications remain unclear [Tarpey et al., 2009].

Mutation in PCDH19

The mutation spectrum observed in PCDH19 is associated with a loss of function in the mutated allele. MRNAs containing mutations that lead to PTCs are prone to degradation through the nonsense-mediated mRNA decay surveillance system present in the fibroblasts of affected individuals. Missense mutations occurring in the extracellular domain may modify the adhesive characteristics of protocadherin 19, potentially leading to a loss of function. In particular, the Asn340Ser and Glu414Gln mutations could influence the amino acids that are crucial for calcium binding [Marini C, 2010; Patel SD, 2006].

Cellular Interference

Males possessing hemizygous PCDH19 mutations do not exhibit seizures or cognitive impairments, indicating that PCDH19 may not be essential in humans [Emond et al., 2009], as other pathways may compensate for its function. Conversely, females with heterozygous PCDH19 mutations experience early-onset seizures and varying degrees of intellectual disability, attributed to mosaicism induced by XCI [Dibbens et al., 2008]. This mosaicism results in a combination of normal and PCDH19-deficient cells, which interferes with cell–cell communication — a phenomenon referred to as "cellular interference" [Wieland et al., 2004]. The case of an affected mosaic male lends support to this hypothesis; however, validation necessitates data from homozygous females or relevant animal models.

Clinical Relevance

Females are possessing with heterozygous PCDH19 mutations that exhibit a diverse array of epileptic phenotypes with seizures commencing in infancy (12.9 months) and frequently provoked by fever (approximately 90% of cases). Seizures are resistant to medication during childhood but it may show improvement with advancing age. The clinical presentation may resemble that of Dravet syndrome (DS), although PCDH19 cases generally manifest later, exhibit less status epilepticus, and demonstrate more favorable long-term prognosis. Photosensitivity, prevalent in DS, is infrequent in PCDH19 cases [Depienne et al., 2009]. The intellectual outcomes vary widely: from normal (27.7%) to mild, moderate, or severe cognitive impairment [Depienne et al., 2011]. The severity of epilepsy does not serve as a predictor for intellectual outcomes.

Genetic Finding

After variant calling and filtering with eight rare variants were examined with seven classified as benign. A remarkable heterozygous variant PCDH19 c.706C>T on the X chromosome which was identified and it is associated with epilepsy. This variant has been described previously as de novo in epilepsy cases often linked to autism and variants at or closely the same codon which have also been implicated in disease [Specchio et al., 2011]. It was absent in maternal DNA and suggesting it may be either de novo or inherited from hemizygous father (without symptoms) possibly due to meiotic nondisjunction [Martin, R.H., 2005].

Genotype-Phenotype Correlation

The link between genotype and phenotype in PCDH19 mutations is weak. Clinical symptoms can vary greatly even among family members. As an example, monozygotic twins may have different symptoms to suggesting that nongenetic factors are especially XCI in females which play a role [Bolduc et al., 2008]. Skewed XCI may lessen the disease's severity by limiting cellular interference. In contrast, the complete skewing might result in asymptomatic carriers while balanced inactivation could worsen symptoms. However, a definitive correlation has not established a clear connection between XCI in blood cells and clinical severity [Marini et al., 2010]. It has not been established as blood does not accurately reflect brain tissue. Validation requires studies using brain tissue or mouse models.

Diagnostic Relevance

Identifying PCDH19 mutations confirms the diagnosis in females with infantile epilepsy and assess increasing risk. Molecular testing is especially important to distinguish from similar

epileptic syndromes. It is recommended for females with early-onset febrile seizures, seizure clusters, treatment-resistant epilepsy, or a family history of affected females. In cases suspected of DS, testing for PCDH19 mutations should include sequencing and deletion analysis, especially when SCN1A mutations are negative. Mutations are found in 5 to 37% of case. Testing in males is complicated by somatic mosaicism and limited detection from blood DNA [Wright and Burton, 2009]. Genetic counselling is challenging because of the gene's unusual inheritance pattern; an asymptomatic father can pass mutations to all his daughters. Prenatal testing, including fetal sex determination, could be offered due to the risk of cognitive impairment. De novo mutations have a low recurrence risk, but germline mosaicism cannot be ruled out [Dibbens et al., 2011].

Future Perspectives

The patient's epilepsy characterized by fever-sensitive, clustered and focal or tonic seizure matches typical PCDH19-related features, indicating the mutation as the likely cause rather than KS gene dosage [Elia et al., 1995]. It is believed that the underlying tissue involves tissue mosaicism which results in abnormal neuronal networks a key characteristic of PCDH19-related epilepsy. Interestingly similar which is called "cellular interference" mechanisms have been observed in EFNB1-related craniofrontonasal syndrome through mouse models [Depienne et al., 2009]. However, we still lack direct evidence for PCDH19 such as brain mosaicism or relevant animal models. The case of a male patient with a heterozygous PCDH19 mutation lends further support to the cellular interference hypothesis [Compagni et al., 2003]. While clinical presentations of PCDH19-related epilepsy are more clarified the precise mechanisms leading to seizures and intellectual disability are not clear. Experimental data from animal models, particularly zebrafish, indicate that *pcdh19* has an important function in early brain development with a specific role in the process of cell migration in the neural plate [Emond et al., 2009]. PCDH19 probably acts in concert with other cadherins when the brain forms [Biswas et al., 2010]. To date only PCDH19 and PCDH15 have been associated with Mendelian disorders, though other protocadherin could potentially play a role in neurodevelopmental diseases such as autism and schizophrenia [Bray et al., 2002].

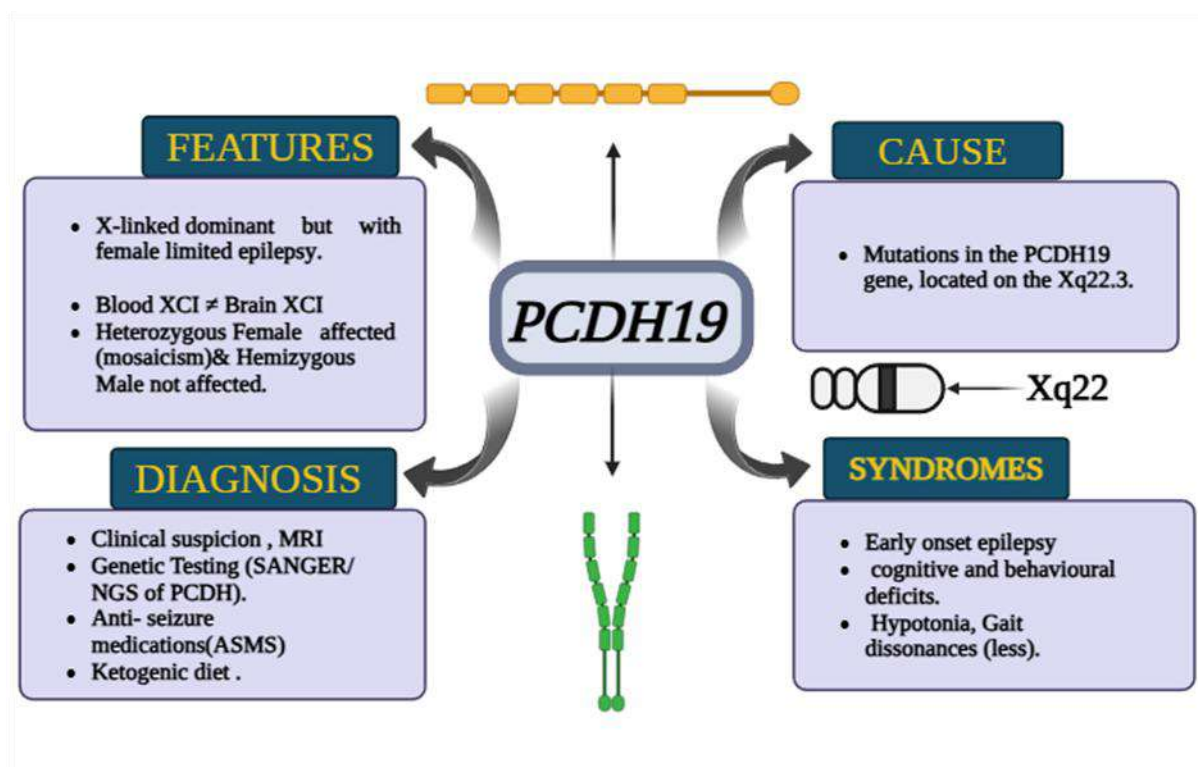


Figure 2: Protocadherin19- Related Infantile Epileptic Encephalopathy

RETT SYNDROME

Rett syndrome (RTT) is an X-linked neurological disorder that mainly affects the girls, and it's a rare one. In the year 1966, this disorder was first described by Andreas Rett. This disorder shows early development. RTT has four clinical stages: stagnation (6-18 months), rapid regression (1-4 years), pseudo stationary (2- potentially life), and late motor deterioration (10- life time). Main symptoms include loss of ability to speak and motor skills, hand-wringing is repetitive, seizures, irregularities in breathing, and gastrointestinal problems [Hagberg B. 2002]. Sometimes the patient faces smaller brain volumes due to reduction in size of the neurons, the dendrites do not grow properly, and imbalances in the neurotransmitter secretion [Armstrong DD, 1992]. Metabolic disturbances can be seen, such as dyslipidemia, dysfunction in mitochondria, oxidative stress gets increased, and abnormal energy metabolisms are common [Sierra C, 2001]. Despite of having these problems, many patients can be benefitted from communication aids. Management is symptomatic, focusing on nutrition, seizures, and orthopedic care. However, sudden death may occur due to cardiovascular and respiratory issues [Kerr AM, 1997].

Mutation in MeCP2

In Brazil a linkage analysis was done in a family; the causative agent is Xq28, and MECP2 gene mutations were found in most cases [Sirianni N, 1998]. MECP2 mutations are mostly de novo and caused RTT; the female is heterozygous [Neul JL, 2008]. RTT cases sometimes include mutations in the X-linked CDKL5 or FOXP1 gene [Sartori S, 2009]. MeCP2 is a nuclear protein that binds to methyl-DNA. In neurons, it is highly expressed and keeps increasing during the development of the brain, indicating that its role is in the maturation of neurons and synaptic function [Kishi N, 2005]. MECP2 produces two isoforms— e1, which is predominant mainly in the brain, and e2, with complex regulation through a long 3' UTR [Pelka GJ, 2005]. MeCP2 contains various functional regions: the methyl-binding domain, the transcriptional repression domain, and a chromatin-interacting C-terminal domain [Nan X, 1993]. It binds methylated CpG and non-CpG sequences, regulating expression of genes by promoting co-repressor complexes such as NCoR/SMRT and mSIN3A [Lyst MJ et al., 2013]. MeCP2 is an intrinsically disordered protein (IDP) that undergoes post-translational modifications (PTMs) [Adams VH, 2007], including phosphorylation and ubiquitination. Besides this, the MeCP2 gene also activates transcription, can perform splicing of mRNA, and acts as a versatile transcriptional modulator [Kokura K, 2001].

Statins that are used for cholesterol control, like Fluvastatin and lovastatin, are used to improve the function of motor skills, lifespan, and cholesterol balance in the brains of mice that are MeCP2 mutants, which benefits RTT [Jain MK, 2005]. Mutations in MeCP2 mainly occur in GABA neurons [Chao,H.T., 2010].

Clinical Overview

Hallmark features of RTT include improper hand use, stereotypic hand movements develop like wringing, clapping, decelerated head growth causing microcephaly, etc. Beside this, various features include abnormalities in breathing, changes in EEG, spasticity, scoliosis, and retardation of growth. Diagnosis depends on some criteria and is discarded if there is any proof of other metabolic, neurological, or structural disorders in the structure of brain. With an increase of the clinical studies, a larger phenotype has been seen, that includes slighter or more severe variation. Though it was thought that it's a disorder only for female but males with RTT symptoms have been found, generally in 47, XXY karyotype, mosaicism, or small mutation MECP condition [Schwartzman JS, 2001].

Management

RTT management need various several approach that mainly points on increasing abilities of the patients and thus providing support to family. Various treatment plans must involve, tailored education, physio social support and access to resources in the community. Medicinal diagnostics including L-carnitine, melatonin, and Mg may increase symptoms, still further studies through clinical trials are required. Difficulties in feeding can be seen due to problem in swallowing [Budden SS, 1997]. Nutritional care, involving gastrostomy and improved practices in feeding, can increase growth. Communication support like eye-pointing and boards help to express. Monitoring in EEG helps to differentiate original seizures. Prolonged QT involves to not take any medications that may arise cardiac arrhythmias [Ellaway CJ, 1999]. 65% of patients are affected by scoliosis and bracing. Early symptoms of motor inflexibility, like elevated Achilles tendon tone, need physiotherapy and orthoses to maintain mobility. Statins that are used for cholesterol control like fluvastatin and lovastatin are used that improve function of motor, lifespan, and cholesterol balance in the brain of mice that are MeCP2 mutant, which is benefitted for RTT.

Retardation involves epilepsy, and treatment should be biomarker-specific. Research on MECP2 was done and was found that it has its role in gene expression in neurons and formation of synapsis. MECP2 mutation destroy these functions. Variation in RTT Phenotype is due to several factor like genetic heterogeneity and X-inactivation. Beside this, mutations in CDKL5 are related to RTT with early-onset seizures. Further identification of MeCP2 and CDKL5 targets is important to understand RTT and development of new targeted therapies.

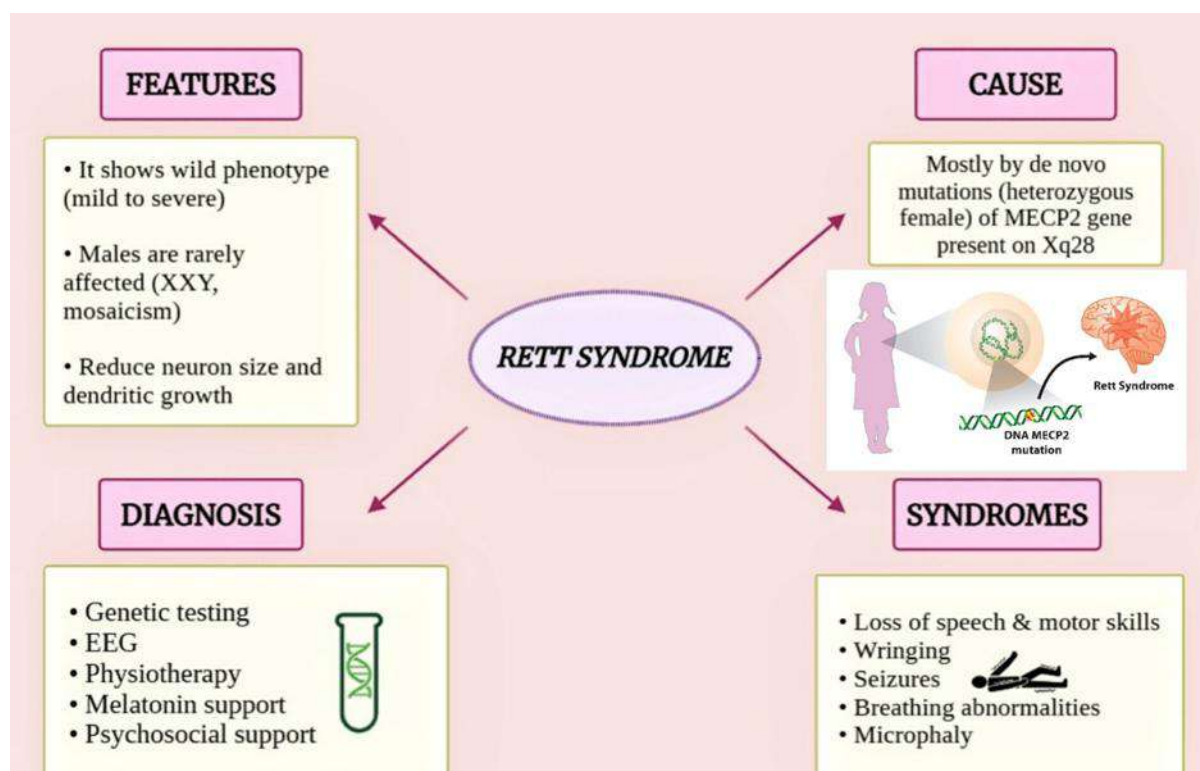


Figure 3: Rett Syndrome

FABRY DISEASE

X-linked inheritance, an aspect of Mendelian inheritance, is how Fabry Disease (FD) is passed down. Disorders that can be inherited as single-gene characteristics and whose inheritance seems to obey Mendel's laws are typically referred to as having "Mendelian inheritance" [Cassidy SB, 2005]. Early on, lysosomal glycosphingolipid deposition-induced microvascular pathology and dysfunction in cells play significant roles. Globotriaosylceramide (Gb3 or GL-3) [De Duve C, 1975] & related glycosphingolipids (galabiosylceramide) start to accumulate within lysosomes, known to be ubiquitous subcellular organelles, when lysosomal exoglycohydrolase α -gal A (α -D-galactoside galactohydrolase) is missing or inadequately active [Kint JA, 1970]. Both sexes experience progressive organ failure as an outcome of aging-related destruction of critical organ systems [Wilcox WR, 2008]. Patients who mainly have heart or kidney issues were later divided into "cardiac variant" [Elleder M, 1990] and "renal variant" [Nakao S, 2003]. Incorrectly, female heterozygotes were labeled as "carriers of the defective gene," which essentially shielded them from developing disease symptoms.

Epidemiology

More than 50 genetically unique lysosomal storage disorders, including FD, occur due to a monogenic imperfection. Despite being pan-ethnic, FD is uncommon, with reported cases ranging from 1 in 476,000 [Poorthuis BJ, 1999] to 1 in 117,000 [Meikle PJ, 1999], causing specific prevalence estimates difficult. Nevertheless, newborn testing initiatives have revealed prevalence rates that are higher than expected. In the country of Italy, about 1 in 3,100 men [Spada M, 2006] and in Taiwan, 1 in 1,500 are affected.

Recognition of X Linkage: Recessive & Dominant Inheritance

X-linked recessive inheritance has 3 characteristics. First, men are nearly the only ones affected by the disorder. Secondly, unaffected carrier females pass it on to their sons; half of the daughters are carriers, and half of the sons are affected. Third, daughters who must carry the trait can pass it to their grandsons, but affected males can't give it to their sons. Family tree info doesn't always match up, so we need to think about cases where the father isn't the biological parent. Also, X-linked dominant inheritance [Harper PS, 2004] doesn't include passing the trait from father to son. Although both sexes are impacted, women are affected more often. As affected females transfer equally to sons and daughters, affected men only transmit to daughters. Because heterozygotes show both normal and mutant gene products, women are typically less impacted than men. The one exemption is transmitting males with the fragile-X premutation [Sherman SL, 1985].

Clinical Overview

The first manifestations associated with other symptoms of the disease more often start in boys, [Zarate YA, 2008] and early neurological impairments are generally observed in the small fibers of the peripheral somatic [Dutsch M, 2002] as well as in the autonomous nervous system [Cable WJ, 1982]. Fabry crises or episodic crises that is extreme anguishing discomfort that begins in extremities and reflects in the rest of the limbs. Chronic pain that is accompanied by paresthesia in burning and feeling as in tingling sensation [Charrow J, 2009]. In kidney, the symptoms of proteinuria as well as high urinary Gb3 excretion are noted. Glomerular endothelial, mesangial, interstitial, and podocyte cells accumulating Gb3 concomitantly cause renal injury, leading to effacement of the foot processes. Cardiac involvement develops in 40 to 60% of patients and is characterized by angina, dyspnea, and LVH [Shah JS, 2005]. Concentric LVH & diastolic dysfunction are common in men. Approximately 40% of patients have RVH that preserves systolic function but has impaired diastolic function [Palecek T, 2008]. Stroke occurs more

frequently in people [Sims K, 2009]. Small vessel disease is the main cause of the cerebrovascular events with hyper perfusion, thrombus-formation contributing to FD [DE Graba T, 2000]. Loss of hearing, vertigo & tinnitus are signs of auditory problems. Respiratory involvement can range from coughing & dyspnea to complete airway blockage [Brown LK, 1997]. Furthermore, anemia, remodeling of blood vessels & intima-media thickening of arteries can be seen. In addition to being carriers, heterozygous females with FD often display a variety of symptoms as a result of XCI, which causes varying organ involvement. Pain, angiokeratoma, cardiac, renal, and cerebrovascular problems are among the possible symptoms [Lyon MF, 1961].

Etiology

The GLA gene indicates lysosomal α -gal A, found on the X chromosome at position Xq22. It covers 7 exons across 12,436 bps. In the GLA gene, different kinds of mutations are seen [Kornreich R, 1990]. More than 585 known mutations have been identified, the majority of which result in an enzyme that is not functional [Elstein D, 2010]. The variable activity of enzymes and progression of the disease can be understood by mutational divergence. While certain sequence modifications & polymorphisms of one nucleotide in the 5' untranslated region (such as p.Asp313Tyr in exon 6) [Froissart R, 2003] are still up for debate as they are benign variants or actual pathogenic mutations [Aerts JM, 2008].

Every single monomer of the homodimeric enzyme human α -gal A is made up of two domains: a C-terminal β -sandwich and an N-terminal (β/α)₈ barrel that occupies the site of action. Catalytic residues D170 (nucleophile) and D231 (acid/base) are among the 15 residues that make up the active site. Departed from the active site, 3 N-linked carbohydrates are linked together at N139, N192, and N215. The dimer's two active sites are spaced roughly 50 Å apart, and the structure is stabilized by a disulfide bond between C142 and C172 [Guce AI, 2010].

Diagnosis

Enzymatic Assay- Girls and adult women who are affected might have α -gal A activity levels in their plasma or leukocytes that appear normal [Linthorst GE, 2005]. Thus, using enzymatic assays isn't the best way to confirm the clinical diagnosis of FD in females. All females should have their status determined by genotyping, which identifies mutations in the GLA gene [Germain DP, 2007].

Genotyping- Nowadays testing based on DNA is the most efficient method for confirming FD. Point mutations can be detected by direct molecular analysis because of the tiny size of the GLA

gene [Germain DP, 2007]. In any case, if enzyme activity is found to be decreased without any obvious mutation being identified, MLPA would be the next most suitable option to detect deletions [Schirinzi A, 2008].

Histology- Light microscopy does not have great value in FD which presents with non-specific results, although lipid staining at times does bring to light storage cells. As for Electron microscopy (EM) is more so used for its diagnostic ability which includes the identification of whorled lysosomal inclusions, we term these ‘zebra bodies’ in heart and kidney biopsies. Also, because of its invasive nature and the fact that we have reliable molecular tools available, we reserve EM for when we have uncertain genetic results in females.

Biomarkers: Globotriaosylceramide- Urinary Gb3 serves as a dependable marker. Perhaps it takes time, but maximum patients can be diagnosed with this [Auray-Blais C, 2008]. Urinary Gb3 does not increase in certain patients with late-onset variants of FD or specific mutations in the GLA gene, for example, p. Asn215Ser [Piraud M, 2005]. Hemizygous males and adult females with the conventional FD have been found to have higher levels of Globotriaosylsphingosine or lysoGb3.

Enzyme Replacement Therapy (ERT)

In 2001, recombinant human α -gal A was used to initiate ERT for FD. There are two versions- agalsidase alfa & beta, approved in Europe [Sakuraba H, 2006], but they are made differently & have different doses. ERT's been proven safe and works well over time, but the rules for when to initiate treatment, specifically for women and kids, aren't the same everywhere in the world. Ongoing studies continue to examine whether similar efficacy can be achieved with alternative therapies, though it's clear that any real conclusions depend on appropriate analytical methods and well-matched baseline characteristics [Hoffmann B, 2007].

Future Perspectives

α - NAGAL enzymes engineered to copy α - gal A and are looking really good at targeting Fabry. This class of the enzyme seems to have some positive effects at suppressing Gb3 in animal and cell model-based assays & may be a safer alternative for ERT. Separately, some mutant enzymes are assisted in their folding by specific chaperones, which include 1-deoxygalactonojirimycin (DGJ) which also plays a role in proper cellular traffic of these enzymes. In the case of FD patients who have responsive GLA mutations, we see very good response from DGJ.

Because of skewed XCI, heterozygous ladies may also show signs and symptoms in X-connected disorders such as FD. Phenotypic variability results from the increased expression of the mutant

gene in specific tissues. The procedure is called "Lyonization", randomly deactivates one of the X chromosomes per cell, resulting in a mosaic of normal and mutant cells that affects symptom intensity and clinical appearance.

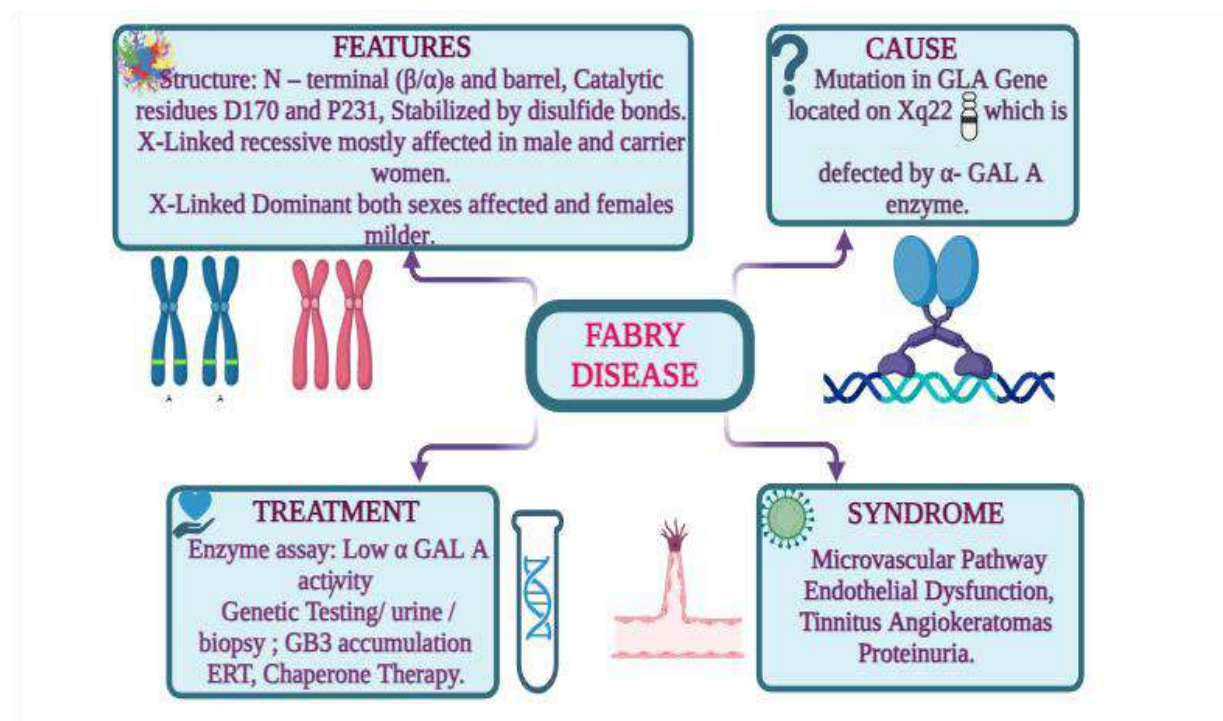


Figure 4: Febry Disease

Conclusion

To sum up X-linked diseases covers up various disorder that affect immunity, neural, and metabolic systems, often caused by specific pattern of inheritance and inactivation of X chromosome. Disorders like XLAS, PCDH19-related epilepsy, RTT, & FD show that X linked genes mutation are responsible for genotype and phenotype correlations that gets complex, sex-specific manifestations and variable expressivity. Due to single X chromosome males shows more severe phenotypes while females show various symptoms due to mosaicism, skewed XCI and genetic heterogeneity. Molecular diagnostics advancement involves sequencing of targeted gene and analysis of deletion have enhanced accuracy of diagnosis, genetic counselling and prognostication. Clinical management also shows diversity from ERT in FD to multidisciplinary care in RTT and control of seizure methods in PCDH19-related disorders. Research ongoing on various mechanisms like PCDH19 “cellular interference” and MECP2 metabolic role in RTT shows the complexity of pathophysiology of X-linked disease. However, innovation in therapies involving pharmacological chaperones, gene therapy & tailored interventions, is reshaping

various treatment in future. All over, these insights change the old view of X-linked disorders as simple Mendelian traits instead of picturing them as dynamic conditions caused by epigenetics, dosage of gene and systemic interactions. Genomic research integration, clinical data and functional studies are included that are important for precision medicine advancement approaches which provide outcomes for individuals affected by these disorders and often debilitating disorders.

References

- [1] Selmi C. Autoimmunity in 2010. *Autoimmune Rev* 2011; 10.
- [2] Invernizzi P, Gershwin ME. The genetics of human autoimmune disease. *J Autoimmune* 2009.
- [3] Gregory MC, Shamshirsaz AA, Kamgar M, Bekherinia MR: Alport syndrome, Fabry disease, and nail patella syndrome. In: *Diseases of the Kidney and Urinary Tract*, 8th Ed., edited by Schrier RW, Boston, Little, Brown and Company, 2007.
- [4] Dibbens et al., 2008; Fabisiak and Erickson, 1990; Juberg and Hellman, 1971; Ryan et al., 1997.
- [5] Depienne et al. 2011. Mutations and deletions in PCDH19 account for various familial or isolated epilepsies in females. *Hum Mutat* 32.
- [6] Burd L, Randall T, Martsolf JT, Kerbeshian J. 1991 Rett syndrome symptomatology of institutionalized adults with mental retardation: comparison of males and females. *Am. J. Ment. Retard.*
- [7] Stephanie M. Kyle^{1,2}, Neeti Vashi^{1,3} and Monica J. Justice^{1,2,3}.
- [8] Brady RO, Gal AE, Bradley RM, Martensson E, Warshaw AL, Laster L: Enzymatic defect in Fabry's disease: ceramide-trihexosidase deficiency. *N Engl J Med* 1967.
- [9] Sweeley CC, Klionsky B: Fabry's disease: classification as a sphingolipidosis and partial characterization of a novel glycolipid. *J Biol Chem* 1963.
- [10] Eaton WW, Rose NR, Kalaydjian A, Pedersen MG, Mortensen PB. Epidemiology of autoimmune diseases in Denmark. *J Autoimmune* 2007.
- [11] Ranke MB, Saenger P. Turner's syndrome. *Lancet* 2001
- [12] Guo X, Su B, Zhou Z, Sha J. Rapid evolution of mammalian X-linked testis microRNAs. *BMC Genomics* 2009.
- [13] Pinheiro I, Dejager L, Libert C. X-chromosome-located microRNAs in immunity: might they explain male/female differences? the X chromosome genomic context may affect X-located miRNAs and downstream signaling, thereby contributing to the enhanced immune response of females. *Bioassays* 2011
- [14] Flinter FA, Cameron JS, Chantler C, Houston I, Bobrow M. Genetics of classic Alport's syndrome. *Lancet* 1988; ii.
- [15] Barker DF, Hostikka SL, Zhou J et al. Tryggvason K. Identification of mutations in the COL4A5 collagen gene in Alport syndrome. *Science* 1990; 247.

- [16] Hudson BG, Reeder's ST, Tryggvason K. Type IV collagen: structure, gene organization and role in human diseases. *J Biol Chem* 268 (35): 1993.
- [17] Kashtan CE: Alport syndromes: Phenotypic heterogeneity of progressive hereditary nephritis. *Pediatr Nephrol* 14: 2000.
- [18] Gubler MC: Diagnosis of Alport syndrome without biopsy? *Pediatr Nephrol* 22: 2007.
- [19] Gross O, Netzer KO, Lambrecht R, Seibold S, Weber M: Meta-analysis of genotype-phenotype correlation in X-linked Alport syndrome: Impact on clinical counseling. *Nephrol Dial Transplant* 17: 2002.
- [20] Barker DF, Pruchno CJ, Xiang X, Atkin CL, Stone EM, Denison JC, Fain PR, Gregory MC: A mutation causing Alport syndrome with tardive hearing loss is common in the western United States. *Am J Hum Genet* 58:1996
- [21] Barker DF, Denison JC, Atkin CL, Gregory MC: Efficient detection of Alport syndrome COL4A5 mutations with multiplex genomic PCR SSCP. *Am J Med Genet* 98: 2001
- [22] Jais JP et al: X-linked Alport syndrome: Natural history in 195 families and genotype-phenotype correlations in males. *J Am Soc Nephrol* 11: 2000
- [23] Morishita H, Yagi T. 2007. Protocadherin family: diversity, structure, and function. *Curr Opin Cell Biol* 19.
- [24] Frank M, Kemler R. 2002. Protocadherin. *Curr Opin Cell Biol* 14.
- [25] Biswas S, Emond MR, Jontes JD. 2010. Protocadherin-19 and N-cadherin interact to control cell movements during anterior neurulation. *J Cell Biol* 191.
- [26] Tarpey PS, Smith R, Pleasance E, Whibley A, Edkins S, Hardy C, O'Meara S, Latimer C, Dicks E, Stratton MR. 2009. A systematic, large-scale resequencing screen of X-chromosome coding exons in mental retardation. *Nat Genet* 41.
- [27] Marini C, Mei D, Parmeggiani L, Norci V, Calado E, Ferrari A, Moreira A, Pisano T, Specchio N, Vigeveno F, Battaglia D, Guerrini R. 2010. Protocadherin 19 mutations in girls with infantile-onset epilepsy. *Neurology* 75.
- [28] Patel et al. 2006. Type II cadherin ectodomain structures: implications for classical cadherin specificity. *Cell* 124.
- [29] Emond MR, Biswas S, Jontes JD. 2009. Protocadherin-19 is essential for early steps in brain morphogenesis. *Dev Biol* 334.
- [30] Dibbens et al., 2008. X-linked protocadherin 19 mutations cause female-limited epilepsy and cognitive impairment. *Nat Genet* 40.
- [31] Wieland I, Jakubiczka S, Muschke P, Cohen M, Thiele H, Gerlach KL, Adams RH, Wieacker P. 2004. Mutations of the ephrin-B1 gene cause craniofrontonasal syndrome. *Am J Hum Genet* 74.
- [32] Depienne et al. Sporadic infantile epileptic encephalopathy caused by mutations in PCDH19 resembles Dravet syndrome but mainly affects females. *PLoS Genet* 5: e1000381.

- [33] Specchio N, Marini C, Terracciano A, Mei D, Trivisano M, Sicca F, Fusco L, Cusmai R, Darra F, Bernardina BD, Bertini E, Guerrini R, Vigeveno F. 2011. Spectrum of phenotypes in female patients with epilepsy due to protocadherin 19 mutations. *Epilepsia* 52.
- [34] Martin, R.H., 2005. Mechanisms of nondisjunction in human spermatogenesis. *Cytogenet. Genome Res.* 111, 245–249.
- [35] Bolduc V, Chagnon P, Provost S, Dube MP, Belisle C, Gingras M, Mollica L, Busque L. 2008. No evidence that skewing of X chromosome inactivation patterns is transmitted to offspring in humans. *J Clin Invest* 118.
- [36] Wright CF, Burton H. 2009. The use of cell-free fetal nucleic acids in maternal blood for non-invasive prenatal diagnosis. *Hum Reprod Update* 15.
- [37] Elia, M., Musumeci, S.A., Ferri, R., Scuderi, C., Del Gracco, S., Stefanini, M.C., 1995. Seizures in Klinefelter's syndrome: a clinical and EEG study of five patients. *Ital. J. Neurol. Sci.*
- [38] Compagni, A., Logan, M., Klein, R., Adams, R.H., 2003. Control of skeletal patterning by ephrinB1-EphB interactions. *Dev. Cell* 5.
- [39] Bray NJ, Kirov G, Owen RJ, Jacobsen NJ, Georgieva L, Williams HJ, Norton N, Spurlock G, Jones S, Zammit S, O'Donovan MC, Owen MJ. 2002. Screening the human protocadherin 8 (PCDH8) gene in schizophrenia. *Genes Brain Behav.*
- [40] Hagberg B. 2002 Clinical manifestations and stages of Rett syndrome. *Ment. Retard. Dev. Disabil. Res. Rev.* 8. (doi:10.1002/mrdd.10020)
- [41] Armstrong DD. 1992 The neuropathology of the Rett syndrome. *Brain Dev.*
- [42] Sierra C, Vilaseca MA, Brandi N, Artuch R, Mira A, Nieto M, Pineda M. 2001 Oxidative stress in Rett syndrome. *Brain Dev.* 23(Suppl 1). (doi:10.1016/S0387-7604(01)00369-2)
- [43] Kerr AM, Armstrong DD, Prescott RJ, Doyle D, Kearney DL. 1997 Rett syndrome: analysis of deaths in the British survey. *Eur. Child Adolesc. Psychiat.* 6(Suppl 1).
- [44] Sirianni N, Naidu S, Pereira J, Pillotto RF, Hoffman EP. 1998 Rett syndrome: confirmation of X-linked dominant inheritance, and localization of the gene to Xq28. *Am. J. Hum. Genet.* (doi:10.1086/302105)
- [45] Neul JL, Fang P, Barrish J, Lane J, Caeg EB, Smith EO, Zoghbi H, Percy A, Glaze DG. 2008 Specific mutations in Methyl-CpG-Binding Protein 2 confer different severity in Rett syndrome. *Neurology.* (doi: 10.1212/01.wnl.0000291011. 54508.aa)
- [46] Sartori S, Di Rosa G, Polli R, Bettella E, Tricomi G, Tortorella G, Murgia A. 2009 A novel CDKL5 mutation in a 47, XXY boy with the early-onset seizure variant of Rett syndrome. *Am. J. Med. Genet. Part A* 149. (doi:10.1002/ajmg.a.32606)
- [47] Kishi N, Macklis JD. 2005 Dissecting MECP2 function in the central nervous system. *J. Child Neurol.* 20. (doi:10.1177/08830738050200091001)

- [48] Pelka GJ, Watson CM, Christodoulou J, Tam PPL. 2005 Distinct expression profiles of Mecp2 transcripts with different lengths of 3'UTR in the brain and visceral organs during mouse development. *Genomics*. (doi:10.1016/j.ygeno.2004.12.002)
- [49] Nan X, Meehan RR, Bird A. 1993 Dissection of the methyl-CpG binding domain from the chromosomal protein MeCP2. *Nucleic Acids Res*. (doi:10.1093/nar/21.21.4886)
- [50] Lyst MJ et al. 2013 Rett syndrome mutations abolish the interaction of MeCP2 with the NCoR/SMRT co-repressor. *Nat. Neurosci*. 16. (doi:10.1038/nn.3434)
- [51] Adams VH, McBryant SJ, Wade PA, Woodcock CL, Hansen JC. 2007 Intrinsic disorder and autonomous domain function in the multifunctional nuclear protein, MeCP2. *J. Biol. Chem*. (doi:10.1074/jbc.M700855200)
- [52] Kokura K, Kaul SC, Wadhwa R, Nomura T, Khan MM, Shinagawa T, Yasukawa T, Colmenares C, Ishii S. 2001 The Ski protein family is required for MeCP2 mediated transcriptional repression. *J. Biol. Chem*. (doi:10.1074/jbc.M105747200)
- [53] Jain MK, Ridker PM. 2005 Anti-inflammatory effects of statins: clinical evidence and basic mechanisms. *Nat. Rev. Drug Discov*. (doi:10.1038/nrd1901)
- [54] Chao, H.T.; Chen, H.; Samaco, R.C.; Xue, M.; Chahrour, M.; Yoo, J.; Gong, S.; Lu, H.-C.; Heintz, N.; Ekker, M.; et al. Dysfunction in GABA signaling mediates autism-like stereotypies and Rett syndrome phenotypes. *Nature* 2010.
- [55] Schwartzman JS, Bernardino A, Nishimura A, Gomes RR, Zatz M. Rett syndrome in a boy with a 47, XXY karyotype confirmed by a rare mutation in the MECP2 gene. *Neuropediatric* 2001.
- [56] Budden SS. Rett syndrome: habilitation and management reviewed. *Eur Child Adol Psych* 1997.
- [57] Ellaway CJ, Sholler G, Leonard H, Christodoulou J. Prolonged QT interval in Rett syndrome. *Arch Dis Child* 1999.
- [58] Cassidy SB, Allason JE. Management of genetic syndromes. Second edition ed., Hoboken, New Jersey: Wiley, 2005.
- [59] De Duve C: Exploring cells with a centrifuge. *Science* 1975.
- [60] Kint JA: The enzyme defect in Fabry's disease. *Nature* 1970.
- [61] Wilcox WR et al.: Females with Fabry disease frequently have major organ involvement: lessons from the Fabry Registry. *Mol Genet Metab* 2008
- [62] Elleder M, Bradova V, Smid F, Budesinsky M, Harzer K, Kustermann-Kuhn B, Ledvinova J, Belohlavek X, Kral V, Dorazilova V: Cardiocyte storage and hypertrophy as a sole manifestation of Fabry's disease. *Virchows Arch Pathol Anat Histopathol* 1990.
- [63] Nakao S, Kodama C, Takenaka T, Tanaka A, Yasumoto Y, Yoshida A, Kanzaki T, Enriquez AL, Eng CM, Tanaka H, Tei C, Desnick RJ: Fabry disease: detection of undiagnosed hemodialysis patients and identification of a "renal variant" phenotype. *Kidney Int* 2003.
- [64] Poorthuis BJ, Wevers RA, Kleijer WJ, Groener JE, de Jong JG, van Weely S, Niezen-Koning KE, van Diggelen OP: The frequency of lysosomal storage diseases in The Netherlands. *Hum Genet* 1999.

- [65] Meikle PJ, Hopwood JJ, Clague AE, Carrey WF: Prevalence of lysosomal storage disorders. *JAMA* 1999.
- [66] Spada M, Pagliardini S, Yasuda M, Tukul T, Thiagarajan G, Sakuraba H, Ponzzone A, Desnick RJ: High incidence of later-onset Fabry disease revealed by newborn screening. *Am J Hum Genet* 2006.
- [67] Harper PS. Practical genetic counselling. 6th ed. 2004, London: Arnold.
- [68] Sherman SL, Jacobs PA, Morton NE, Froster-Iskenius U, Howard-Peebles PN, Nielsen KB, et al. Further segregation analysis of the fragile X syndrome with special reference to transmitting males. *Hum Genet* 1985.
- [69] Zarate YA, Hopkin RJ: Fabry's disease. *Lancet* 2008.
- [70] Cable WJ, Kolodny EH, Adams RD: Fabry disease: impaired autonomic function. *Neurology* 1982.
- [71] Charrow J: A 14-year-old boy with pain in hands and feet. *Pediatr Ann* 2009.
- [72] Shah JS, Hughes DA, Sachdev B, Tome M, Ward D, Lee P, Mehta AB, Elliott PM: Prevalence and clinical significance of cardiac arrhythmia in Anderson-Fabry disease. *Am J Cardiol* 2005.
- [73] Palecek T, Dostalova G, Kuchynka P, Karetova D, Bultas J, Elleder M, Linhart A: Right ventricular involvement in Fabry disease. *J Am Soc Echocardiogr* 2008.
- [74] Sims K, Politei J, Banikazemi M, Lee P: Stroke in Fabry disease frequently occurs before diagnosis and in the absence of other clinical events: natural history data from the Fabry Registry. *Stroke* 2009.
- [75] DE Graba T, Azhar S, Dignat-George F, Brown E, Boutiere B, Altarescu G, McCarron R, Schiffmann R: Profile of endothelial and leukocyte activation in Fabry patients. *Ann Neurol* 2000.
- [76] Brown LK, Miller A, Bhuptani A, Sloane MF, Zimmerman MI, Schilero G, Eng CM, Desnick RJ: Pulmonary involvement in Fabry disease. *Am J Respir Crit Care Med* 1997.
- [77] Lyon MF: Gene action in the X-chromosome of the mouse (*mus musculus* L.). *Nature* 1961.
- [78] Kornreich R, Bishop DF, Desnick RJ: α -Galactosidase A gene rearrangement causing Fabry disease. Identification of short direct repeats at breakpoints in an Alu-rich gene. *J Biol Chem* 1990.
- [79] Gal A: Molecular genetics of Fabry disease and Genotype-phenotype correlation. In *Fabry disease*. Edited by: Elstein D, Altarescu G, Beck M. Dordrecht, Heidelberg, London, New-York: Springer; 2010.
- [80] Froissart R, Guffon N, Vanier MT, Desnick RJ, Maire I: Fabry disease: D313Y is an α -galactosidase A sequence variant that causes pseudo deficient activity in plasma. *Mol Genet Metab* 2003.
- [81] Aerts JM, Groener JE, Kuiper S, Donker-Koopman WE, Strijland A, Ottenhoff R, van Roomen C, Mirzaian M, Wijburg FA, Linthorst GE, Vedder AC, Rombach SM, Cox-Brinkman J, Somer Harju P, Boot RG, Hollak CE, Brady RO, Poorthuis BJ: Elevated globotriaosylsphingosine is a hallmark of Fabry disease. *Proc Natl Acad Sci USA* 2008.
- [82] Guce AI, Garman SC: The structure of human α -galactosidase A and implications for Fabry disease. In *Fabry disease*. Edited by: Elstein D, Altarescu G, Beck M. Dordrecht, Heidelberg, London, New-York: Springer; 2010.
- [83] Linthorst GE, Vedder AC, Aerts JM, Hollak CE. Screening for Fabry disease using whole blood spots fails to identify one-third of female carriers. *Clin Chim Acta* 2005.

- [84] Germain DP, Shabbeer J, Cotigny S, Desnick RJ. Fabry disease: twenty novel alpha-galactosidase A mutations and genotype-phenotype correlations in classical and variant phenotypes. *Mol Med* 2002.
- [85] Schirinzi A, Centra M, Prattichizzo C, Gigante M, De Fabritiis M, Giancaspro V, et al. Identification of GLA gene deletions in Fabry patients by Multiplex Ligation-dependent Probe Amplification (MLPA). *Mol Genet Metab*; 2008.
- [86] Auray-Blais C, Cyr D, Ntwari A, West ML, Cox-Brinkman J, Bichet DG, et al. Urinary globotriaosylceramide excretion correlates with the genotype in children and adults with Fabry disease. *Mol Genet Metab* 2008.
- [87] Piraud M, Piraud M, de Goiffon F, Froissart R, Maire I, Vanier MT. [Globotriaosylceramide measurement in urine.]. *Med Sci (Paris)* 2005;21 (11 Suppl).
- [88] Sakuraba H, Murata-Ohsawa M, Kawashima I, Tajima Y, Kotani M, Ohshima T, Chiba Y, Takashiba M, Jigami Y, Fukushige T, Kanzaki T, Itoh K: Comparison of the effects of agalsidase alfa and agalsidase beta on cultured human Fabry fibroblasts and Fabry mice. *J Hum Genet* 2006.
- [89] Hoffmann B, Schwarz M, Mehta A, Keshav S: Gastrointestinal symptoms in 342 patients with Fabry disease: prevalence and response to enzyme replacement therapy. *Clin Gastroenterol Hepatol* 2007.

Chapter 3:

Molecular Basis of Phytochemical Protection Against NSAID-Induced Organ Toxicity in Mammals: A Contemporary Review

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Abstract

Non-steroidal anti-inflammatory drugs (NSAIDs) represent extensively prescribed therapeutic agents valued for their analgesic and antipyretic properties. These compounds exert their effects by inhibiting cyclooxygenase (COX) enzymes, thereby suppressing prostaglandin synthesis- a critical mediator of inflammatory responses in mammals. However, prolonged NSAID administration induces severe pathophysiological complications across multiple organ systems, including gastrointestinal mucosal damage, nephrotoxicity, cardiotoxicity, and hepatotoxicity. Conventional strategies to mitigate these adverse effects have yielded limited success. Emerging evidence demonstrates the therapeutic potential of phytochemicals as protective agents against NSAID-induced organ damage. These plant-derived bioactive compounds exhibit pleiotropic protective mechanisms through their antioxidant, anti-inflammatory, immunomodulatory, and anti-apoptotic properties. Recent investigations reveal that phytochemicals such as curcumin, quercetin, and resveratrol confer gastroprotective, cardioprotective, and hepatoprotective effects primarily by attenuating oxidative stress and suppressing pro-inflammatory mediators. The natural origin, dietary accessibility, and sustainability of phytochemical sources position them as promising candidates for therapeutic development. Advanced delivery systems, including nano formulations of curcumin and resveratrol, alongside combination therapies employing multiple phytochemicals, represent innovative strategies to enhance bioavailability and therapeutic efficacy. This review consolidates current understanding of phytochemical-mediated protection against NSAID toxicity in mammalian tissues, examining molecular mechanisms and therapeutic applications. Understanding these protective pathways is essential for developing evidence-

based interventions that preserve NSAID therapeutic benefits while minimizing organ-specific toxicity across mammalian species.

Keywords: Anti-inflammatory, Antioxidant, Phytochemicals, Pathophysiology, NSAIDs.

1. INTRODUCTION:

Non-steroidal anti-inflammatory drugs (NSAIDs) form the most unremarkably used group of prescribed medicine and non-prescribed medicine therapies for pain fever and redness in world and animals (Rainsford, 2013). Despite broad non-subjective pertinence during degenerative or no use of NSAIDs many inauspicious/pathophysiologic consequences can occur, especially in the gastrointestinal renal, cardiovascular, or hepatic system (Wallace, 2008; Lanas and Chan, 2017) of rules. These personal effects are for the most part COX drug-addicted, due to the suppression of prostaglandin (Vane and Botting, 1998) establishment and through with aerobic/nitrosamine strain mitochondrial loser, and photograph to weave impairment. Gathering latest studies suggests that the long term use of NSAIDs also diminishes mucosal wholeness, disrupts redox homeostasis and amplifies inflammatory cascade; thus, bearing possibly sincere risk in mammalian healthcare (Mousavi et al., 2020; Ricciotti and FitzGerald, 2011). Phytochemicals secondary winding metabolites of meditative plants have attracted much attending and show equiprobable preserving roles in opposite of such inauspicious personal effects (González-Peña et al., 2021; Pandey et al., 2021).

Phytochemicals, bioactive compounds extracted from meditative plants have in the past decades have drawn big attending as potential modulators for drug connected toxicities (Calderón-Montaña et al., 2011; Panche et al., 2016). The phytochemicals like polyphenols, flavonoids, terpenoids, alkaloids and saponins have assorted pharmacologic properties such as antioxidant, anti-inflammatory, cytoprotective immunomodulatory (Cowan, 1999; Kumar and Pandey, 2013) etc. Their pack rat potency toward free radicals' pro-/anti-inflammatory mediators, shelter of mitochondrial role and ordinance of gene reflexion represent their potency for discussion of NSAID-evoked pathologic changes (Surh, 2003; Sultana and Anwar, 2008). It is interesting to quotation that plant compounds such as curcumin, quercetin, resveratrol and catechins have also evidenced to faded stomachic ulcer aerobic strain and nephrotoxicity which is related with NSAID therapy in mammals (Goel et al., 2001; Al-Rejaie et al., 2013; Ren et al., 2019).

Curcumin is a polyphenolic palmatifid found in genus *curcuma longa*. It showed personal effects that protect the put up in the liver. These personal effects occur through with mechanisms

that subdue oxidation (Goel et al., 2008; Hewlings and Kalman, 2017) and redness. Quercetin and resveratrol are flavonoids and stilbenes found in many places. They have noticeable abilities to dispatch free radicals. They also commute instigative cytokines. Because of this they subdue stomachic ulcer and kidney impairment from oxidation. This impairment caused by NSAIDs. From green tea, catechins also have this effect. Similarly, in, from milk thistle as well has this effect. Both have been reportable to amend how mitochondria work. They also lessen lipid peroxidation. This advance shows their potency for therapy (Surai, 2015; Federico et al., 2017).

The flow state of affairs emphasizes on the usable role of phytochemicals which would help in preventing the toxicities caused by NSAIDs. In the time of the growing need for safer remedial adjuvants, phytochemicals show forebode to dispatch the drawbacks of handed down NSAID discussion. All the same, issues like pharmacokinetic variableness ‘bioavailability and normalization of preparations derivable from plants carry on being major obstacles (Scalbert et al., 2005; Wink, 2015). In order to advancement phytochemical-based interventions in no subjective carry out a thoroughgoing judgement of their mechanical efficaciousness is undependable, in the use of plastic approaches in drug expression.

2. NSAIDs

NSAIDs (non-steroidal anti instigative drugs) are a type of medicament used for their pain pill (pain relieving), febrifuge [fever reducing) and anti-instigative personal effects. In broad, they can be classified according to chemic social system and COX enzyme specificity. Based on chemic social system they are segmented into salicylates, propionic acid derivatives (isobutylphenyl propionic acid, naprosyn or ansaid, oruvail) acetic acid derivatives [indocin and diclofenac] (oxicams or enolic acid derivatives] feldene and meloxicam and the fenamates (mefenamic acid] including some exclusive COX 2 inhibitors (coxibs) including celecoxib and etoricoxib. The other agents nimesulide and torodal go to this group. NSAIDs are classified based on their selectivity for enzymes as non-exclusive COX inhibitors (e.g., empirin, isobutylphenyl propionic acid diclofenac ‘indocin) advantageous COX2 inhibitors (e.g., meloxicam, nimesulide] and exclusive COX2 inhibitors (e.g., celecoxib ‘etoricoxib) (Vane & Botting, 1998; Rang et al., 2021; Grosser et al., 2017).

They are used for a motley of non-subjective purposes. They are unremarkably used to subdue pain, including worry toothache‘ and musculoskeletal pain as well as reducing fever and redness (Rang et al., 2021). They help to lower fever peculiarly the non-exclusive ones such as isobutylphenyl propionic acid and aspirin (Grosser et al., 2017). Due to their powerful anti-instigative effectuate, they are practical in degenerative disorders such as- arthritic arthritis,

degenerative joint disease, ankylosing spondylitis, gout, bursitis and tendinitis (Vane & Botting, 1998). Low-dose aspirin assumes an unusual role as an antiplatelet agent in the case of myocardial infarct and stroke. NSAIDs are often used as dysmenorrhea (catamenial cramps) drugs such as isobutylphenyl propionic acid and mefenamic acid, and they are delineated as being especially usable for that purpose (Zahradnik et al., 2010). For short-term direction of pain which let in post-operative pains. Chemicals like diclofenac and acetophenone are in most use.

3. NSAIDs-INDUCED PHYSIOLOGICAL COMPLICATION IN MAMMALS

Though NSAIDs are very actual to facilitate pain, fever and flush the use of NSAIDs in mammals causes respective physical problems by inhibiting COX, a key enzyme of the prostaglandin deductive reasoning. By suppression of COX1 which is creditworthy for modulating tutelar prostaglandins and results in minimized stomachic mucus secretions and less hydrogen carbonate product, NSAIDs can lead to stomachic duodenal ulcer and erosions and GI bleeding (Vonkeman and van de Laar, 2010). They negatively sham renal prostaglandins which in turn may spoil renal blood flow peculiarly in unhealthy or hypotensive animals resulting in acute renal failure and water keeping and in some cases local death in papilloma (Carey et al., 2018). Long term use of some NSAIDs can also cause hepatotoxicity through with dose-in relation to or individual mechanisms. Cardiovascular organization - exclusive COX2 inhibitors may pose a higher risk of developing general high blood pressure myocardial infarct and thrombotic events [due to disruption of the equilibrium between prostacyclin and thromboxane]. Furthermore, in mammals the NSAID is able to break up fruitful homeostasis acting as an inhibitor of ovulation, implantation, and prostaglandin. These complications instance the fine line betwixt curative efficaciousness and side personal effects of NSAIDs in human and vet medication alike.

3.1 Liver toxicity

Hepatotoxicity is a clinically remarkable disease though it is rare and causes inauspicious core of NSAIDs in mammalian variety. NSAID-correlated hepatotoxicity is by mostly two main mechanisms: dose-addicted (built in) and individual (occasional) interactions. The dose-addicted form is extraordinary and is mostly seen with supra-hyper-dosages or prolonged dosages which impregnate the hepatic detoxifying paths. This summons includes mitochondrial scathe, aerobic tenseness and plain hepatocyte wound that emerges in hepatocyte local death (Boelsterli, 2002).

The individual type is both clinically more appropriate and occasional. It can find even in the comportment of therapeutical doses in susceptible subjects as a resolution of genetical polymorphisms, unsusceptible reactions or neutered drug metabolic process. For exemplify

hepatotoxicity of diclofenac is well secure due to its metabolite coevals and unsusceptible reactions. Symptoms can range from upraised liver enzymes [ALT, AST, ALP] to acerbity, from cholestasis to hepatocellular local death and in wicked cases to fulminant hepatic failure (Aithal et al., 2004).

Unlike hepatotoxic responses to confirmed NSAID presidential term have been genuine in dogs, cats and horses in veterinarian medication, return of individual hepatocellular local death has been related with carprofen in doges specially in Labrador Retrievers. Horses may show biochemical certify of hepatocellular wound during long butazolidin therapy.

3.2 Inflammation

NSAID elicited inflaming is unremarkably self-contradictory, as these drugs are intentional to decrement pain, fever, and inflaming through and through a decrement of the natural action of cyclooxygenase (COX) enzymes with accompanying decrement in prostaglandin synthetic thinking. Yet with immoderate or iterative use the light handed equaliser betwixt protecting and provocative mediators may be noncontinuous. suppression of COX—1 by NSAIDs decreases the synthetic thinking of prostaglandins that help to sustain blood flow to the tissues and that protect the stomachal mucous membrane, causing aggravation ulcer and decentralized provocative reactions in intestines. In the kidneys, reduced prostaglandin purpose can lead to cut blood flow resulting in provocative renal injury (Harirforoosh and Jamali, 2013), in addition to predestined individuals may go through hypersensitivity or insusceptible motivated reactions to NSAIDs which can arouse general inflaming such as skin rashes, bronchospasms, or even hepatic inflammation (Kowalski et al., 2013). Despite their direct role as anti provocative drugs, NSAIDs can paradoxically move provocative responses decussate unlike tissues in the body.

3.3 Pro-apoptotic changes

In addition, from blocking COX enzymes, NSAIDs also leads to pro apoptotic alterations through and through COX-breakaway pathways. They step in with the latent of the mitochondrial tissue layer and advance the discharge of cytochrome-c which triggers caspases and causes apoptosis (Zhou, Liu, & Shapiro, 2005). To get over anti apoptotic proteins like BCL-2 and regulating pro-apoptotic members like Bak and Bax, NSAIDs commute the equaliser of the BCL-2 category proteins. in addition, they gain cell death signalling (Shiff & Rigas, 1999) by producing excited o variety and inducing endoplasmic second stomach punctuate. These modifications invoice for NSAIDs' degrading personal effects on variety meat such as the venter,

kidney and liver as well as their accomplishable anticancer personal effects by causing tumour cells to experience apoptosis (Piazuelo & Lanás, 2015).

3.4 Tissue damage

Tissue injury caused due to NSAIDs is coupled with varied mechanics and interferes with synthetic thinking of prostaglandin and toxic metabolites. suppression of COX1 weakens the epithelial roadblock and causes gastritis, peptic ulcers and gastrointestinal bleeding in the gastrointestinal tract by lowering preventive prostaglandins mucosal blood flow, and hydrogen carbonate secretion (Wallace, 2008). Because prostaglandins typically keep the kidneys perfused, NSAID elicited crushing may lead to vasoconstriction ischaemia, vascular local death and even continual kidney failure (Whelton, 1999) over time. doomed NSAIDs like diclofenac may add to hepatotoxicity by producing excited metabolites which activate aerobic punctuate mitochondrial dysfunction, and insusceptible mediate hurt. This might end in cholestatic legal injury or hepatocellular local death (Aithal, 2004). By upsetting the sense of equilibrium betwixt prostacyclin [vascular dilatation, anti-thrombotic] and thromboxane (vasoconstrictive pro-thrombotic) NSAIDs can also activate cardiovascular problems by enhancing the risk of thrombosis, myocardial infarct, ischemic stroke (Antman et al., 2005) and tissue paper degeneracy in varied organ systems gets worse at the cancellous level by a gain in aerobic punctuate mitochondrial legal injury and apoptosis (Harirforoosh, Asghar, & Jamali, 2013).

4. MOLECULAR PATHWAY OF NSAID INDUCED CELL DAMAGE

NSAIDs initiation cell scathe chiefly through a compounding of prostaglandin depletion mitochondrial dysfunction, and aerobic strain–unvoluntary apoptosis. At the cellular level suppression of cyclooxygenase enzymes (COX1 and COX2) lowers the deduction of prostaglandins, which commonly protect cells by regulating blood flow mucosal wholeness and mitochondrial constancy. Without prostaglandins, cells lose their defence team against acid aerobic radicals, and calcium surcharge. NSAID metabolic process inside cells generates excited oxidative stress variety (ROS) and electrophilic intermediates that flak proteins, lipids and DNA, initiating aerobic strain and lipid peroxidation of membranes. within mitochondria NSAIDs uncouple aerobic phosphorylation by disrupting the electron exaltation chain, leading to loss of mitochondrial tissue layer potentiality and opening of the permeability conversion pore [MPTP]. This causes expiration of cytochrome c into the cytosol which activates caspase-9 and caspase-3, driving the inherent apoptotic tract. At the same time mitochondrial dysfunction increases intracellular Ca^{2+} assemblage activating proteases and endonucleases that disgrace cytoskeletal

proteins and DNA. NSAIDs also upregulate pro—apoptotic proteins [Bax, Bak, and p53] while suppressing anti-apoptotic Bcl2, tipping the symmetricalness toward programmed cell death. In terrible cases overwhelming aerobic strain and ATP depletion lead to local death scarred by serum tissue layer bust and anarchical outflow of cellular table of contents (Liu et al., 2015; Bjarnason et al., 2018; Varga et al., 2020).

5. REMEDIAL MEASURES ADOPTED SO FAR

Therapeutic measures adoptive so far regarding NSAID [Non-Steroidal Anti rabble-rousing Drug] perniciousness have for the most part centred on minimizing unfavourable personal effects while maintaining curative benefits. One major coming is the ontogenesis of discriminating COX2 inhibitors (like celecoxib), which aim to thin out redness and pain without gravely impairing COX 1 mediate stomachal protective covering, thereby lowering the risk of stomachal ulcers. In objective practise co prescription drug of gastroprotective agents such as proton pump inhibitors (omeprazole, pantoprazole) or H2 sensory receptor blockers are a demotic scheme to preclude NSAID evoked stomachal and duodenal ulcer (Lanza et al., 2009). The use of prodrugs and azotic oxide-donating NSAIDs has also been explored to raise mucosal blood flow and thin out gastrointestinal harm (Wallace, 2008). For patients at high risk current or transdermal formulations are sometimes used to avoid general perniciousness. in addition, monitoring liver and kidney go during chronic NSAID therapy has turn into a key prophylactic device measuring stick. explore into phytochemicals and antioxidants (such as curcumin quercetin and resveratrol) has shown call in counteracting aerobic strain and mitochondrial dysfunction evoked by NSAIDs. last, patient of department of education, dose optimization, and dodging of chronic or excess NSAID use are well thought out necessary non-pharmacologic remedies to limit unfavourable outcomes (Sostres et al., 2013).

6. PHYTOCHEMICALS

Phytochemicals are the naturally occurring bioactive compounds found in plants that are not necessary nutrients but exert world-shaking health-promoting personal effects in human race and animals. They are secondary winding metabolites produced by plants to protect themselves against pathogens, environmental strain and herbivores, and when eaten they put up caring curative and prophylactic device roles against diverse diseases. Phytochemicals are loosely sorted into various categories based on their chemic anatomical structure and begotten body process including phenolics (flavonoids, phenoplast acids, tannins, lignans) alkaloids (morphia, berberine, quinine), terpenoids (carotenoids saponins steroids), organosulfur compounds

(glucosinolates, allyl sulfides), and glycosides. Each group exhibits clear pharmacologic actions such as antioxidant anti-rabble rousing anti-carcinogenic, hepatoprotective and cardioprotective properties. Due to these multifarious begotten activities, phytochemicals have gained hearty involvement in biomedical inquiry, specially as potentiality alternatives or reciprocal agents to counterfeit drugs like NSAIDs for reducing perniciousness and enhancing curative efficaciousness (Liu, 2004; Pandey and Rizvi, 2009; Shahidi and Ambigaipalan, 2015).

7. SOURCES OF PHYTOCHEMICALS

i) Silymarin	Milk thistle (<i>Silybum marianum</i>)
ii) Glycyrrhizin	Licorice root (<i>Glycyrrhiza glabra</i>)
iii) Capsaicin	Red chili peppers (<i>Capsicum annuum</i>)
iv) Berberine	Tree turmeric, <i>Berberis</i> species
v) Gingerols	Ginger (<i>Zingiber officinale</i>)
vi) Apigenin	Parsley, chamomile, celery
vii) Kaempferol	Broccoli, spinach, kale, tea
viii) Catechins	Cocoa, dark chocolate, tea
ix) EGCG (Epigallocatechin gallate)	Green tea (<i>Camellia sinensis</i>)
x) Resveratrol	Grapes, red wine, peanuts
xi) Quercetin	Onions, apples, berries, green tea
xii) Curcumin	Turmeric (<i>Curcuma longa</i>)

8. PROTECTIVE ROLE OF PHYTOCHEMICALS AGAINST DIVERSE PATHOPHYSIOLOGICAL CONDITIONS

Phytochemicals the by nature occurring bioactive compounds stage in plants, showing world-shaking caring roles against divers' pathophysiological conditions evoked by non-steroidal, anti-rabble-rousing drugs [NSAIDs]. Long-term or high dose NSAID governance is often related to with gastrointestinal ulcer hepatic perniciousness, renal damage, aerobic strain, and pro-apoptotic changes in mammalian tissues. Phytochemicals such as curcumin (from curcuma longa], quercetin (from onions and apples], resveratrol (from grapes and peanuts], silymarin (from milk thistle), epigallocatechin gallate (from green tea] and glycyrrhizin [from liquorice] act through and through triple mechanisms including inhibition of excited atomic number 8 variety, suppression of rabble rousing mediators sweetening of antioxidant enzymes stabilisation of cell membranes, and input of cytoprotective mucus secretion. These bioactive not only

preclude weave scathe at stomachal, hepatic, and renal levels but also baffle apoptosis and elevate cancellate haunt mechanisms, thereby reducing the unfavourable outcomes of NSAID photo. Their multi targeted actions make them promising curative adjuncts for minimizing drug evoked complications and high spot the grandness of dietetic phytochemicals in prophylactic device and reciprocal practice of medicine (Surh, 2003; Hussain et al., 2016; Rauf et al., 2018).

Target Organ / Condition	Key Phytochemicals	Protective Mechanisms
Gastrointestinal Tract (ulceration, mucosal erosion)	Curcumin, Quercetin, Capsaicin, Gingerols	Enhanced stomachal mucus and hydrogen carbonate secretion, thin out aerobic strain subdues COX 2 and pro rabble-rousing cytokines quicken mucosal healing.
Liver (hepatotoxicity, oxidative stress, apoptosis)	Silymarin, Resveratrol, Curcumin, Berberine	stabilize hepatocyte membranes raise glutathione and antioxidant enzymes, thin out lipid peroxidation subdue mitochondrial dysfunction preclude apoptosis
Kidney (nephrotoxicity, oxidative stress, inflammation)	Quercetin, EGCG, Catechins, Kaempferol	salvage excited oxygen atoms variety touch on renal antioxidant defence team, conquer rabble-rousing pathways, protect renal hollow cells from apoptosis.
Systemic Oxidative Stress and Inflammation	Resveratrol, Apigenin, Quercetin, Curcumin	Suppress NF- κ B and MAPK signalling, reduce cytokine release, improve mitochondrial function, enhance overall redox balance.
Apoptotic and Pro-inflammatory Pathways	Apigenin, Curcumin, Berberine, Quercetin	Inhibit caspase activation, downregulate COX-2, TNF- α , IL-1 β , IL-6, and modulate cell survival pathways.

9. EFFICACY OF PHYTOCHEMICALS AGAINST NSAIDS INDUCED SIDE EFFECTS

Phytochemicals show noteworthy efficaciousness in reducing NSAID elicited side effects through their power to inflect aerobic, seditious, and apoptotic pathways. NSAIDs, work by inhibiting cyclooxygenase [COX] enzymes abridge prostaglandin deductive reasoning which not only lowers flush but also disrupts shielding mechanisms in the gastrointestinal tract, liver and

kidneys and results in ulceration (Wallace, 2008; Bjarnason et al., 2018). This leads to accumulated aerobic accent, mitochondrial dysfunction and apoptosis. Phytochemicals such as curcumin, quercetin and resveratrol undermine these personal effects through and through denary mechanisms. Curcumin inhibits NF κ B and COX 2 signalling reducing pro seditious cytokine eject while enhancing antioxidant enzyme action thereby protecting against stomachal and hepatic combat injury. Quercetin acts as a free ultra magpie and modulates the Nrf2 nerve pathway strengthening endogenic antioxidant defences and preventing renal aerobic impairment. Resveratrol exerts anti apoptotic personal effects by regulating Bcl 2 sept proteins and stabilizing mitochondrial membranes thus reducing NSAID elicited cell death (Shaito et al., 2020). In addition, polyphenols advance mucosal healing by enhancing mucus product and epithelial regeneration, countering gastrointestinal ulcer. By targeting these interrelated molecular pathways, phytochemicals render multi-faceted guarding against the inauspicious physiologic complications of NSAIDs, highlighting their sanative likely as completing agents.

10. CONCLUSION

Non-steroidal anti-inflammatory drugs (NSAIDs) comprise one of the most oft used classes of sanative agents mainly quantitative for their pain pill, febrifuge and anti seditious properties. They are extensively unarbitrary for both acute and progressive conditions such as fever, musculoskeletal pain creaky arthritis degenerative arthritis, and other seditious disorders. Despite their obligatory role in nonclassical medication, NSAIDs pose noteworthy health risks due to their pharmacologic chemical mechanism of legal action. By inhibiting cyclooxygenase (COX) enzymes NSAIDs abridge the deductive reasoning of prostaglandins that intermediate flush and pain; even so this also unknowingly interferes with the physiologic shielding roles of prostaglandins in the gastrointestinal tract kidneys and liver. As issue, long term or high dose organization often leads to pathophysiological complications in mammals (Grosser et al., 2017).

The effects of NSAIDs draw out over denary organ systems. Gastrointestinal ulcer and bleeding are among the most average outcomes due to belittled stomachal mucus secretion and hydrogen carbonate product. Hepatic toxicity, defined by overhead liver enzymes, cholestasis hepatocellular local death and bitterness, is a noteworthy complicating in susceptible individuals (Björnsson, 2016). Renal impairment arises from lessened renal blood flow and aerobic accent while cardiovascular risks such as high blood pressure and thrombosis are connected peculiarly with discriminating COX 2 inhibitors (Fitzgerald, 2004; Solomon et al., 2005). At the cellular level, NSAIDs can kick up aerobic accent mitochondrial dysfunction and pro apoptotic changes,

leading to tissue paper impairment and flush. These prejudicious personal effects emphasize the fine counterpoise betwixt sanative gain and perniciousness in NSAID usage.

To treat these challenges, respectable tending has been orientated towards shielding strategies. The approaches let in the evolution of discriminating COX-2 inhibitors like celecoxib, which aim to render anti seditious personal effects while reducing gastrointestinal harm. notwithstanding, even these drugs carry risks, peculiarly cardiovascular. In this linguistic context phytochemicals are naturally occurring bioactive compounds from plants, have emerged as promising alternatives or connected agents. Substances such as curcumin (from curcuma domestica), resveratrol (from grapes) quercetin (from onions and apples), and epigallocatechin gallate (from green tea) tell powerful antioxidant anti-seditious, and cytoprotective properties (Aggarwal and Harikumar, 2009; Russo et al., 2014). These compounds undermine NSAID elicited aerobic accent influence seditious signalling pathways, protect mitochondrial unity, and abridge apoptotic cell death. Their broad accessibility and comparatively safe profile make them fetching candidates for reducing NSAID allied perniciousness in mammals. Incorporation of phytochemicals and nutraceuticals into handling regimens may boost render a interactive shielding effectuates, continuing inquiry into the molecular mechanisms of NSAID perniciousness and the shielding role of unplanted compounds holds anticipate for safer long-term direction of pain and flush.

11. FUTURE PERSPECTIVES

The upcoming view on the advantageous role of phytochemicals against NSAID evoked pathophysiological conditions in mammals is promising focusing on various key areas.

- i. Targeted Molecular Mechanisms:* Phytochemicals are being explored for their power to inflect circumstantial incendiary pathways, such as NF- κ B, MAPKs, STAT and Nrf2 signalling. These pathways are essential in degenerative inflaming and phytochemicals that can tempt them may offer sanative benefits in conditions like arthritis and neurodegenerative diseases.
- ii. Gut Microbiota Modulation:* the fundamental interaction betwixt NSAIDs and the gut microbiota is an emerging area of search. Phytochemicals may help touch on a sound microbiome equilibriize possibly alleviating NSAID evoked enteropathy and enhancing boilers suit gut health.
- iii. Synergistic Therapies:* Combining phytochemicals with button down NSAIDs could concentrate needed dosages, thereby minimizing side personal effects. For case, ascorbic acid

has been shown to raise the inhibiting outcome of acetylsalicylic acid on COX2 mediate prostaglandin E2 product, suggesting likely for compounding therapies.

- iv. ***Personalized Medicine:*** Advancements in pharmacogenomics may enable the ontogeny of personal anti-inflammatory treatments. By tailoring phytochemical based therapies to individualistic familial profiles efficaciousness can be maximized while minimizing harmful personal effects.
- v. ***Sustainable Drug Development:*** The green synthetic thinking of nanomaterials from phytochemicals offers a sustainable plan of attack to drug ontogeny. These bio-based nanomaterials can be used in biosensing and targeted drug bringing systems, enhancing the specificity and efficaciousness of treatments.

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REFERENCES

- Aggarwal, B.B. and Harikumar, K.B., 2009. Potential therapeutic effects of curcumin, the anti-inflammatory agent, against neurodegenerative, cardiovascular, pulmonary, metabolic, autoimmune and neoplastic diseases. *The international journal of biochemistry & cell biology*, 41(1), pp.40-59.
- Aithal, G.P., 2004. Diclofenac-induced liver injury: a paradigm of idiosyncratic drug toxicity. *Expert Opinion on Drug Safety*, 3(6), pp.519-523.
- Al-Rejaie, S.S., Aleisa, A.M., Sayed-Ahmed, M.M., Al-Shabanah, O.A., Abuohashish, H.M., Ahmed, M.M. and Hafez, M.M., 2013. Protective effect of rutin on the oxidative stress, inflammatory response, and apoptosis in acetaminophen-induced nephrotoxicity in rats. *Journal of Applied Toxicology*, 33(3), pp.209–217.
- Antman, E.M., Bennett, J.S., Daugherty, A., Furberg, C., Roberts, H. and Taubert, K.A., 2007. Use of nonsteroidal antiinflammatory drugs: an update for clinicians: a scientific statement from the American Heart Association. *Circulation*, 115(12), pp.1634-1642.
- Bjarnason, I., Scarpignato, C., Holmgren, E., Olszewski, M., Rainsford, K.D. and Lanas, A., 2018. Mechanisms of damage to the gastrointestinal tract from nonsteroidal anti-inflammatory drugs. *Gastroenterology*, 154(3), pp.500-514.
- Björnsson, E.S., 2016. Hepatotoxicity by drugs: the most common implicated agents. *International journal of molecular sciences*, 17(2), p.224.
- Boelsterli, U.A., 2002. Mechanisms of NSAID-induced hepatotoxicity: focus on nimesulide. *Drug safety*, 25(9), pp.633-648.
- Calderón-Montaña, J.M., Burgos-Morón, E., Pérez-Guerrero, C. and López-Lázaro, M., 2011. A review on the dietary flavonoid kaempferol. *Mini-Reviews in Medicinal Chemistry*, 11(4), pp.298–344.
- Carey, R.M., Calhoun, D.A., Bakris, G.L., Brook, R.D., Daugherty, S.L., Dennison-Himmelfarb, C.R., Egan, B.M., Flack, J.M., Gidding, S.S., Judd, E. and Lackland, D.T., 2018. Resistant hypertension: detection, evaluation, and management: a scientific statement from the American Heart Association. *Hypertension*, 72(5), pp.e53-e90.
- Chu, L.Y., Liou, J.Y. and Wu, K.K., 2015. Prostacyclin protects vascular integrity via PPAR/14-3-3 pathway. *Prostaglandins & other lipid mediators*, 118, pp.19-27.
- Contos, M.J., Choudhury, J., Mills, A.S. and Sanyal, A.J., 2004. The histologic spectrum of nonalcoholic fatty liver disease. *Clinics in liver disease*, 8(3), pp.481-500.
- Cowan, M.M., 1999. Plant products as antimicrobial agents. *Clinical Microbiology Reviews*, 12(4), pp.564–582.
- Federico, A., Dallio, M. and Loguercio, C., 2017. Silymarin/silybin and chronic liver disease: a marriage of many years. *Molecules*, 22(2), p.191.
- FitzGerald, G.A., 2004. Coxibs and cardiovascular disease. *New England Journal of Medicine*, 351(17), pp.1709-1711.
- Goel, A., Kunnumakkara, A.B. and Aggarwal, B.B., 2008. Curcumin as “Curecumin”: from kitchen to clinic. *Biochemical Pharmacology*, 75(4), pp.787–809.
- Goel, R.K., Sairam, K. and Dorababu, M., 2001. Mechanism of anti-ulcerogenic effect of plant flavonoids: role of prostaglandins and nitric oxide. *Phytotherapy Research*, 15(2), pp.101–106.
- González-Peña, D., León-González, A.J., Daddaoua, A., Zarzuelo, A. and Gálvez, J., 2021. Role of polyphenols in the modulation of oxidative stress and inflammation induced by NSAIDs. *Frontiers in Pharmacology*, 12, p.633273.
- Grosser, T., Smyth, E. and FitzGerald, G.A., 2011. Anti-inflammatory, antipyretic, and analgesic agents; pharmacotherapy of gout. *Goodman and Gilman's the pharmacological basis of therapeutics*, 12, pp.959-1004.

- Harirforoosh, S., Asghar, W. and Jamali, F., 2013. Adverse effects of nonsteroidal antiinflammatory drugs: an update of gastrointestinal, cardiovascular and renal complications. *Journal of Pharmacy & Pharmaceutical Sciences*, 16(5), pp.821-847.
- Harirforoosh, S., Asghar, W. and Jamali, F., 2013. Adverse effects of nonsteroidal antiinflammatory drugs: an update of gastrointestinal, cardiovascular and renal complications. *Journal of Pharmacy & Pharmaceutical Sciences*, 16(5), pp.821-847.
- Hewlings, S.J. and Kalman, D.S., 2017. Curcumin: a review of its effects on human health. *Foods*, 6(10), p.92.
- HP, Z., 2010. Nonsteroidal anti-inflammatory drugs and hormonal contraceptives for pain relief from dysmenorrhea: a review. *Contraception*, 81, pp.185-196.
- Hung, S.C., Deng, W.P., Yang, W.K., Liu, R.S., Lee, C.C., Su, T.C., Lin, R.J., Yang, D.M., Chang, C.W., Chen, W.H. and Wei, H.J., 2005. Mesenchymal stem cell targeting of microscopic tumors and tumor stroma development monitored by noninvasive in vivo positron emission tomography imaging. *Clinical cancer research*, 11(21), pp.7749-7756.
- Hussain, T., Tan, B., Yin, Y., Blachier, F., Tossou, M.C. and Rahu, N., 2016. Oxidative stress and inflammation: what polyphenols can do for us?. *Oxidative medicine and cellular longevity*, 2016(1), p.7432797.
- Kowalski, M.L., Asero, R., Baybek, S.E.V.İ.M., Blanca, M., Blanca-Lopez, N., Bochenek, G., Brockow, K., Campo, P., Celik, G.Ü.L.F.E.M., Cernadas, J. and Cortellini, G., 2013. Classification and practical approach to the diagnosis and management of hypersensitivity to nonsteroidal anti-inflammatory drugs. *Allergy*, 68(10), pp.1219-1232.
- Kumar, S. and Pandey, A.K., 2013. Chemistry and biological activities of flavonoids: an overview. *The Scientific World Journal*, 2013, p.162750.
- Lanas, Á. and Chan, F.K., 2017. Peptic ulcer disease. *The Lancet*, 390(10094), pp.613–624.
- Lanza, F.L., Chan, F.K., Quigley, E.M. and Practice Parameters Committee of the American College of Gastroenterology, 2009. Guidelines for prevention of NSAID-related ulcer complications. *Official journal of the American College of Gastroenterology| ACG*, 104(3), pp.728-738.
- Liu, R.H., 2004. Potential synergy of phytochemicals in cancer prevention: mechanism of action. *The Journal of nutrition*, 134(12), pp.3479S-3485S.
- Mousavi, S.H., Baharara, J. and Asghari, M.H., 2020. The protective role of antioxidants in NSAID-induced gastrointestinal disorders. *Current Drug Metabolism*, 21(4), pp.289–299.
- Panche, A.N., Diwan, A.D. and Chandra, S.R., 2016. Flavonoids: an overview. *Journal of Nutritional Science*, 5, e47.
- Pandey, K.B. and Rizvi, S.I., 2009. Plant polyphenols as dietary antioxidants in human health and disease. *Oxidative medicine and cellular longevity*, 2(5), pp.270-278.
- Pandey, K.B., Rizvi, S.I. and Mishra, N., 2021. Plant polyphenols in the prevention of NSAID-associated gastrointestinal complications. *Phytotherapy Research*, 35(4), pp.2001–2013.
- Rainsford, K.D., 2013. *Anti-inflammatory and anti-rheumatic drugs*. Springer Science & Business Media.
- Rang, H.P., Dale, M.M., Ritter, J.M., Flower, R.J. and Henderson, G., 2011. *Rang & Dale's pharmacology*. Elsevier Health Sciences.
- Rauf, A., Imran, M., Suleria, H.A.R., Ahmad, B., Peters, D.G. and Mubarak, M.S., 2017. A comprehensive review of the health perspectives of resveratrol. *Food & function*, 8(12), pp.4284-4305.
- Reed, J.R., dela Cruz, A.L.N., Lomnicki, S.M. and Backes, W.L., 2015. Inhibition of cytochrome P450 2B4 by environmentally persistent free radical-containing particulate matter. *Biochemical pharmacology*, 95(2), pp.126-132.

- Ren, J., Lu, Y., Qian, Y., Chen, B., Wu, T., Ji, G. and Shi, L., 2019. Curcumin attenuates NSAID-induced gastric damage by regulating oxidative stress and inflammation. *Journal of Food Biochemistry*, 43(4), p.e12759.
- Ricciotti, E. and FitzGerald, G.A., 2011. Prostaglandins and inflammation. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 31(5), pp.986–1000.
- Russo, M., Spagnuolo, C., Tedesco, I., Bilotto, S. and Russo, G.L., 2012. The flavonoid quercetin in disease prevention and therapy: facts and fancies. *Biochemical pharmacology*, 83(1), pp.6-15.
- Scalbert, A., Johnson, I.T. and Saltmarsh, M., 2005. Polyphenols: antioxidants and beyond. *The American Journal of Clinical Nutrition*, 81(1), pp.215S–217S.
- Shahidi, F. and Ambigaipalan, P., 2015. Phenolics and polyphenolics in foods, beverages and spices: Antioxidant activity and health effects—A review. *Journal of functional foods*, 18, pp.820-897.
- Shaito, A., Posadino, A.M., Younes, N., Hasan, H., Halabi, S., Alhababi, D., Al-Mohannadi, A., Abdel-Rahman, W.M., Eid, A.H., Nasrallah, G.K. and Pintus, G., 2020. Potential adverse effects of resveratrol: A literature review. *International journal of molecular sciences*, 21(6), p.2084.
- Shiff, S.J. and Rigas, B., 1997. Nonsteroidal anti-inflammatory drugs and colorectal cancer: evolving concepts of their chemopreventive actions. *Gastroenterology*, 113(6), pp.1992-1998.
- Solomon, S.D., McMurray, J.J., Pfeffer, M.A., Wittes, J., Fowler, R., Finn, P., Anderson, W.F., Zauber, A., Hawk, E. and Bertagnolli, M., 2005. Cardiovascular risk associated with celecoxib in a clinical trial for colorectal adenoma prevention. *New England Journal of Medicine*, 352(11), pp.1071-1080.
- Sostres, C., Gargallo, C.J. and Lanas, A., 2013. Nonsteroidal anti-inflammatory drugs and upper and lower gastrointestinal mucosal damage. *Arthritis research & therapy*, 15(Suppl 3), p.S3.
- Sultana, S. and Anwar, F., 2008. Flavonols attenuate oxidative stress-mediated cytotoxicity induced by non-steroidal anti-inflammatory drugs. *Toxicology In Vitro*, 22(2), pp.491–499.
- Surai, P.F., 2015. Silymarin as a natural antioxidant: an overview of the current evidence and perspectives. *Antioxidants*, 4(1), pp.204–247.
- Surh, Y.J., 2003. Cancer chemoprevention with dietary phytochemicals. *Nature Reviews Cancer*, 3(10), pp.768–780.
- Vane, J.R. and Botting, R.M., 1997. Mechanism of action of anti-inflammatory drugs. *Recent advances in prostaglandin, thromboxane, and leukotriene research*, pp.131-138.
- Varga, Z.V., Ferdinandy, P., Liaudet, L. and Pacher, P., 2015. Drug-induced mitochondrial dysfunction and cardiotoxicity. *American Journal of Physiology-Heart and Circulatory Physiology*, 309(9), pp.H1453-H1467.
- Vonkeman, H.E. and van de Laar, M.A., 2010. Nonsteroidal anti-inflammatory drugs: adverse effects and their prevention. In *Seminars in arthritis and rheumatism* (Vol. 39, No. 4, pp. 294-312). WB Saunders.
- Wallace, J.L., 2008. Prostaglandins, NSAIDs, and gastric mucosal protection: why doesn't the stomach digest itself?. *Physiological Reviews*, 88(4), pp.1547–1565.
- Whelton, A., 1999. Nephrotoxicity of nonsteroidal anti-inflammatory drugs: physiologic foundations and clinical implications. *The American journal of medicine*, 106(5), pp.13S-24S.
- Wink, M., 2015. Modes of action of herbal medicines and plant secondary metabolites. *Medicines*, 2(3), pp.251–286.

Chapter 4:

3D Cell Culture Systems: Advances and Applications in Biomedical Research

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Abstract

Three-dimensional cell culture systems have emerged as transformative technologies bridging the gap between traditional two-dimensional monolayer cultures and complex in vivo tissue environments. Unlike conventional flat cultures where cells grow on rigid plastic surfaces under non-physiological conditions, 3D systems enable cells to organize into architectures that recapitulate native tissue structures, cell-cell interactions, extracellular matrix relationships, and microenvironmental gradients of oxygen, nutrients, and signaling molecules. This dimensional complexity profoundly influences cellular behavior including gene expression, protein synthesis, metabolism, differentiation, and drug responses, making 3D cultures more physiologically relevant models for biomedical research. Multiple 3D culture platforms have been developed, each with distinct advantages and applications. Scaffold-based systems including hydrogels and decellularized matrices provide structural support mimicking extracellular matrix. Scaffold-free approaches including spheroids and organoids rely on cells' intrinsic self-assembly capabilities to form complex structures. Microfluidic organs-on-chips integrate 3D cultures with controlled fluid flow, mechanical forces, and multi-tissue interfaces. These diverse platforms are revolutionizing drug discovery by providing predictive models for efficacy and toxicity screening, advancing cancer research through tumor spheroids and patient-derived organoids that preserve intratumoral heterogeneity, enabling regenerative medicine applications including tissue engineering and personalized therapeutics, and facilitating disease modeling where patient-derived cells recreate pathological phenotypes in vitro. This review examines the major 3D culture technologies, their biophysical and biochemical properties, comparative advantages and limitations, and applications across biomedical research domains. We discuss technical challenges including standardization, scalability, analytical characterization, and cost that must be addressed for widespread adoption. Emerging innovations including bioprinting, advanced biomaterials, integration with biosensors, and combination with genetic engineering are explored. As 3D culture technologies mature and become more accessible, they promise to accelerate therapeutic development, reduce animal experimentation, and enable precision medicine approaches tailored to individual patient biology.

Keywords: 3D cell culture, organoids, spheroids, tissue engineering, organs-on-chips, hydrogels, tumor models, drug screening, regenerative medicine

1. Introduction

Cell culture has served as a cornerstone of biological research and biotechnology for over a century, enabling investigation of cellular and molecular processes in controlled environments isolated from organismal complexity. Traditional two-dimensional (2D) monolayer culture, where cells adhere to flat plastic or glass surfaces, has yielded fundamental insights into cell biology and facilitated development of biologics, vaccines, and therapeutic screening assays (Duval et al., 2017). However, the artificial nature of 2D culture—cells flattened against rigid substrates, lacking tissue architecture, exposed uniformly to media components, and experiencing non-physiological mechanical forces—creates profound differences from *in vivo* environments that limit translational relevance.

The recognition that dimensionality profoundly influences cellular phenotype has driven development of three-dimensional (3D) culture systems that better recapitulate tissue environments. In native tissues, cells exist within complex 3D architectures, surrounded by extracellular matrix (ECM), engaged in multidirectional cell-cell interactions, and exposed to gradients of oxygen, nutrients, growth factors, and waste products (Jensen & Teng, 2020). These spatial and microenvironmental features regulate gene expression, signal transduction, metabolism, and functional differentiation. When cultured in 3D, cells exhibit morphologies, proliferation rates, differentiation patterns, and drug responses that more closely resemble *in vivo* behavior compared to 2D counterparts.

Multiple 3D culture platforms have emerged, each designed to recapitulate specific aspects of tissue organization. Scaffold-based systems employ natural or synthetic materials providing structural support and biochemical cues, including hydrogels (collagen, Matrigel, alginate, synthetic polymers) that encapsulate cells in ECM-mimicking environments, and decellularized tissues preserving native ECM architecture (Gunti et al., 2021). Scaffold-free approaches leverage cells' intrinsic self-assembly capabilities, generating spheroids through forced aggregation or organoids through directed differentiation of stem cells into miniature organ-like structures with multiple cell types and spatial organization. Microfluidic organs-on-chips integrate 3D cultures with perfusion systems, mechanical forces, and multiple tissue compartments, modeling organ-level physiology and inter-tissue communication (Ingber, 2016).

The applications of 3D culture span biomedical research domains. In drug discovery, 3D tumor models provide more predictive platforms for anticancer drug screening, better recapitulating chemoresistance mechanisms observed clinically. Patient-derived organoids enable personalized medicine by testing drug responses on individual patients' cells before treatment (Dutta et al., 2017). In regenerative medicine, 3D engineered tissues serve as grafts for repair or replacements, while organoids model development and disease mechanisms. Toxicology benefits from 3D liver and kidney models that better predict human toxicity. Cancer research employs tumor spheroids preserving intratumoral heterogeneity and microenvironmental features driving progression and therapeutic resistance.

This review provides comprehensive examination of 3D cell culture technologies, their biophysical principles, comparative characteristics, and biomedical applications. We discuss scaffold-based and scaffold-free systems, emerging organs-on-chip platforms, and specialized applications in cancer, regenerative medicine, and drug development. Technical challenges including reproducibility, characterization, and scalability are critically evaluated alongside innovations addressing these limitations. Our goal is to provide researchers with conceptual understanding necessary for selecting appropriate 3D systems for their applications while highlighting the transformative potential of these technologies for advancing biomedical science and translational medicine.

2. Scaffold-Based 3D Culture Systems

2.1 Hydrogel-Based Cultures

Hydrogels—crosslinked polymer networks swollen with water—provide tunable 3D environments mimicking ECM properties. Natural hydrogels including collagen, Matrigel (basement membrane extract), fibrin, and hyaluronic acid offer inherent bioactivity through integrin-binding motifs, protease-cleavable sites, and growth factor sequestration capabilities (Duval et al., 2017). Cells encapsulated within these materials experience 3D matrix architecture, enabling natural morphology and cell-matrix interactions absent in 2D culture. Matrigel, rich in laminin, collagen IV, and growth factors, supports organoid culture and maintains stem cell pluripotency, though batch-to-batch variability and undefined composition present challenges for standardization.

Synthetic hydrogels including polyethylene glycol (PEG), polyvinyl alcohol, and peptide-based materials offer defined composition and tunable mechanical and biochemical properties. Mechanical stiffness, controlled through polymer concentration and crosslinking density,

influences cell fate with soft matrices (0.1-1 kPa) promoting neural differentiation and stiff matrices (10-40 kPa) favoring osteogenic differentiation (Jensen & Teng, 2020). Biochemical functionalization through conjugated peptides (RGD for integrin binding, IKVAV for neural cells) provides specific cellular signals. Degradability through hydrolytic or enzymatic mechanisms allows cells to remodel their microenvironment, essential for invasion, morphogenesis, and tissue formation.

Table 1: Comparison of Major 3D Cell Culture Platforms

Platform Type	Method	Key Advantages	Main Limitations	Typical Applications	Cell Density	Cost
Hydrogel (Natural)	Cell encapsulation in ECM-like materials	Bioactive, cell-responsive, commercial availability	Batch variation, undefined composition, limited tunability	Organoids, stem cell culture, tissue engineering	$0.5-5 \times 10^6$ cells/mL	Moderate
Hydrogel (Synthetic)	Cell encapsulation in engineered polymers	Defined composition, tunable properties, reproducible	Lack bioactivity (requires functionalization), optimization needed	Mechanobiology, engineered tissues, controlled studies	$0.5-5 \times 10^6$ cells/mL	Moderate-High
Spheroids (hanging drop)	Gravity-driven aggregation	Simple, no materials, uniform size	Low throughput, manual, difficult media changes	Drug screening, stem cell aggregates	10^2-10^4 cells/spheroid	Low
Spheroids (ultra-low attachment)	Spontaneous aggregation on non-adhesive surfaces	High throughput, scalable, commercial plates	Size heterogeneity, aggregation time varies	High-throughput screening, tumor models	10^2-10^4 cells/spheroid	Low-Moderate
Organoids	Stem cell self-organization in Matrigel	Organ-like complexity, multiple cell types, physiological architecture	Complex protocols, expertise needed, variability	Disease modeling, drug testing, developmental biology	10^3-10^6 cells/organoid	High
Bioprinted Constructs	Layer-by-layer	Precise spatial control,	Equipment cost, limited materials,	Tissue engineering,	Variable	High

	deposition	complex architectures, customizable	specialized expertise	personalized medicine		
Organs-on-Chips	Microfluidic perfusion systems	Physiological flow, mechanical forces, multi-tissue interfaces	Complex fabrication, specialized equipment, scale-up challenges	Organ physiology, drug metabolism, toxicity	Variable	High

Note. ECM = extracellular matrix. Cost categories: Low (<\$100/experiment), Moderate (\$100-1,000), High (>\$1,000). Cell densities and costs are approximate and application-dependent.

2.2 Decellularized Matrices and Bioprinted Scaffolds

Decellularized tissues obtained by removing cellular components while preserving native ECM architecture provide scaffolds with organ-specific composition and structure. Perfusion of detergents, enzymes, or physical methods through tissues removes cells, leaving intact collagen fibers, basement membranes, and vascular networks (Gunti et al., 2021). Recellularization with relevant cell types creates functional tissue constructs for transplantation or in vitro models. Decellularized liver, heart, lung, and kidney scaffolds have been recellularized and shown to support cell survival, differentiation, and tissue-specific functions. However, complete decellularization without damaging ECM ultrastructure remains technically challenging, and immunogenic residual cellular components may trigger rejection.

Bioprinting technologies enable fabrication of 3D tissue constructs with precise spatial control over cell placement, biomaterial deposition, and architectural features. Extrusion bioprinting deposits cell-laden bioinks through nozzles, building structures layer-by-layer (Gunti et al., 2021). Inkjet bioprinting ejects droplets containing cells onto substrates, offering higher resolution but limited to low-viscosity materials. Laser-assisted bioprinting transfers cells from donor to receiver substrates with micrometer precision. Bioprinted constructs can recreate complex tissue features including vascular networks, cellular zonation, and multi-tissue interfaces impossible to achieve with conventional methods. Challenges include maintaining cell viability during printing, achieving adequate mechanical strength, and vascularizing large constructs for nutrient delivery.

3. Scaffold-Free Systems: Spheroids and Organoids

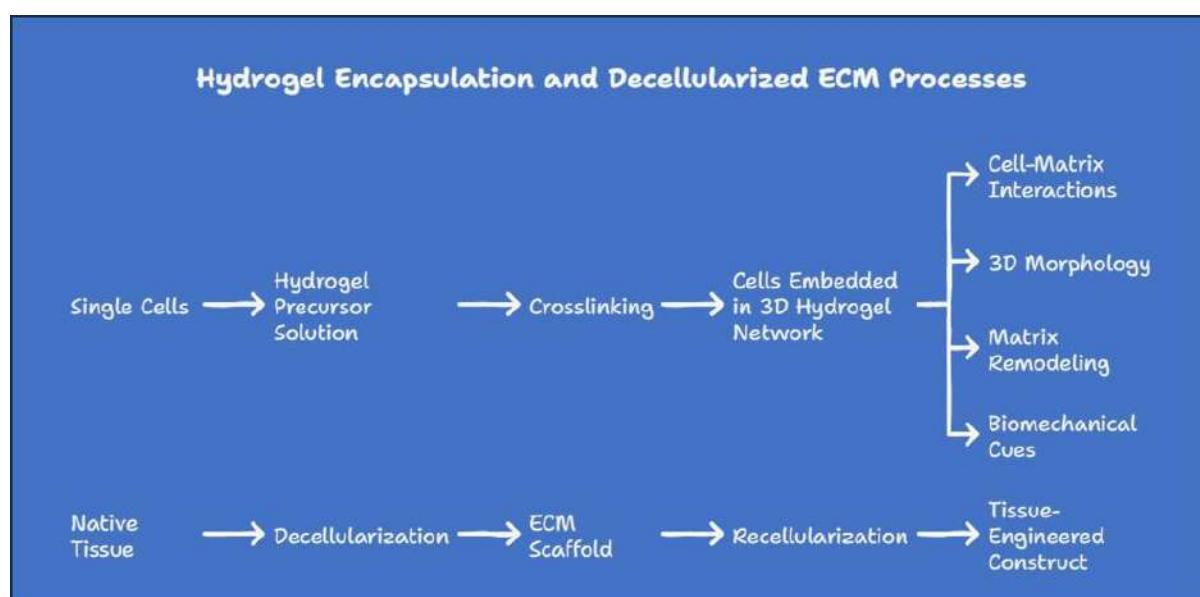
3.1 Tumor Spheroids and Multicellular Aggregates

Spheroids—three-dimensional aggregates of cells—form spontaneously when cells are prevented from attaching to substrates, instead interacting with each other to create compact structures. Multiple methods generate spheroids including hanging drop culture where gravity draws cells to droplet apex, ultra-low attachment plates with non-adhesive surfaces promoting aggregation, and spinner flask cultures where continuous agitation prevents surface attachment (Nunes et al., 2019). These scaffold-free approaches rely on cells' intrinsic adhesion molecules including cadherins to form cohesive structures.

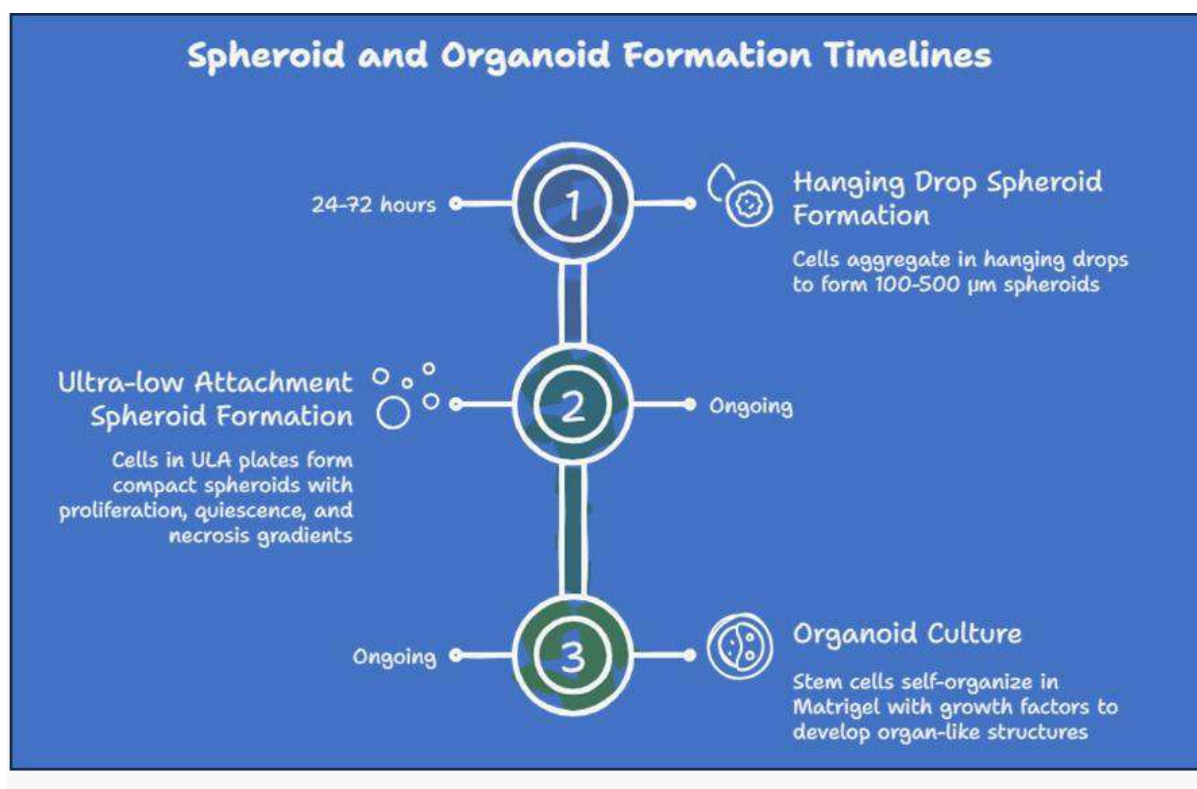
Tumor spheroids particularly have become widely adopted cancer models. Unlike 2D cancer cell monolayers where all cells proliferate rapidly and are equally exposed to drugs, spheroids develop gradients of oxygen, nutrients, and pH similar to avascular tumor microenvironments (Nunes et al., 2019). Outer proliferating cell layers surround quiescent intermediate zones and necrotic cores, recapitulating histological features of tumors. This heterogeneity influences drug responses, with chemotherapy often killing proliferating outer cells while sparing quiescent interior cells that subsequently repopulate tumors. Spheroids also develop drug resistance mechanisms including hypoxia-induced proteins, altered metabolism, and enhanced DNA repair absent in 2D cultures.

3D Cell Culture Technologies: Methods and Cellular Organization

A. SCAFFOLD-BASED SYSTEMS



B. SCAFFOLD-FREE SYSTEMS



C. STRUCTURAL COMPARISON

2D Culture:	3D Culture:
[Cells on flat surface]	[Cells in 3D environment]
- Uniform morphology	• Natural 3D morphology
- All cells proliferating	• Proliferation gradients
- Uniform drug exposure	• Diffusion gradients
- Altered gene expression	• In vivo-like expression
↓	↓
Limited physiological relevance	Enhanced physiological relevance

Note. Panel A shows scaffold-based approaches using hydrogels or decellularized ECM. Panel B illustrates scaffold-free spheroid and organoid formation methods. Panel C contrasts cellular organization and microenvironmental gradients between 2D and 3D cultures. ULA = ultra-low attachment; ECM = extracellular matrix.

Co-culture spheroids incorporating multiple cell types better model tumor complexity. Cancer cells co-cultured with fibroblasts, endothelial cells, and immune cells recapitulate tumor-stroma interactions driving invasion, angiogenesis, and immune evasion (Nunes et al., 2019). Patient-derived tumor spheroids preserve intratumoral heterogeneity and can predict clinical drug responses, enabling personalized oncology applications. Automated spheroid culture platforms and high-content imaging systems facilitate high-throughput drug screening in 3D formats.

3.2 Organoids: Miniaturized Organ Models

Organoids—self-organized 3D structures derived from stem cells or organ progenitors—represent the most sophisticated in vitro tissue models, recapitulating organ architecture, multiple cell types, and functional properties. Organoid generation typically involves embedding pluripotent or tissue-specific stem cells in Matrigel and providing morphogens and growth factors that guide self-organization through developmental pathways (Dutta et al., 2017). The resulting structures contain multiple differentiated cell types arranged in organ-specific patterns: intestinal organoids develop crypts and villi; brain organoids form cortical layers; kidney organoids contain nephron-like structures.

Patient-derived organoids established from biopsies or surgical specimens preserve individual genetic backgrounds and disease phenotypes, enabling personalized medicine applications. Cystic fibrosis patient-derived airway organoids maintain CFTR mutations and can predict responses to CFTR modulators, guiding treatment selection (Dutta et al., 2017). Cancer patient-derived organoids (PDOs) grown from tumor biopsies preserve tumor heterogeneity and genetic complexity, providing platforms for drug sensitivity testing that correlate with clinical responses. Biobanks of PDOs representing diverse cancer types and genotypes facilitate drug discovery and resistance mechanism studies.

Organoid limitations include lack of vascular networks limiting size and causing central necrosis, absence of immune components unless specifically co-cultured, and variability in size and differentiation between organoids and experiments (Jensen & Teng, 2020). Standardization efforts focus on defined culture media replacing Matrigel, automated culture systems for reproducibility, and quantitative characterization assays. Integration of organoids with microfluidic systems (organs-on-chips) addresses vascularization and enables perfusion, mechanical forces, and multi-organ interactions.

4. Organs-on-Chips and Microfluidic Systems

4.1 Microfluidic Culture Platforms

Organs-on-chips combine 3D cell culture with microfluidic systems, creating dynamic in vitro models that recapitulate organ-level physiology including perfusion, mechanical forces, and multi-tissue interfaces. These devices typically comprise transparent polymer channels (often PDMS—polydimethylsiloxane) containing cell-laden compartments with controlled fluid flow mimicking blood or interstitial fluid circulation (Ingber, 2016). Continuous perfusion provides nutrients, removes waste, and establishes shear stress on cells, influencing differentiation and function. Mechanical actuation can mimic breathing motions in lung-on-chip or peristalsis in gut-on-chip devices.

The lung-on-chip, a pioneering example, comprises two microfluidic channels separated by a porous membrane with alveolar epithelial cells on one side and endothelial cells on the other, creating an air-liquid interface (Ingber, 2016). Cyclic mechanical stretching mimics breathing motions, inducing physiological changes in gene expression, barrier function, and surfactant production. This system recapitulates pulmonary edema, bacterial infection, and drug-induced toxicity more accurately than static cultures or animal models. Similar approaches have created liver-on-chip for drug metabolism studies, kidney-on-chip for nephrotoxicity assessment, and blood-brain barrier-on-chip for CNS drug delivery.

Table 2: Applications of 3D Cell Culture Systems Across Biomedical Research Domains

Research Domain	3D Culture Type	Specific Applications	Key Advantages Over 2D	Notable Examples/Findings	Current Limitations
Cancer Research	Tumor spheroids, PDOs	Drug screening, resistance mechanisms, metastasis modeling	Recapitulate hypoxia, gradients, drug resistance	PDO sensitivity predicts drug response	Lacks tumor microenvironment complexity
Drug Discovery	Spheroids, organoids, organ-chips	Efficacy screening, toxicity testing, PK/PD modeling	Improved clinical translation, reduced animal use	Liver-on-chip predicts human hepatotoxicity	Throughput lower than 2D, higher cost

<i>Regenerative Medicine</i>	Bioprinted tissues, organoids	Tissue grafts, cell therapy products, transplantation	Functional tissue constructs, patient-specific	Bioprinted skin grafts in clinical trials	Vascularization, innervation challenges
<i>Disease Modeling</i>	Patient-derived organoids	Genetic diseases, infectious diseases, pathophysiology	Maintain patient mutations, physiological relevance	CF organoids predict drug response	Lacks systemic/immune components
<i>Stem Cell Biology</i>	Organoids, 3D differentiation	Developmental studies, cell fate determination, niche modeling	Recapitulate in vivo developmental programs	Cerebral organoids model cortical development	Variability, limited maturation
<i>Immunology</i>	3D tumor models, lymphoid organoids	Immune-cancer interactions, vaccine testing, inflammation	Cell-cell interactions, migration, tissue context	3D models improve CAR-T screening	Difficult to incorporate immune cells
<i>Toxicology</i>	Liver/kidney organoids, multi-organ chips	Hepatotoxicity, nephrotoxicity, systemic effects	Organ-specific metabolism, human-relevant	Multi-organ chip predicts systemic toxicity	Limited organ crosstalk, complexity

Note. PDOs = patient-derived organoids; PK/PD = pharmacokinetics/pharmacodynamics; CF = cystic fibrosis; CAR-T = chimeric antigen receptor T cell.

4.2 Multi-Organ Integration and Physiological Modeling

Linking multiple organs-on-chips creates body-on-chip systems modeling inter-organ communication, systemic drug distribution, and multi-organ toxicity. A drug metabolized by liver-on-chip produces metabolites that flow to kidney-on-chip for excretion or heart-on-chip to assess cardiotoxicity, recapitulating pharmacokinetic/pharmacodynamic relationships in humans (Ingber, 2016). These integrated systems enable assessment of off-target effects, drug-drug interactions, and cumulative toxicity impossible to evaluate in isolated single-organ cultures or even animal models with different metabolic pathways.

Challenges for organs-on-chips include complexity of fabrication requiring microfabrication expertise, difficulty maintaining multiple cell types with different media requirements in connected systems, and scaling for high-throughput applications (Ingber, 2016). Standardization

of device designs, protocols, and readouts is needed for regulatory acceptance and widespread adoption. Despite challenges, regulatory agencies including FDA are engaging with organ-chip developers to establish qualification processes, recognizing potential for improving drug development and reducing animal testing.

5. Applications in Drug Discovery and Development

5.1 Predictive Toxicology and Efficacy Screening

A major driver for 3D culture adoption is improved prediction of human drug responses. Attrition rates in drug development remain high, with many candidates failing in clinical trials due to lack of efficacy or unexpected toxicity not predicted by preclinical models (Antoni et al., 2015). 2D cultures oversimplify biology, while animal models have species-specific differences in drug metabolism, target expression, and physiological responses. 3D human cell models offer intermediate complexity with human-relevant biology.

Hepatotoxicity, a leading cause of drug attrition, is better predicted by 3D liver models than conventional hepatocyte cultures. Liver organoids and spheroids maintain metabolic enzyme expression, synthesize albumin and clotting factors, and respond to hepatotoxins with injury patterns resembling clinical liver damage (Antoni et al., 2015). Drug metabolism capacity persists longer in 3D than 2D hepatocyte cultures, enabling assessment of metabolite toxicity. Nephrotoxicity similarly benefits from kidney organoid models expressing transporters and metabolic enzymes mediating drug-induced renal injury.

Anticancer drug screening in 3D tumor models reveals resistance mechanisms masked in 2D cultures. Chemotherapy penetration into spheroid interiors is limited by diffusion barriers and cell density, mimicking poor drug penetration in solid tumors (Nunes et al., 2019). Hypoxic cores activate resistance pathways including HIF-1 α that are absent in 2D cultures with uniform oxygen. Dose-response curves in 3D typically show higher IC₅₀ values (lower potency) than 2D, better correlating with clinical dosing requirements. High-throughput 3D screening platforms combining automated spheroid formation, liquid handling, and image analysis enable testing thousands of compounds in physiologically relevant formats.

5.2 Personalized Medicine and Patient-Specific Testing

Patient-derived 3D cultures enable prospective drug testing to guide treatment selection, realizing precision medicine goals. Tumor organoids established from patient biopsies can be tested against panels of chemotherapies, targeted agents, and immunotherapies within weeks,

providing patient-specific drug sensitivity profiles before treatment initiation (Dutta et al., 2017). Clinical studies are demonstrating concordance between organoid drug responses and patient outcomes, validating this approach for treatment guidance.

Beyond oncology, patient organoids model genetic diseases and test therapeutic interventions. Cystic fibrosis intestinal organoids exhibit impaired CFTR function and can be rescued by CFTR modulators, with organoid responses predicting clinical benefit (Dutta et al., 2017). This enables precision medicine for CF where genotype alone imperfectly predicts drug responsiveness. Similar approaches apply to metabolic disorders, ciliopathies, and infectious diseases where patient-derived organoids model disease phenotypes and therapeutic responses.

6. Challenges and Future Directions

6.1 Standardization, Reproducibility, and Scalability

Despite tremendous promise, 3D culture faces challenges limiting widespread adoption. Standardization remains problematic, with protocols varying between laboratories and commercial reagent variability (particularly Matrigel batch effects) affecting reproducibility (Jensen & Teng, 2020). Organoid heterogeneity in size, morphology, and differentiation within and between experiments complicates quantitative analysis and inter-study comparisons. Standardized protocols, defined media formulations replacing Matrigel, and automated culture systems are being developed to address reproducibility.

Scalability for high-throughput applications requires adaptation of culture methods and analytical techniques. Manual organoid culture is labor-intensive and low-throughput; automated liquid handling systems, microwell formats, and bioprinted arrays are enabling scalability (Gunti et al., 2021). Analytical challenges include extracting RNA/protein from 3D structures, quantifying drug penetration, and assessing spatial heterogeneity. Advanced imaging including confocal microscopy, light-sheet microscopy, and high-content imaging provide spatially resolved data, while computational image analysis extracts quantitative metrics.

6.2 Emerging Technologies and Integration

Several emerging technologies are enhancing 3D culture capabilities. Bioprinting with multiple bioinks and cell types creates complex tissue architectures with defined spatial organization (Gunti et al., 2021). Sensors integrated into 3D cultures enable real-time monitoring of oxygen, pH, metabolites, and bioelectrical activity. Gene editing including CRISPR allows creation of isogenic organoid lines differing only in specific mutations, enabling causal variant analysis and

drug target validation. Immune cell co-cultures incorporating T cells, macrophages, and dendritic cells model immune-tumor interactions for immunotherapy development.

Integration of 3D cultures with other technologies creates powerful platforms. Combination with organ-on-chip microfluidics adds perfusion and mechanical forces (Ingber, 2016). Coupling with single-cell genomics reveals heterogeneity and identifies rare populations. Integration with artificial intelligence and machine learning enables predictive modeling, image analysis, and optimization of culture conditions. These convergent technologies will drive next-generation 3D culture systems with enhanced physiological relevance and analytical capabilities.

Conclusion

Three-dimensional cell culture systems have fundamentally transformed biomedical research by providing models that bridge the gap between oversimplified 2D cultures and complex in vivo environments. The recognition that dimensionality profoundly influences cellular behavior—morphology, gene expression, metabolism, differentiation, and drug responses—has driven adoption of 3D platforms across research domains. From simple spheroids to sophisticated organoids and organs-on-chips, these technologies offer unprecedented opportunities to study human biology in controlled yet physiologically relevant contexts.

The diverse 3D culture platforms available today—scaffold-based hydrogels and bioprinted constructs, scaffold-free spheroids and organoids, microfluidic organs-on-chips—each provide distinct advantages suited to specific applications. Hydrogels offer tunable biochemical and mechanical properties mimicking extracellular matrix. Spheroids provide simple, scalable models of tissue architecture and microenvironmental gradients. Organoids self-organize into miniature organs with remarkable cellular complexity and functional properties. Organs-on-chips add perfusion, mechanical forces, and multi-tissue interfaces approaching organ-level physiology.

Applications span the translational research spectrum. In drug discovery, 3D models improve prediction of human efficacy and toxicity, potentially reducing costly late-stage failures and animal use. Cancer research employs tumor spheroids and patient-derived organoids that preserve intratumoral heterogeneity and resistance mechanisms, enabling mechanistic studies and personalized drug testing. Regenerative medicine utilizes 3D engineered tissues for transplantation and patient-specific therapeutics. Disease modeling with patient-derived organoids recreates pathological phenotypes in vitro, facilitating drug development and precision medicine.

Significant challenges remain before 3D cultures fully replace 2D systems and animal models. Standardization of protocols, culture conditions, and analytical methods is essential for reproducibility and regulatory acceptance. Scalability for high-throughput applications requires automation and miniaturization while maintaining biological complexity. Vascularization of large constructs to overcome diffusion limitations represents a critical technical barrier. Cost reduction through defined media, reusable devices, and efficient manufacturing will expand accessibility beyond specialized laboratories.

Emerging innovations promise to address current limitations and expand 3D culture capabilities. Advanced biomaterials with precisely controlled properties will better recapitulate native ECM. Bioprinting technologies will create increasingly complex tissue architectures with defined cellular organization. Integration with sensors, microfluidics, and genetic engineering will enable real-time monitoring, dynamic control, and systematic perturbation of 3D systems. Artificial intelligence and machine learning will optimize culture conditions, analyze complex datasets, and predict therapeutic responses from 3D model data.

The trajectory of 3D cell culture technology points toward increasingly sophisticated models that faithfully recapitulate human tissue physiology while remaining experimentally tractable. As these systems mature and become more standardized and accessible, they will accelerate therapeutic development, enable true personalized medicine, reduce reliance on animal experimentation, and provide unprecedented insights into human biology in health and disease. The convergence of 3D culture with other cutting-edge technologies promises to usher in a new era of biomedical research where in vitro models approach the complexity and predictive power of in vivo systems while offering the control and accessibility that make them indispensable research tools.

References

1. Antoni, D., Burckel, H., Josset, E., & Noel, G. (2015). Three-dimensional cell culture: A breakthrough in vivo. *International Journal of Molecular Sciences*, 16(3), 5517-5527. <https://doi.org/10.3390/ijms16035517>
2. Dutta, D., Heo, I., & Clevers, H. (2017). Disease modeling in stem cell-derived 3D organoid systems. *Trends in Molecular Medicine*, 23(5), 393-410. <https://doi.org/10.1016/j.molmed.2017.02.007>
3. Duval, K., Grover, H., Han, L. H., Mou, Y., Pegoraro, A. F., Fredberg, J., & Chen, Z. (2017). Modeling physiological events in 2D vs. 3D cell culture. *Physiology*, 32(4), 266-277. <https://doi.org/10.1152/physiol.00036.2016>

4. Edmondson, R., Broglie, J. J., Adcock, A. F., & Yang, L. (2014). Three-dimensional cell culture systems and their applications in drug discovery and cell-based biosensors. *Assay and Drug Development Technologies*, 12(4), 207-218. <https://doi.org/10.1089/adt.2014.573>
5. Fatehullah, A., Tan, S. H., & Barker, N. (2016). Organoids as an in vitro model of human development and disease. *Nature Cell Biology*, 18(3), 246-254. <https://doi.org/10.1038/ncb3312>
6. Griffith, L. G., & Swartz, M. A. (2006). Capturing complex 3D tissue physiology in vitro. *Nature Reviews Molecular Cell Biology*, 7(3), 211-224. <https://doi.org/10.1038/nrm1858>
7. Gunti, S., Hoke, A. T. K., Vu, K. P., & London, N. R. (2021). Organoid and spheroid tumor models: Techniques and applications. *Cancers*, 13(4), 874. <https://doi.org/10.3390/cancers13040874>
8. Ingber, D. E. (2016). Reverse engineering human pathophysiology with organs-on-chips. *Cell*, 164(6), 1105-1109. <https://doi.org/10.1016/j.cell.2016.02.049>
9. Jensen, C., & Teng, Y. (2020). Is it time to start transitioning from 2D to 3D cell culture? *Frontiers in Molecular Biosciences*, 7, 33. <https://doi.org/10.3389/fmolb.2020.00033>
10. Langhans, S. A. (2018). Three-dimensional in vitro cell culture models in drug discovery and drug repositioning. *Frontiers in Pharmacology*, 9, 6. <https://doi.org/10.3389/fphar.2018.00006>
11. Murphy, S. V., & Atala, A. (2014). 3D bioprinting of tissues and organs. *Nature Biotechnology*, 32(8), 773-785. <https://doi.org/10.1038/nbt.2958>
12. Nunes, A. S., Barros, A. S., Costa, E. C., Moreira, A. F., & Correia, I. J. (2019). 3D tumor spheroids as in vitro models to mimic in vivo human solid tumors resistance to therapeutic drugs. *Biotechnology and Bioengineering*, 116(1), 206-226. <https://doi.org/10.1002/bit.26845>

Chapter 5:

Beyond the Live Bug: A Review on the Therapeutic Potential and Mechanistic Insights of Postbiotics

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Abstract

The burgeoning field of microbiome science has established the profound impact of gut health on overall human physiology, popularizing the use of live probiotic microorganisms. However, the administration of live bacteria presents inherent challenges related to stability, standardization, and safety, particularly in vulnerable populations. This has catalyzed the emergence of a novel therapeutic paradigm focused on "postbiotics"-preparations of inanimate microorganisms and/or their metabolic byproducts that confer a health benefit to the host. This review provides a comprehensive analysis of the postbiotic concept, framing it as a scientifically tractable and safer alternative to live probiotics. We dissect the complex composition of postbiotics, including inanimate microbial cells, crucial cell-wall components (e.g., peptidoglycan, teichoic acids), and a vast array of soluble factors and metabolites such as short-chain fatty acids (SCFAs), organic acids, enzymes, and antimicrobial peptides. The review systematically elucidates the multi-faceted mechanisms of action through which postbiotics exert their effects, including the enhancement of gut barrier function, direct modulation of host immune responses, competitive exclusion of pathogens, and metabolic signalling. We synthesize the growing body of experimental evidence from in vitro and in vivo models that demonstrates the efficacy of postbiotics in the context of gastrointestinal disorders, metabolic disease, and immune homeostasis. By critically evaluating the distinct advantages of postbiotics over probiotics, namely their superior safety profile, stability, and potential for precise dosing and by addressing the current challenges related to characterization and clinical validation, we outline a roadmap for the future of this exciting field, which is poised to deliver a new generation of microbiome-based therapeutics.

Keywords: Postbiotics, Probiotics, Gut Microbiome, Short-Chain Fatty Acids (SCFAs), Gut Barrier Function, Immunomodulation, Cell-Free Supernatant.

1. Introduction

In the past two decades, our understanding of human health has been revolutionized by the recognition that we are not solitary organisms, but complex ecosystems. The human gastrointestinal tract is home to a dense and dynamic community of trillions of microorganisms—the gut microbiota—whose collective genome, the microbiome, vastly outnumbers our own (Human Microbiome Project Consortium, 2012). This microbial community is not a passive bystander but an active metabolic "organ" that plays a fundamental role in digestion, nutrient synthesis, immune system development, and protection against pathogens. An imbalance in this community, termed dysbiosis, is now implicated in a vast array of human diseases, ranging from inflammatory bowel disease (IBD) and metabolic syndrome to autoimmune disorders and even neurological conditions (Lynch & Pedersen, 2016).

This paradigm shift has fueled an explosive growth in strategies aimed at modulating the gut microbiota for therapeutic benefit. The most well-known of these are probiotics, defined as "live microorganisms that, when administered in adequate amounts, confer a health benefit on the host" (Hill et al., 2014). The market for probiotic supplements and functional foods has grown into a multi-billion-dollar industry. Alongside probiotics are prebiotics, which are substrates (typically non-digestible fibers) that are selectively utilized by host microorganisms to confer a health benefit, essentially acting as "food" for beneficial gut bacteria.

Despite their widespread use and documented benefits for certain conditions, the administration of live probiotics is not without its challenges. The viability of the microorganisms is sensitive to manufacturing processes, storage conditions, and passage through the acidic environment of the stomach, leading to issues with product stability and inconsistent dosing. More significantly, safety concerns have been raised regarding the use of live bacteria in certain vulnerable populations, including the critically ill, premature infants, and immunocompromised individuals, where there is a potential risk of systemic infection (bacteremia) or adverse immune reactions (Didari et al., 2014).

These limitations have spurred scientific inquiry into a logical and compelling alternative: if the health benefits of probiotics are mediated by the structural components of the bacteria or the molecules they produce, is it necessary to administer the live "bug" itself? This question has given rise to the concept of "postbiotics." A postbiotic is, in essence, the beneficial effect of a probiotic, but delivered without the live microorganism. This approach offers the potential to harness the power of the microbiome with enhanced safety, stability, and standardization, representing a significant evolution in the field of gut health therapeutics. This review will

explore the definition, composition, mechanisms of action, and therapeutic potential of postbiotics, a field poised to move "beyond the live bug."

2. Literature Search Methodology

This comprehensive review was compiled through a systematic search of the scientific literature to synthesize the current understanding of postbiotics and their therapeutic potential. The literature search was conducted across major scientific databases, including PubMed, Scopus, and Web of Science, with Google Scholar used for supplementary searching. The search included publications up to December 2025.

The search strategy was designed to capture the evolution of the postbiotic concept. Keywords included "postbiotics," "paraprobiotics," "inactivated probiotics," "heat-killed bacteria," "cell-free supernatant," "metabolic byproducts," "probiotics," and "gut microbiota." These terms were combined with functional and mechanistic keywords such as "immunomodulation," "gut barrier," "anti-inflammatory," "short-chain fatty acids (SCFAs)," "tight junctions," and with disease-specific terms like "inflammatory bowel disease (IBD)," "necrotizing enterocolitis," and "irritable bowel syndrome (IBS)." Specific bacterial genera known for their probiotic effects, such as "*Lactobacillus*," "*Lactiplantibacillus*," and "*Bifidobacterium*," were also included in the search queries.

3. The "Biotics" Family: A Conceptual Framework

The terminology surrounding microbiome-based therapies has evolved rapidly. To understand postbiotics, it is useful to place them in the context of the broader "biotics" family.

Probiotics: The foundational concept. Live microorganisms (e.g., *Lactiplantibacillus rhamnosus* GG) that confer a health benefit when consumed in adequate amounts (Hill et al., 2014).

Prebiotics: The "food" for beneficial microbes. Non-digestible compounds (e.g., inulin, fructo-oligosaccharides) that are selectively utilized by host microorganisms, leading to a health benefit (Gibson et al., 2017).

Synbiotics: A synergistic combination of probiotics and prebiotics in a single product.

Postbiotics: The newest member of the family. In 2021, a consensus panel of experts convened by the International Scientific Association for Probiotics and Prebiotics (ISAPP) established a formal definition: **"a preparation of inanimate microorganisms and/or their components that confers a health benefit on the host"** (Salminen et al., 2021). This definition is critical because it provides clear boundaries. A postbiotic must be derived from a microorganism, it must be inanimate (e.g., heat-killed, UV-irradiated), and it must have demonstrated a health benefit in

a target population. This distinguishes postbiotics from purified microbial metabolites, which would be considered drugs, and from undefined fermented foods. The term "paraprobiotic" is often used synonymously with inactivated microbial cells, which now fall under the broader umbrella of postbiotics.

4. Composition and Characterization of Postbiotics

A postbiotic preparation is a complex mixture. Its beneficial effects can stem from various components, which can be broadly divided into the inanimate microbial cells themselves and the soluble factors they produced during fermentation.

4.1. Inanimate Cellular Components

Even when non-viable, the structural components of the microbial cell can have profound biological effects, primarily through interaction with the host's immune system.

Peptidoglycan (PGN): A major component of the cell wall of both Gram-positive and Gram-negative bacteria. PGN fragments can be recognized by host pattern recognition receptors (PRRs) like Toll-like receptor 2 (TLR2) and NOD-like receptors (NOD1, NOD2), triggering immune signalling cascades (Travassos et al., 2004).

Lipoteichoic Acid (LTA): Found in the cell wall of Gram-positive bacteria like *Lactiplantibacillus*. Like PGN, LTA is a potent immunomodulator that signals through TLR2.

Surface-Layer Proteins (SLPs): These proteins form a crystalline array on the surface of many probiotic bacteria. They are involved in adhesion to intestinal epithelial cells and can directly modulate host immune responses, often promoting anti-inflammatory signalling (Hynönen & Palva, 2013).

Pili and Fimbriae: These hair-like appendages, even when denatured, can interact with host cells and modulate immune signalling.

4.2. Soluble Metabolites (The "Secretome")

The cell-free supernatant from a probiotic fermentation is a rich cocktail of bioactive molecules.

Short-Chain Fatty Acids (SCFAs): These are the primary end-products of bacterial fermentation of dietary fiber. The main SCFAs—**butyrate, propionate, and acetate**—are metabolic superstars. Butyrate is the preferred energy source for colonocytes (the cells lining the colon), helping to maintain gut barrier integrity. All three SCFAs act as crucial signalling molecules, binding to host G-protein coupled receptors (GPCRs) like GPR41, GPR43, and GPR109A on various cell types, including immune cells and enteroendocrine cells, thereby influencing inflammation, metabolism, and appetite (Koh et al., 2016).

Other Organic Acids: Lactic acid and succinic acid contribute to lowering the colonic pH, which inhibits the growth of many pathogens.

Bacteriocins: These are small, ribosome-synthesized antimicrobial peptides (e.g., nisin) that can selectively kill competing bacteria, including pathogens, without harming the producing strain.

Vitamins and Amino Acids: Probiotic bacteria can synthesize essential vitamins, such as vitamin K and several B vitamins (e.g., folate, biotin), as well as functional amino acids.

Enzymes: Secreted enzymes like bile salt hydrolases (BSH) can deconjugate bile acids, impacting host lipid metabolism.

5. Mechanisms of Action

The therapeutic effects of postbiotics arise from their ability to modulate host physiology through several key mechanisms.

5.1. Enhancement of Gut Barrier Function:

A healthy intestinal barrier is critical for preventing the translocation of harmful substances (like bacterial endotoxins) from the gut lumen into the bloodstream. Postbiotics can strengthen this barrier in several ways:

They can upregulate the expression of genes encoding tight junction proteins, such as occludin, claudins, and zonula occludens-1 (ZO-1), which form the "seals" between adjacent epithelial cells (Putaala et al., 2008).

SCFAs, particularly butyrate, provide energy to colonocytes, promoting their health and turnover.

They can stimulate the production of mucus, which forms a protective physical barrier over the epithelium.

5.2. Immunomodulation:

This is perhaps the most significant mechanism of postbiotic action. By interacting with immune cells in the gut-associated lymphoid tissue (GALT), postbiotics can help to maintain immune homeostasis.

Cell wall components (PGN, LTA) are recognized by TLRs on immune cells like dendritic cells and macrophages. This interaction can trigger signalling pathways (e.g., NF- κ B, MAPKs) that lead to the production of cytokines.

Crucially, the outcome of this signalling is context-dependent and strain-specific. Some postbiotics promote a pro-inflammatory response (e.g., producing IL-12) that is beneficial for clearing pathogens. Others promote an anti-inflammatory response by inducing the production of cytokines like IL-10 and TGF- β , and by driving the differentiation of naïve T cells into regulatory T cells (Tregs), which are essential for suppressing excessive inflammation (Šrůtková et al., 2015).

5.3. Antimicrobial Activity:

Postbiotics can directly inhibit the growth of enteric pathogens. This is achieved through the action of secreted bacteriocins and the lowering of gut pH by organic acids, creating an environment that is inhospitable to many harmful bacteria like *Salmonella* and *Clostridium difficile* (Gareau et al., 2010).

5.4. Metabolic and Signalling Effects:

SCFAs produced by probiotics are potent signalling molecules. By binding to their cognate GPCRs on enteroendocrine L-cells, they can stimulate the release of gut hormones like GLP-1 and PYY, which regulate glucose homeostasis and promote satiety. Their influence on immune cells via GPCRs also links gut metabolism directly to inflammatory status (Koh et al., 2016).

6. Therapeutic Potential and Experimental Evidence

The multi-faceted mechanisms of postbiotics give them broad therapeutic potential.

In Vitro Evidence: A wealth of *in vitro* studies has demonstrated the bioactivity of postbiotics. Cell-free supernatants from *Lactiplantibacillus plantarum* have been shown to protect Caco-2 intestinal cell monolayers from damage and to reduce the inflammatory response induced by LPS. Inactivated cells and purified cell wall components have been shown to directly modulate cytokine production by peripheral blood mononuclear cells (PBMCs) and dendritic cells, often promoting an anti-inflammatory IL-10 response.

In Vivo Evidence: Animal models have provided strong evidence for the efficacy of postbiotics in various disease contexts.

Inflammatory Bowel Disease (IBD): In mouse models of colitis (e.g., DSS-induced colitis), oral administration of heat-killed *L. plantarum* or its cell-free supernatant has been shown to significantly reduce disease severity, decrease inflammatory cell infiltration in the colon, restore gut barrier integrity, and rebalance the cytokine profile (Taverniti & Guglielmetti, 2011).

Necrotizing Enterocolitis (NEC): In neonatal rat models of NEC, a devastating inflammatory disease of the gut in premature infants, postbiotics have shown protective effects, reducing intestinal damage and mortality. This is particularly important given the safety concerns of using live probiotics in this fragile population.

Metabolic Syndrome: In mice fed a high-fat diet, administration of postbiotics has been shown to improve insulin sensitivity, reduce weight gain, and lower systemic inflammation, often linked to the beneficial effects of SCFAs.

7. Advantages of Postbiotics Over Probiotics

The shift towards postbiotics is driven by several clear and compelling advantages:

1. **Safety:** This is paramount. As non-viable preparations, postbiotics carry no risk of causing infection, making them suitable for immunocompromised patients, the critically ill, and premature infants. There is also no risk of transferring antibiotic resistance genes.
2. **Stability:** Postbiotics are resistant to heat, pressure, and enzymatic degradation. This means they have a much longer shelf life and are far easier to incorporate into a wide range of products, from shelf-stable foods and beverages to pharmaceuticals, without the need for a cold chain.
3. **Standardization and Dosing:** The composition of a postbiotic can be chemically analyzed, and the concentration of key components (e.g., cell mass, SCFA levels) can be standardized. This allows for precise, reproducible dosing, which is a fundamental requirement for pharmaceutical development and a major challenge for live probiotics, whose viable count can vary significantly.

8. Clinical and Translational Relevance

The application of postbiotics in human health is a rapidly growing field.

Infant Nutrition: Several infant formulas are now supplemented with heat-inactivated *Bifidobacterium lactis*, which has been shown in clinical trials to support immune health and reduce gastrointestinal infections in infants (Salminen et al., 2021).

Gastrointestinal Disorders: Clinical trials have shown that certain postbiotic preparations can be effective in managing symptoms of irritable bowel syndrome (IBS) and in reducing the incidence and duration of infectious diarrhoea.

Dermatology: Topically applied postbiotics are being used in skincare products to modulate the skin microbiome, reduce inflammation, and strengthen the skin barrier.

9. Current Limitations and Knowledge Gaps

As a nascent field, the study of postbiotics faces several important challenges.

Standardization of Production: There is a need for standardized protocols for the production of postbiotics (e.g., specific inactivation methods, fermentation conditions) to ensure product consistency and allow for meaningful comparison between studies.

Identification of Active Molecules: A postbiotic preparation is a complex mixture. A major challenge is "de-convoluting" this mixture to identify the specific molecule or combination of molecules responsible for the observed health benefit.

Lack of Large-Scale Clinical Trials: The single greatest hurdle is the need for more large-scale, well-designed, placebo-controlled randomized controlled trials (RCTs) to definitively establish the efficacy of specific postbiotic preparations for specific health indications in human populations.

10. Future Directions

The future of postbiotics is focused on moving from a complex mixture to a more defined and engineered therapeutic.

Mechanism-Driven Discovery: A more systematic approach is needed, starting with a desired health outcome (e.g., reducing gut inflammation) and screening libraries of postbiotic preparations from diverse probiotic strains (e.g., novel *Lactiplantibacillus* strains from traditional fermented foods) to find the most potent candidates.

Advanced Characterization: The application of advanced 'omics' technologies—metabolomics, proteomics, and lipidomics—will be crucial for comprehensively characterizing the composition of postbiotic preparations and identifying novel bioactive molecules.

Bio-engineering and Synthetic Postbiotics: Once key active molecules are identified, they can be produced in pure form via chemical synthesis or recombinant fermentation. This leads to the possibility of creating "synthetic postbiotics"—rationally designed cocktails of purified components tailored for a specific therapeutic effect.

Targeted Delivery Systems: Advanced formulation technologies, such as encapsulation in pH-sensitive polymers, can be used to develop delivery systems that release postbiotic components in specific parts of the gastrointestinal tract (e.g., the colon) to maximize their local effect.

Conclusion

The concept of postbiotics represents a significant and logical evolution in our approach to modulating the gut microbiome for health. By moving "beyond the live bug" to the inanimate microorganisms and their beneficial metabolites, we can overcome the primary challenges of safety, stability, and standardization that have limited the pharmaceutical development of probiotics. Postbiotics act through a sophisticated array of mechanisms, including strengthening the gut barrier, balancing the immune system, and providing key metabolic signals to the host. While the field is still young and requires a concerted effort in standardization and rigorous clinical validation, postbiotics hold immense promise. They represent a bridge between traditional probiotic concepts and modern pharmaceutical science, paving the way for a new generation of precisely defined, safe, and highly effective microbiome-based therapeutics.

References

- Didari, T., et al. (2014). The safety of probiotics in clinical use. *Expert Opinion on Drug Safety*, 13(7), 935–947.
- Gareau, M. G., et al. (2010). Probiotic-induced suppression of *E. coli* O157:H7-mediated intestinal epithelial cell damage. *Applied and Environmental Microbiology*, 76(19), 6533–6541.
- Gibson, G. R., et al. (2017). Expert consensus document: The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. *Nature Reviews Gastroenterology & Hepatology*, 14(8), 491–502.
- Hill, C., et al. (2014). Expert consensus document: The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nature Reviews Gastroenterology & Hepatology*, 11(8), 506–514.
- Human Microbiome Project Consortium. (2012). Structure, function and diversity of the healthy human microbiome. *Nature*, 486(7402), 207–214.
- Hynönen, U., & Palva, A. (2013). Lactobacillus surface layer proteins: From self-assembly to designer biomaterials. *Applied Microbiology and Biotechnology*, 97(23), 9877–9896.
- Koh, A., et al. (2016). From dietary fiber to host physiology: Short-chain fatty acids as key bacterial metabolites. *Cell*, 165(6), 1332–1345.
- Lynch, S. V., & Pedersen, O. (2016). The human intestinal microbiome in health and disease. *The New England Journal of Medicine*, 375(24), 2369–2379.
- Putala, H., et al. (2008). Probiotic bacteria and their cell wall components can protect intestinal epithelial barrier function from damage induced by enteropathogenic *Escherichia coli*. *Gut*, 57(6), 762–770.
- Salminen, S., et al. (2021). The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of postbiotics. *Nature Reviews Gastroenterology & Hepatology*, 18(9), 649–667.
- Šrůtková, D., et al. (2015). The effect of a multispecies probiotic mixture on the immune response in a mouse model of colitis. *Journal of Functional Foods*, 17, 308–316.
- Taverniti, V., & Guglielmetti, S. (2011). The immunomodulatory properties of probiotic microorganisms beyond their viability (ghost probiotics: Paraprobiotics). *International Journal of Molecular Sciences*, 12(11), 7853–7874.
- Travassos, L. H., et al. (2004). Toll-like receptor 2-dependent bacterial sensing does not occur via peptidoglycan. *EMBO Reports*, 5(10), 1000–1006.

Chapter 6:

Newer Insights Into Nanoparticle Based SiRNA Delivery In Cancer Cells

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Abstract

With a complicated pathophysiology, cancer is one of the main causes of mortality and morbidity. Chemotherapy, radiation therapy, targeted therapy, and immunotherapy are examples of traditional cancer treatments. However, restrictions like cytotoxicity, lack of selectivity and multi-drug resistance provide a significant obstacle to effective cancer treatment. One of the most innovative methods for conducting gene therapy is RNA interference (RNAi), which may inhibit the transcription or expression of particular genes. Small interfering RNA (siRNA) offers enormous therapeutic potential for diseases like cancer caused on by aberrant gene overexpression or mutation. The stability and efficient delivery of siRNA in vivo are the main obstacles to the use of siRNA therapies. An immune-hostile tumor microenvironment and physiological circulatory system hurdles make the administration of stable siRNA difficult. The efficient intracellular siRNA delivery is promised by the advancements in nanotechnology and nanoparticle-mediated delivery technologies. For siRNA to be effectively delivered into cancer cells, safe, stable, and effective nanoengineered delivery methods must be designed and validated. This review synthesizes the latest developments in nanoparticle delivery methods for siRNA-based cancer treatments and the challenges and future developments of siRNA delivery vectors.

Keywords: - Cancer, Chemotherapy, Mutation, Nanotechnology, Nanoparticles, RNAi, siRNA, Transcription, Tumor microenvironment.

Introduction

Cancer continues to be a primary cause of morbidity and mortality worldwide, and the disease burden is rapidly increasing (Mishra et al 2017). An estimated 9.7 million people died from cancer and 20 million new cases were reported in 2022. It was projected that 53.5 million people survived five years after receiving a cancer diagnosis. 1 in 5 people will get cancer at some point in their lives and 1 in 9 men and 1 in 12 women die from it (WHO, 2024). Multiple genes exhibit abnormalities in their function during the genesis and later development of cancer. These abnormalities may result from the overexpression of normal genes or from alleles that have mutations and abnormal functions. Thus, to some extent, tumors can also be referred to as genetic diseases (Chen et al 2021). Treatment for this disease is extremely tough due to its multifaceted character (Isazadeh et al 2023). The three primary traditional cancer-based treatment modalities are radiation, chemotherapy, and surgery. Furthermore, a number of modern therapies have surfaced in recent years, including immunotherapy, gene therapy, chemodynamic therapy, photodynamic therapy (PDT), photothermal therapy (PTT), and others. Chemotherapy is the most commonly used therapeutic approach among various treatment modalities; yet, it inevitably causes adverse effects on healthy tissues. Similar to this, other therapeutic approaches have not produced good clinical outcomes. Since important carcinogenic markers and pathways have been identified, gene therapy has emerged as a new class of pharmacological medications with notable therapeutic benefits in the battle against cancer. Gene therapy exhibits superior selectivity and specificity for related genes when compared to chemotherapy. (Chen et al 2021).

Recent advances in integrated knowledge systems and precision medicine have begun to offer rationally designed personalized treatments that are based on specific therapeutic targets and the selective modulation of the molecular mechanisms of disease (Isazadeh et al. 2023). The molecular mechanism of gene silencing of a specific mRNA corresponds to the degradation of that mRNA by RNA molecules that are antisense to the mRNA coding sequence, thereby preventing the translation of that mRNA into its corresponding proteomic product. This process is termed RNA interference (RNAi). In the first stage of RNAi, longer double-stranded RNA is processed and cleaved into siRNA molecules that contain a 2-nucleotide overhang at the 3' end of each strand. This processing is the responsibility of the RNase III-like enzyme known as Dicer. The RNA-induced silencing complex (RISC) is one of the several RNA silencing complexes that bind to the processed siRNA. The more stable 5'-end of the siRNA strand is usually integrated into the active RISC complex once the siRNA strands are separated within the RISC complex. After the antisense single-stranded siRNA component directs and aligns the RISC complex on the target mRNA, the mRNA is cleaved by the catalytic RISC protein, which is a member of the argonaute family (Ago2). Increasing understanding of the molecular

mechanisms underlying endogenous RNA interference has led to the development of siRNAs as novel nucleic acid medications for the treatment of cancer and other incurable disorders. In 2001, Elbashir and colleagues successfully employed synthetic siRNAs for silencing and established the fundamentals of siRNA structure and RNAi dynamics, laying the groundwork for the creation of RNAi applications (Dana et al. 2017). RET/PTC in papillary thyroid carcinoma, HER2 in nasopharyngeal tumor, VEGF in tumor angiogenesis, PD-L1 in ovarian cancers, FAK in pancreatic adenocarcinoma, and EWS-Flil gene expression in Ewing's sarcoma have all been suggested as targets for this approach (Ali et al. 2012).

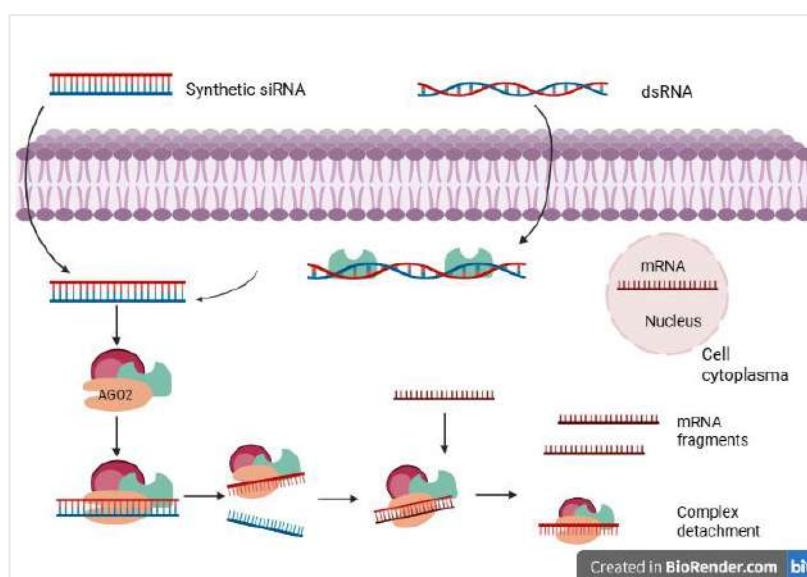


Fig1: RNA interference mediated by siRNA (Picture created through Biorender)

Through the silencing of particular genes linked to the progression of cancer, siRNA, a significant kind of gene expression regulation, opens up new therapeutic options. It has numerous benefits for treating tumors, including minimal dosage needs, good specificity, and a comparatively easy drug development procedure. However, there are a number of barriers to the efficient distribution and efficacy of siRNA technology. Naked siRNA is a type of physiologically active nucleic acid material with great specificity; yet, the following issues prevent its practical use: 1) siRNA is easily broken down by RNase and extremely unstable in the body's physical environment. 2) Since the cell membrane is a negatively charged hydrophobic structure and the siRNA molecule is a negatively charged hydrophilic material, free siRNA finds it difficult to penetrate the cell by interacting with the cell membrane; 3) siRNA will trigger the release of inflammatory cytokines and interferons and activate the body's innate immune system upon entering; 4) siRNA may cause toxicity by degrading the target mRNA as well as the miRNA that the cell expresses. Therefore, the main obstacle to

siRNA's therapeutic use is its delivery. One of the key areas of siRNA medication research is now the creation and manufacture of secure and efficient siRNA delivery vehicles (Chen et al. 2021). Currently, rapid growth in nanotechnology provides new possibilities in the development of anticancer therapies using siRNA. siRNA can be delivered to tumor cells to shut down specific genes and can be transported to tumor sites by nanoparticles through the enhanced permeability and retention (EPR) effect. Chemotherapy, as well as other innovative cancer therapies, may one day integrate siRNA with the help of future clinician-scientists, translational medical researchers, and materials scientists to refine the best siRNA delivery vectors and remove the barriers to their clinical use (Chen et al. 2021).

1. NANOPARTICLES AS CARRIER OF siRNA DELIVERY:

The cytoplasm of cancer cells is the target of siRNA used in cancer treatment to produce RNA interference. A good siRNA vector should encourage endocytosis, enrich siRNA at the target site, and greatly increase its biological stability. Nanoparticles offer the following benefits for siRNA delivery: 1) shielding and limiting premature degradation of siRNA; 2) enhancing the capacity to target tumors using the passive or active targeting action of nanoparticles; 3) internalization through nanoparticle endocytosis; 4) immune response avoidance through siRNA encapsulation within the nanoparticles. Therefore, nanocarriers can effectively address the issues associated with siRNA administration, such as simple degradation, poor targeting ability, poor internalization ability, easy activation of immune response, and so forth. Viral and nonviral vectors are now the two primary types of vectors used in siRNA delivery methods (Chen et al. 2021). The inherent ability of viral vectors to transfer genetic material into cells makes them superior in terms of release; nevertheless, their usage is restricted by significant toxicities (Mishra et al. 2017). An alternative to viral vectors is nonviral vectors. Numerous non-viral vectors have been created and are now a widely used and effective research tool for determining the structure, regulation, and function of genes. Despite being less effective than viruses at delivering genes, they have a number of benefits, including ease of use, trouble-free quality control, biocompatibility, and the fact that they elicit little to no particular immune reaction (Ali et al. 2012). Numerous natural and artificial nanocarriers, including liposomes, micelles, exosomes, inorganic materials (carbon nanotubes, quantum dots, and gold nanoparticles), and synthetic organic polymers (polyethyleneimine (PEI), dendrimer, and cyclodextrin) have been developed for effective siRNA delivery. A small number of these have even been subjected to clinical testing (Mishra et al. 2017).

2. LIPID-BASED NANOPARTICLES:

The most preferred substance for siRNA delivery is lipid-based nanoparticles, and liposomes are now employed in 20% of gene therapy clinical trials. The review by Kim et al. revealed that, with average knockdown efficiency of $79.3 \pm 15.2\%$ and $80.4 \pm 13.8\%$, respectively, lipid-based methods often exhibit quite high gene silencing effects both *in vitro* and *in vivo* (Chen et al. 2021). Lipid-based nano vectors for siRNA have been employed by the majority of research labs and biotechnological/pharmaceutical enterprises (Shen et al. 2012).

2.1. Liposomes/ lipoplexes:

Liposomes/lipoplexes have been thoroughly investigated as nonviral delivery systems for plasmids and siRNA (Lee et al. 2013). Lipoplexes are complexes that form between nucleic acids, primarily plasmid DNA, and cationic lipids. Neutral liposomes have better pharmacokinetics and are more biocompatible than cationic lipids, but because neutral lipids and anionic polynucleotides do not interact during formation, they have poor entrapment efficiency. Landen Jr. et al. created a technique for creating 1,2-dioleoylsn-glycero-3-phosphatidylcholine-(DOPC-) encapsulated siRNA liposomes that entails dissolving siRNA and DOPC in excess tertiary-butanol with the non-ionic detergent Tween 20 in order to improve entrapment efficiency. In an orthotopic model of ovarian cancer, DOPC-encapsulated siRNA that targeted the oncoprotein EphA2 was very successful in lowering EphA2 expression 48 hours after a single dosage was administered. Intravenous or intraperitoneal injections of DOPC-encapsulated siRNA were very successful in decreasing tumor weight and *in vivo* expression of target genes (such as EphA2, FAK, neuropilin-2, or IL-8) in animal models of various human malignancies (Mishra et al. 2017).

Negatively charged siRNA forms lipoplexes with cationic lipids, such 1,2-dioleoyl-3-trimethylammonium-propane (DOTAP) and dioleoyl phosphatidylethanolamine. *In vitro* transfer of plasmid DNA or siRNA into mammalian cells is frequently accomplished using cationic liposomes. Despite their effective siRNA uptake, cationic liposomes have had limited success in *in vivo* gene silencing, most likely because of their intracellular stability and consequent inability to release siRNA contents. Lastly, the toxicity of cationic liposomes has restricted their efficacy. By encouraging the generation of reactive oxygen intermediates, the use of cationic liposomes *in vivo* caused pulmonary inflammation and dose-dependent damage. When liposomes are coated with hydrophilic molecules like polyethylene glycol (PEG), the reticuloendothelial system (RES) absorbs less, increasing the circulatory half-life. Santel et al. created a unique liposomal siRNA formulation in 2006 called siRNA-lipoplex/AtuPLEX, which is based on cationic lipids and contains neutral fusogenic and PEG-modified lipid components for better cellular absorption, pharmacokinetics, and siRNA release

efficiency. By using this formulation to target genes specific to endothelia, including TIE-2 or CD31 (platelet endothelial cell adhesion molecule-1), they showed that the related mRNAs and proteins were downregulated in mice. Atu027 is, a lipoplexed siRNA molecule, particularly targeting the expression of protein kinase N3, a downstream effector of the phosphoinositol-3-kinase signalling cascade. According to reports, Atu027 inhibits hematogenous metastasis to the target organ lung in a variety of animal lung metastasis models and inhibits lymph node metastasis in orthotopic prostate and pancreatic cancer mouse models. In patients with colorectal cancer that has spread to the liver, Silence Therapeutics (London, UK) is conducting a phase I trial with Atu027, which was well tolerated up to a dose of 0.180 mg/kg and was not linked to dose-dependent toxicities (Lee et al. 2013).

2.2. Stable Nucleic Acid Lipid Particles and Lipidoids:

For the systemic administration of siRNA, solid lipid-based systems such as cationic solid-lipid nanoparticles and stable nucleic acid lipid particles (SNALPs) have been created as substitutes for emulsions, liposomes, microparticles and polymeric nanoparticles. Jeffs and colleagues came up with a new method, called spontaneous vesicle formation, that quickly and dependably manufactures stabilized plasmid lipid particles for non-viral, systemic gene therapy. Morrissey and colleagues created SNALPs (PEGylated lipid nanoparticles) using this method. These nanoparticles contain siRNA that is surrounded by a lipid bilayer with neutral lipids and PEG-lipid fusion promoters. In animals with replicating HBV, siRNA that is stable and targets the hepatitis B virus (HBV) RNA was incorporated into SNALPs, and upon intravenous administration, the amount of HBV DNA was decreased. In addition, a weekly dose sustained decreases in blood HBV DNA for up to six weeks. Zimmermann and colleagues noted a significant dose-dependent suppression of ApoB mRNA in the livers of mice and nonhuman primates following intravenous administration of SNALPs containing ApoB-targeting siRNAs. After one administration of 2.5 mg/kg SNALP-formulated siRNA, ApoB mRNA expression in the liver was reduced by 90%, and this was well tolerated for 11 consecutive days (Lee et al. 2013).

Lipidoid nanoparticles are tailored delivery agents, consisting of PEG-modified lipids, cholesterol, and lipids that are designed to deliver specific siRNAs. Akinc et al. created a novel chemical technique to enable the quick synthesis of a sizable library of lipidoids and evaluated their effectiveness in siRNA delivery in order to enhance SNALP-mediated delivery. The 98N₁₂-5 lipidoid-based siRNA formulation, the top candidate, demonstrated a 75%–90% decrease in the expression of ApoB or FVII factor in hepatocytes from mice and nonhuman primates. This formulation lowered toxicity by

enabling gene silencing at orders of magnitude lower siRNA doses than those needed by the original SNALP formulation (Lee et al. 2013).

3. POLYMERIC NANOPARTICLES:

Lipids and polymers are widely used materials for siRNA delivery. For siRNA delivery, polymer nanoparticles are usually made out of two kinds of polymers: cationic block copolymers and cationic polymers. Cationic polymers are more widely used. Because of their high positive charge, cationic polymers create electrostatic nanoparticles with siRNA's negative charge due to the phosphate groups (PO_3^{4-}) and the positive amine groups (NH_3^+). Presently, cationic polymers such as polyethyleneimine (PEI), poly-L-lysine (PLL), and cyclodextrin polymers are being used more than all others. PEI is considered to be an excellent vector for gene transfer among them due to its high gene transfection efficiency. It acts as a "proton sponge" in tumor cells, facilitating endosome escape and cell uptake, enhancing transfection effectiveness, and shielding oligonucleotides from corresponding nuclease degradation. However, PEI exhibits extreme cytotoxicity, which varies according to the number of branches and molecular weight. Low molecular PEI is an effective method to reduce cytotoxicity. Previous studies have been conducted in which a low molecular weight PEI/siRNA complex was intraperitoneally injected at tumor sites, and demonstrated that the complex was able to silence the HER2 gene and inhibit tumor growth. Adding PEG to the complex may reduce the cytotoxicity of PEI. Kim et al. prepared and synthesized a polyelectrolyte complex of PEI/PEG conjugated to siRNA. Once it gathered in the tumor region, the vascular endothelial growth factor (VEGF) deletion prevented microvessel formation, which in turn prevented the growth of the tumor. PEG modification also helps to lower immunogenicity. Prior research has demonstrated the widespread usage of polymer carriers based on chitosan and poly(L-lactide) PEG-poly(L-lactide) triblock copolymer (PLLA-PEG-PLLA), which exhibit low immunogenicity and strong biocompatibility, in gene and drug delivery. It is important to remember that PEG is not very biodegradable. It will build up in the lysosome when taken in large quantities or over an extended period of time, which will hinder the regular function of some catabolic lysosomal enzymes. Consequently, it is best to employ as much low molecular weight PEG as feasible (Chen et al. 2021).

4. INORGANIC NANOPARTICLES:

There are several inorganic nanoparticles that are thought to be promising siRNA delivery vehicles for both therapeutic and imaging purposes. They consist of carbon nanotubes and metallic material such as gold (Mishra et al. 2017).

4.1. Carbon Nanotubes (CNTs)

CNTs are nanomaterials with intriguing chemical and physical characteristics that have just lately come to light as a novel approach to gene therapy, bioengineering, and cancer treatment. Because of their nanoneedle shape, it has been suggested that CNTs can readily pass through the plasma membrane and move straight into the target cells' cytoplasm via a mechanism that is independent of endocytosis and does not cause cell death. Single-walled and multiwalled carbon nanotubes are the two types of CNTs. For the transport of siRNA, a number of functionalized CNTs have been developed and tested. Single-walled CNTs functionalized with $\text{CONH}-(\text{CH}_2)_6-\text{NH}_3^+ \text{Cl}^-$ were employed by Zhang et al. as siRNA carriers. They significantly reduced tumor growth by releasing siRNA from the side-wall of the nanotube to inhibit the production of telomerase reverse transcriptase. It has been demonstrated that CNTs functionalized with amine-terminated PEG (phospholipid (PL)-PEG2000- NH_2) are effective at delivering siRNA to human T cells. Dendron-CNTs and ammonium-functionalized CNTs have also been shown to be effective siRNA delivery vehicles with minimal cytotoxicity. Nonetheless, despite the lack of clarity surrounding the underlying mechanisms, a number of research have examined the possible toxicity of CNTs (Lee et al. 2013).

4.2. Gold Nanoparticles:

Gold nanoparticles (AuNPs) have become a viable siRNA delivery carrier because of their high surface-to-volume ratio, ease of manufacturing, outstanding biocompatibility, and ease of surface functionalization. SiRNA delivery using AuNPs has been thoroughly investigated (Lee et al. 2013). Gold has unusual properties. It can be shaped into many different sizes, shapes, and even particles as small as 1-100 nanometers. Due to their unique physico-chemical properties, gold nanoparticles (AuNPs) have been widely used as radiosensitizers, contrast agents, and as photothermal agents in the diagnosis and treatment of cancer. Compared to traditional carriers, the surface chemistry of gold makes AuNPs ideal for the delivery of siRNA. As an example, AuNPs can be paired with amine and thiol groups, which allows for surface modification with functional moieties, which can include stabilizing groups, bioactive polymers, and targeting ligands. This technique has been used to enable targeted siRNA delivery for cancer therapy. In recent studies, it has been determined that via bioactive targeting ligands, it is possible to achieve the specific delivery of site- modified delivery of AuNPs to particular target cells or organs. In response to cell surface receptor [transferrin receptor (TfR) and folate receptor (FR)]- overexpression, two different bioconjugated AuNPs were created for the study: 1) an anionic transferrin (Tf) tagged AuNP, AuNPs-PEG-Tf, and 2) a cationic folic acid (FA) tagged AuNP, AuNPs-PEI-FA, for prostate cancer. For the initial screening of AuNPs as nontoxic gene delivery vectors, the AuNPs were characterized and their cell-specific uptake quantified (Guo et al.

2016). Furthermore, gold nanorods can also be utilized in the delivery of siRNA to target cells or tissues (Lee et al. 2013).

4.3 Silica Nanoparticles:

The discovery of Mobile Crystalline Material-41 (MCM-41), has sparked significant research, particularly with regards to the modification and application of mesoporous silica materials in the field of drug delivery. The first recorded drug delivery application with mesoporous silica nanoparticles (MSNPs) occurred in 2001. These nanoparticles are particularly advantageous because they can be modified to create an effect tissue specific accumulation, enhanced intracellular uptake and localization and controlled and/ or sustained therapeutic release. These attributes combined with their therapeutic loading and release potential, have allowed mesoporous silica nanoparticles to be used in the preclinical delivery of drug and nucleic acid therapies for the treatment of various diseases including cancer. An example of this is the work of Shen et al., where cyclodextrin and polyethylene imine functionalized MSNPs were used to deliver siRNA cancer therapeutics. Cyclodextrin-grafted polyethyleneimine (CP) is designed to electrostatically complex siRNA and then facilitates the endosomal escape of siRNA. Surface-functionalized MSNPs can be employed as effective and secure nanocarriers for siRNA delivery, according to Xia et al. To transport siRNA effectively, they added a functional group to the surface of MSNPs that improves and permits cellular absorption and siRNA delivery, respectively, in addition to conventional drug administration (Li et al 2016).

Table 1: Different Nanomaterials for siRNA Delivery

Nanocarriers	Advantages
1. Lipids	Gene silencing effect is strong, synthesis method is simple, adjustable continuous release behaviour
2. Polymers	Large surface to mass ratio, size is ultra-small and can be controlled, functionalizable structure, high thermodynamic stability and dynamic stability
3. Inorganic nanoparticles	Surface area is high, highly controllable morphology, particle structure is clear, large pore volume

CONCLUSION

In recent decades, siRNA-based gene therapy has received significant attention due to its ability to degrade mRNA in the cytoplasm and disrupt the expression of particular genes. SiRNA delivered by NPs may precisely and effectively cause cell death, hence reducing tumor formation and cell proliferation, by limiting the gene expression or protein translation of cancer-related proteins. SiRNA can be effectively delivered to tumor cells via nanocarriers, where it will decrease gene expression. Currently, a number of RNAi-based therapies have advanced quickly into clinical trials. The clinical trials of siRNA-containing nanoparticles against cancer are included in Table 2.

Table 2: Clinical trials – siRNA- containing nanoparticles against cancer (PKN3 – Protein Kinase N3, SNALP - Stable Nucleic Acid Lipid Particle)

SiRNA drug	Nanocarriers	Target gene	Disease
Atu027	Cationic lipoplex	PKN3	Pancreatic ductal carcinoma
siG12D LODER	Polymer	KRAS G12D	Pancreatic ductal carcinoma
TKM-PLK1	SNALP	PCSK9	Hepatocellular carcinoma
SiRNA-EphA2-DOPC	Neutral liposome	EphA2	Advanced cancers

With regards to effective in vivo siRNA delivery, while much progress has been made in the last couple years, many obstacles still remain to be addressed to optimize the formulation with respect to safety, effectiveness, and selectivity. The development of siRNA still faces many issues due to the disposition of nanocarrier-mediated siRNA still having several issues as to the range of applicability: debatable immunogenicity and toxicity. While nanocarriers are arguably immunogenic when compared to viral vectors, they still face and are required to bypass several barriers associated with the immune system (e.g. immune clearance, Endothelial Tissue Adhesion and Retention, Organ Stasis, etc.) which ultimately leads to the restriction of the efficient delivery of drug and consequently the obstruction of their clinical use. If in vivo immunogenicity, cytotoxicity, and off-targeted immune activation are to be addressed, future research will be focused on these areas. The development of more advanced, biocompatible, and fully biodegradable controlled drug delivery systems is warranted for the more advanced use of RNAi-based cancer therapy.

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CONFLICT OF INTEREST

The authors have no conflict of interest.

REFERENCES

1. Mishra, D. K., Balekar, N., & Mishra, P. K. (2017). Nanoengineered strategies for siRNA delivery: from target assessment to cancer therapeutic efficacy. *Drug delivery and translational research*, 7(2), 346-358.
2. <https://www.who.int/news/item/01-02-2024-global-cancer-burden-growing--amidst-mounting-need-for-services>.
3. Chen, M., Dong, C., & Shi, S. (2021). Nanoparticle-mediated siRNA delivery and multifunctional modification strategies for effective cancer therapy. *Advanced Materials Technologies*, 6(10), 2001236.
4. Isazadeh, H., Oruji, F., Shabani, S., Behrooz, J., Nasiri, H., Isazadeh, A., & Akbari, M. (2023). Advances in siRNA delivery approaches in cancer therapy: challenges and opportunities. *Molecular biology reports*, 50(11), 9529-9543.
5. Dana, H., Chalbatani, G. M., Mahmoodzadeh, H., Karimloo, R., Rezaiean, O., Moradzadeh, A., ... & Gharagouzlo, E. (2017). Molecular mechanisms and biological functions of siRNA. *International journal of biomedical science: IJBS*, 13(2), 48.
6. Ali, H. M., Urbinati, G., Raouane, M., & Massaad-Massade, L. (2012). Significance and applications of nanoparticles in siRNA delivery for cancer therapy. *Expert review of clinical pharmacology*, 5(4), 403-412.
7. Lee, J. M., Yoon, T. J., & Cho, Y. S. (2013). Recent developments in nanoparticle-based siRNA delivery for cancer therapy. *BioMed research international*, 2013(1), 782041.
8. Shen, H., Sun, T., & Ferrari, M. (2012). Nanovector delivery of siRNA for cancer therapy. *Cancer gene therapy*, 19(6), 367-373.
9. Li, J., Xue, S., & Mao, Z. W. (2016). Nanoparticle delivery systems for siRNA-based therapeutics. *Journal of Materials Chemistry B*, 4(41), 6620-6639.

10. Guo, J., O'Driscoll, C. M., Holmes, J. D., & Rahme, K. (2016). Bioconjugated gold nanoparticles enhance cellular uptake: A proof of concept study for siRNA delivery in prostate cancer cells. *International journal of pharmaceutics*, 509(1-2), 16-27
11. Ebrahimi, N., Manavi, M. S., Nazari, A., Momayezi, A., Faghihkhorsani, F., Abdulwahid, A. H. R. R., ... & Fang, W. (2023). Nano-scale delivery systems for siRNA delivery in cancer therapy: New era of gene therapy empowered by nanotechnology. *Environmental research*, 239, 117263.

Chapter 7:

From Skin Ally to Antimicrobial Arsenal: Exploring *Staphylococcus hominis*, Hominicin & Emerging Peptide Weapons

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Abstract

Research on new bioactive molecules has gained pace with the alarming development of antimicrobial resistance; bacteriocins and antimicrobial peptides (AMPs) have been promising candidates for replacing traditional antibiotics. This bipodal action is best exemplified by *Staphylococcus hominis* (*S. hominis*), a common human skin commensal causing bacteremia, septicemia, endocarditis, and endophthalmitis apart from inducing colonization resistance and immunity against infections. Appearance of methicillin-resistant *S. hominis* (MRSHo) underlines the urgency of alternative treatment approaches. Hominicin, a broad-spectrum and heat-stable AMP, is one of its antibacterial weapons and has proven strong activity against methicillin-resistant and vancomycin-intermediate *Staphylococcus aureus* (MRSA, VISA). More recently, a pore-forming daptide bacteriocin known as hominlysin has been recognized as central to skin colonization resistance. The thiopeptide bacteriocin micrococcin P1 (MP1), secreted by both *S. hominis* and other *streptococcal* species, has proven promising in vivo at reducing *S. aureus* infection and accelerating wound closure when delivered using nanoparticle-based technologies. Bacteriocin gene clusters (BGCs) continue to be revealed by genomic and in silico research and remind of this commensal species latent antibacterial richness. Peptide discovery and optimization are simultaneously being revolutionized by developments in computational biology, such as OmegAMP, a deep-learning generative algorithm for the development of targeted AMPs, and the HMD-AMP system of AMP annotation. These strategies collectively bring new possibilities of AMPs designed from *S. hominis* being exploited for drug applications. With a focus on homininicin and hominlysin, this review synthesizes the latest data on *S. hominis* being a producer of bacteriocins and AMPs. In addition to signaling the problems of resistance, clinical role, and new computational resources on AMP engineering, the paper culminates by proposing possible avenues of transforming these findings into novel antimicrobial strategies.

Keywords: Antimicrobial peptides (AMPs), Antimicrobial resistance, Bacteriocins, Hominicin, *Staphylococcus hominis*.

1. Introduction

The Antimicrobial resistance (AMR) is considered to pose the most significant threats to public health across the world, with deaths in a world where resistant infections are increasing alarmingly. In 2019, AMR was recognized as the third leading cause of mortality globally, highlighting the rapid decline of traditional antimicrobial therapeutics (Murray et al., 2022). The profound enrichment of resistant bacterial populations has posed an urgent need for alternative anti-infective strategies, which arises from the inappropriate use, overuse, and extended incorporation of antibiotics in both human and veterinary medicine (Cotter et al., 2013).

Some of the most promising alternatives are antimicrobial peptides (AMPs) and bacteriocins, which shows a variety of mechanisms of action, structural diversity and a relatively low capability for the development of resistance. AMPs are short, protein- like, often peptides which are stable at high temperatures and can suppress the microbial growth by targeting membranes which makes bacterial evasion difficult. Bacteriocins are another type of AMPs which are ribosomal synthesized and can inhibit closely related and phylogenetically distant species. These are highly selective that show potency at low concentrations and are stable at across a broad range of environmental conditions (Cotter et al., 2013). Ever since their discovery in 1925, they have been classified into various groups such as class I lantibiotics, which consists of post-translationally modified amino acids like lanthionine and also exhibit membrane active properties.

The skin microbiome in humans serves as a rich reservoir for organisms producing bacteriocins and AMPs. Coagulase-negative staphylococci (CoNS) are some of the major inhabitants of healthy skin, flourishing in a hostile environment having an acidic pH, fatty acids, dessication , ultraviolet radiation and endogeneous antimicrobial compounds (Byrd et al., 2018). These commensals play a crucial role in maintaining skin homeostasis by forming an epidermal barrier, moderating host-immune responses and releasing antimicrobial molecules that inhibits opportunistic pathogens through various mechanism collectively known as colonization resistance (Nakatsuji et al., 2013; Parlet et al., 2019; Libertucci et al., 2019; Bay et al., 2020).

Although, *Staphylococcus epidermidis* is the most thoroughly investigated CoNS species, its dual nature as a harmless commensal and a harmful pathogen is linked with infections related to implants, biofilm formation and inflammatory skin conditions like atopic dermatitis and netherton syndrome that restricts its therapeutic applications (Otto, 2008; Foster et al., 2013; Paharik et al., 2016; Williams et al., 2020; Severn et al., 2022). Comparatively, *S. hominis*, the second most common CoNS species found on human skin is a notably promising source for the secretion of antimicrobial substances with lower pathogenic risks. Particularly, *S. hominis* does

not proliferate in lesions related to atopic dermatitis and is found to be linked with protective effects against pathogenic colonization.

Several AMPs showing great potency are derived from *S. hominis* such as hominicin which is an anti-staphylococcal AMP that can withstand high temperatures and particularly active against methicillin-resistant and vancomycin-intermediate *S. aureus* (MRSA, VRSA) and hominlysin, a daptide that increases colonization resistance by forming pores. Additionally, the bacteria also produce a thiopeptide antibiotic, micrococcin P1 (MP1) which has a strong efficacy in topical and wound-healing. Further studies through genomic analyses had revealed that lanthipeptide and bacteriocin biosynthetic gene clusters had high prevalence in *S. hominis*, indicating that its antimicrobial repertoire is still incompletely explored (Fernández-Fernández et al., 2025).

Despite the promising antimicrobial capabilities of *S. hominis*, it also presents certain crucial clinical limitations. The rising prevalence of methicillin-resistant and multi-drug-resistant strains of *S. hominis* has been associated with various hospital acquired infections, like bacteremia, endocarditis, urinary tract infections and complications related to devices (Szemraj et al., 2025; Muraki et al., 2022), emphasizing its dual nature.

Recent advances in computational biology had led to the discovery of novel AMPs. Machine learning tools like HMD-AMP has enabled accurate multi-label annotation of AMP activity (Yu et al., 2021), and diffusion-based generative models like OmegaAMP has enhanced the deliberate design of peptides with controlled physiochemical and antimicrobial properties (Soares et al., 2025). This review combines current findings into *S. hominis* biology, AMP generation and resistance patterns, with specific focus on hominicin, hominlysin and micrococcin P1, and discussing how computational frameworks are shaping the development of next-generation anti-infectives.

2. Biology & Commensal Role of *Staphylococcus hominis*

2.1. Skin Commensal — Frequent Inhabitant of Human Skin Micro biota

Staphylococcus hominis is a leading coagulase-negative staphylococci (CoNS) colonizer of human skin. It has been frequently recovered from various cutaneous sites in both genome and culture-based microbiome studies, hinting that *S. hominis* is a stable, long-term resident of the cutaneous micro biota (Severn et al., 2022). The draft genome of strain Hudgins consists genes involved in the utilization of variety of carbohydrates and amino acids, probable capsule production, and type IV pili, all features that likely support colonization, adhesion, and survival on the skin surface. This genomic profile highlights metabolic adaptability of *S. hominis* and provides a plausible explanation for its persistence on the skin (Calkins et al., 2016).

This structural and metabolic plasticity observed suggests that *S. hominis* is well adjusted to thrive in the challenging cutaneous milieu, which comprehends microbial competition, acidity, dessication, and ultraviolet exposure. Correspondingly, *S. hominis* confirms ecological suitability as a persistent and prolific skin commensal with a broad distribution across individuals and anatomical locations.

2.2 Protective Mechanisms — Quorum Sensing Interference & Colonization Resistance

S. hominis is not merely a passive skin resident; it actively contributes to protecting us against opportunistic pathogens. In a seminal study, (Severn et al., 2022) demonstrated that six different autoinducing peptides produced by *S. hominis* are capable of effectively inhibiting the agr quorum sensing system in the skin pathogen *Staphylococcus aureus*—in addition to their effects on *S. epidermidis*. This inhibition reduces the expression of *S. aureus* virulence genes without inhibiting its growth; thus, this is a quorum-quenching rather than bactericidal activity.

Using mass spectrometry, the researchers revealed the structures of three new AIPs, worked with conditioned media from *S. hominis* cultures, and showed that synthetic versions of these AIPs inhibit all four known agr classes of *S. aureus*. Most primarily, synthetic AIP-II remarkably reduced MRSA-induced dermonecrotic lesions in a mouse skin injury model. These data reveal real in vivo protective potential for AIPs produced by *S. hominis* (Severn et al., 2022).

This quorum-quenching strategy fits within the colonization resistance concept: commensal microbes strengthen the host's barrier by simply dialing down the virulence of pathogens rather than abolishing them completely. The advantage is that there is less collateral damage, a preserved native microbiota balance, and potentially lower pressure for resistance.

This protective role further underlines the value of *S. hominis* as a symbiont, furthering a balance in the microbiome–host–pathogen interrelation rather than simply inhabiting the skin.

2.3 Genomic Insights — Metabolic Versatility, Virulence Determinants, and Antibiotic Resistance

Whole-genome sequencing has revealed that *S. hominis* strains harbor genetic characteristics with both beneficial and harmful capacity. The draft genome of the Hudgins strain consists of 2.2 million base pairs encoding more than 2,000 proteins, involving genes for the synthesis of the polysaccharide capsules (poly- γ -glutamate capsule), type IV pili, metabolic enzymes and various factors correlated with virulence (Calkins et al., 2016).

A more recent comparative genomic analysis of the MDR, methicillin-resistant *S. hominis* clinical isolate (ShoR14) expands the competence capacity of the species as a hospital-associated pathogen. The study has demonstrated that ShoR14 has an acquired mobile genetic element that

carries the virulence factors, antibiotic resistance genes, plasmids, prophages, and various genomic islands. Methicillin resistance was due to a new variant of the SCCmec type VIII cassette. It also consists genes for tetracycline, macrolides, chloramphenicol, and antiseptics, which may be plasmid-borne. (Al-Trad et al.,2022).

The genomic variability contributes to the clinical diversity as seen in *S. hominis* isolates, which can act as harmless commensals supporting cutaneous health as well as multidrug-resistant pathogens causing severe systemic infections. These studies highlight the necessity for continuous monitoring, genome-based strain identification and careful risk-benefit evaluation when taking into account molecules derived from *S. hominis* for therapeutic applications to prevent unintended promotion of antimicrobial resistance.

3. Hominicin: Structure, Stability & Antimicrobial Spectrum

3.1 Discovery & Characterization

During the bioactivity screening of commensal bacterial isolates, it was initially identified that a *Staphylococcus hominis* strain MBBL 2-9 produces a novel antimicrobial peptide, homininicin. Afterwards, using some sequential extraction techniques like, chloroform extraction, ion-exchange chromatography and reverse phase high-performance liquid chromatography (HPLC), the peptides were purified from the culture supernatant which yielded a biologically active fraction with potent antibacterial activity against clinically relevant pathogens (Kim et al., 2010). Various antimicrobial assays illustrated that homininicin exhibited strong inhibitory activities against methicillin-susceptible *Staphylococcus aureus* (ATCC 25923), some antibiotic-resistant strains including methicillin-resistant *S.aureus* (MRSA) (ATCC11435) and vancomycin-intermediate *S.aureus* (VISA) (CCARM 3501) (Kim et al., 2010). These results established homininicin as a peptide having significant activity against gram-positive clinical isolates, especially antibiotic-resistant ones.

3.2 Molecular Features

Hominicin has a molecular mass of approximately 2038.4 Da as revealed by mass spectrometry analysis, placing it within the range of low molecular weight bacteriocins (Kim et al., 2010). Using LC-MS and NMR spectroscopy detailed sequencing was performed which determined the primary structure of homininicin. This has shown that homininicin contain several uncommon amino acids, like dehydro residues (Dhb, dehydrobutyrine; Dmp, dehydromethylproline) along with some other modified residues that are characteristic of class I lantibiotics, but they don't consist the characteristic thioether bridges which are typically found in lantibiotic structures (Kim et al., 2010).

The peptide sequence of hominycin- Dmlle-Dhb-Pro-Ala-Dhb-Pro-Phe-Dhb-Pro-Ala-Ile-Thr-Glu-Ile-Dhb-Ala-Ala-Val-Ile-Ala-Dmp shows no significant similarity to previously reported antimicrobial peptides (AMP), clearly indicating that hominycin is a structurally unique among known AMPs (Kim et al., 2010). Hominycin can be represented as a distinct variant within the broad class of staphylococcal antibiotic like peptides due to the presence of dehydrated amino acids and post-translational modifications.

3.3. Robust stability

One of the most striking features of hominycin is its exceptional physiochemical stability, which significantly enhances its therapeutical potential. Thermal stability assays like exposure to high temperatures, including autoclave conditions as well as prolonged incubation at elevated temperatures showed that the peptide retains antimicrobial activity (Kim et al., 2010) and thus such heat tolerance acts as a consistent feature for many small, post-translationally modified bacteriocins that resist Denaturation and degradation.

Hominycin also displays a broad range of pH stability along with thermal resilience remaining active across a wide range of acidic to basic environments (from pH 2.0-10.0). This characteristic feature allows the peptide for various applications in different environmental conditions, such as skin surfaces, wound beds or digestive tract niches.

Despite this robustness, hominycin is found to be sensitive to proteinase K treatment which resulted in the peptide cleavage and complete loss of antimicrobial activity indicating that intact peptide structure is essential for function. Contrastingly, hominycin is resistant to several other proteases, including trypsin, pepsin and lipase, highlighting its structural resilience (Kim et al., 2010).

3.4. Antimicrobial spectrum

Gram positive bacteria especially, *Staphylococcus aureus* and its resistant strains are at the core of hominycin's antimicrobial spectrum. In vitro assays showed strong efficacy against:

Methicillin-susceptible *S. aureus* (MSSA)

Vancomycin-intermediate *S. aureus* (VISA) strains and

Methicillin-resistant *S. aureus* (MRSA) (Kim et al., 2010)

Related literature has shown that hominycin acts at low minimum inhibitory concentrations (MICs) against these strains in comparison to other staphylococcal bacteriocins (Newstead et al., 2020). Due to its small size, stability and resistance to common proteases hominycin is considered as a favourable candidate among other potential AMPs.

4. Other antimicrobial peptides from *Staphylococcus hominis*

4.1. Hominlysin – A novel daptide bacteriocin

In addition to hominycin, *Staphylococcus hominis* seems to produce other potent AMPs. A recent study had identified a novel daptide bacteriocin- hominlysin, which is encoded in a skin derived isolate. This peptide is relatively small (~21 amino acids) having a measured m/z ratio of ~1020. Based on the function aspect, hominlysin has displayed antimicrobial activity against *Staphylococcus* species by disrupting the membrane or pore formation in the target cells (Nguyen et al., 2025).

According to the biophysical tests like voltage-clamp lipid bilayer, it has been found that hominlysin creates channels in bacterial membranes rich in peptide, which results in the transmembrane dissipation and cell death. In addition to this, the hominlysin gene locus in *S. aureus* which is sufficient to confer antimicrobial activity is expressed heterologously, has proved that the peptide alone drives membrane targeting and killing in a foreign host. Importantly, purified hominlysin has reduced *S. aureus* burden, inflammation, and transepithelial water loss providing in vivo protection in a murine skin infection model. This highlights its potent efficacy as a therapeutic agent for skin infection against gram positive pathogens (Nguyen et al., 2025).

4.2. Micrococcin P1 and broader AMP landscape

4.2.1 Thiopeptide micrococcin P1 (MP1)

Micrococcin P1 (MP1), a thiopeptide bacteriocin is known for its potent efficacy against gram positive bacteria, including MRSA and other resistant pathogens (Ciufolini et al., 2010). This type of bacteriocins function primarily by inhibiting protein synthesis by binding to ribosomal components and thus impeding elongation (Ovchinnikov et al., 2021).

MP1 biosynthetic gene clusters occur in *S. hominis* and related CoNS variants as revealed by the genomic mining and comparative analyses of *Staphylococcus* genomes. MP1 clusters were identified along with lanthipeptide and other bacteriocin gene clusters through the *in-silico* screening of 22 commensal *Staphylococcus* genomes, indicating that *S. hominis* and other related species can act as natural repositories of various AMPs (Fernández-Fernández et al., 2025)

4.2.2 MP1 infection models and nanoparticle delivery

MP1 producing *S. hominis* strain had been shown to lower the *S. aureus* infections in vivo. This thiopeptide when applied as a “probiotic” topical approach in murine infection models has been found to significantly accelerate wound healing and reduced abscess formation as compared to controls (Liu et al., 2020).

Researchers have created certain nanoparticle delivery systems that encapsulate MP1 for increased efficacy and to overcome the physiochemical limitations associated with natural bacteriocins like hydrophobicity and poor solubility. For a instance, as compared to free MP1, MP1 loaded PEG-PCL naoparticles has reduced bacterial burden and inflammation by maintain antimicrobial efficacy and improving outcomes in both localized and systemic infections. By improving stability, bioavailability, and targeting infection sites these delivery strategies enhance the translational potential of bacteriocin-based therapeutics.

Table 1: Key Antimicrobial Peptides Produced by *Staphylococcus hominis*

Peptide	Approx. Size	Mechanism of Action	Target Pathogens	Key Features
Hominicin	~2038 Da	Membrane disruption	<i>S. aureus</i> , MRSA, VISA	Heat- and pH-stable
Hominlysin	~21 aa	Pore formation	Staphylococci	Rapid bactericidal action
Micrococcin P1	Thiopeptide	Ribosomal inhibition	<i>S. aureus</i>	Wound-healing potential

5. Computational Approaches for Antimicrobial Peptide Discovery

AMPs discovery and optimization has been accelerated due to the recent developments in computational biology and artificial intelligence. By facilitating high throughput technologies, functional annotations and logical refining of AMP candidates has reduced major drawbacks of conventional experiment workflows, which is particularly valuable in the light of enhancing antimicrobial resistance.

5.1. OmegaAMP: Controlled generative modeling of AMPs

With a precise control over physiochemical properties, antimicrobial activity profiles and species specificity, OmegaAMP is a deep learning-based model designed for targeted AMP generation. OmegaAMP creates peptides that closely mimic naturally occurring AMPs in terms of amino acid composition, structural diversity and biophysical limitations by integrating protein language model embeddings along with diffusion based generative modeling (Soares et al., 2025). OmegaAMP reduces reliance on post hoc filtering and reducing false positives by enabling direct conditioning during sequence generation as compared to previous generative-discriminative pipelines. This feature is particularly important for creating selective AMPs that preserves

commensal species like *Staphylococcus hominis* while targeting harmful bacteria like *Staphylococcus aureus*. As a result, this tool provides a logical groundwork for producing optimized variants of naturally occurring peptides, such as hominicin, with increased stability and antimicrobial efficiency.

5.2. HMD-AMP: Hierarchical multi-label annotation of AMPs

A hierarchical multi-label deep forest framework called as HMD-AMP was created to accurately annotate AMPs and their antimicrobial target spectra. By simultaneously predicting antimicrobial activity and target organism types such as gram positive and gram-negative bacteria, fungi and viruses, HMD-AMP leverages protein language model-derived embeddings to capture the multifunctional nature of AMPs. This method improves the robustness against overfitting and shuffled sequences while overcoming the drawbacks of binary AMP/non-AMP classification. In the context of *S. hominis* HMD-AMP supports prioritization of candidates for experimental validation by facilitating the functional annotation of bacteriocins and cryptic AMP gene clusters which are identified through genome mining.

5.3. Relevance to *Staphylococcus hominis*-derived AMPs

A powerful strategy for unlocking the antimicrobial potential of commensal bacteria is the integration of combined generative and annotation based computational approaches. In addition to experimental approaches on peptides like hominicin, hominlysin and micrococcin P1, platforms such as OmegaAMP and HMD-AMP has enabled for the systemic discovery and optimization of bacteriocins and AMPs. Together, these tools support the translation of *S.hominis*-derived peptides into next-generation anti-infective therapeutics and fasten the development of specific, micro biota-sparing antimicrobial agents.

6. Challenges and future perspectives

6.1. Translational challenges in *Staphylococcus hominis*-derived AMPs

Despite the increasing evidence highlighting the antimicrobial potential of peptides derived from *Staphylococcus hominis*, several drawbacks limit their applications in clinical settings. At the mechanistic and molecular levels, the lack of complete characterization of bacteriocin marks as the primary challenge. Even though peptides like hominicin, hominlysin and microccin P1 has shown efficient potency against multi-drug-resistant pathogens, the information regarding their pharmacokinetics data, immunogenicity, and long-term stability is limited. Additionally, the restricted antimicrobial range of some bacteriocins, although beneficial for preserving microbiota, might hinder their application in the form of standalone treatments. Further

complications in development pipelines can occur due to manufacturing scalability, formulation stability and vulnerability to proteolytic degradation.

6.2. Future directions and emerging opportunities

Recent advances in genomics, bioinformatics and artificial intelligence have offered promising approaches to address these challenges. By combining genome mining with computational annotation tools like HMD-AMP, it is observed that they can systematically uncover hidden AMP gene clusters in *S.hominis*. Additionally, generative model like OmegaAMP allows for the rational optimization of peptide sequences thereby increasing stability, selectivity and antimicrobial potency. These tools minimize cytotoxicity and resistance development while facilitating the development of next generation AMP variants which retains biological efficiency. Topical and localized uses, such as wound dressing, coatings for implants, delivery systems based on nanoparticles, has shown great potential towards AMPs derived from *S.hominis*, as they avoid certain issues related to systemic toxicity. Additionally, incorporating peptides from *S. hominis* in therapeutic strategies that spare microbiota is found to align well with precision medicine initiatives intended for the preservation of commensal ecosystems while specifically targeting pathogens like *S. aureus*. Future research should aim on in vivo validation, standardized safety profiling and comparative studies with traditional antibiotics.

Conclusion

Staphylococcus hominis holds a distinctive role at the intersection of microbiome studies, antimicrobial resistance and therapeutic approaches. Increasing evidence shows that this prevalent skin commensal produces a wide range of bacteriocins and AMPs including hominicin, hominlysin and micrococcin P1, exhibiting considerable efficacy against clinically significant and multi-drug-resistant pathogens. *S. hominis* highlights an underexploited yet highly promising source for generating new antimicrobials when combined with emerging computational discovery platforms. Utilizing its antimicrobial potential through integrated-experimental approaches can lead to innovative, targeted, resistance-resilient anti-infective treatments in the era following traditional antibiotics.

References

- Al-Trad, E. I., Che Hamzah, A. M., Puah, S. M., Chua, K. H., Kwong, S. M., Yeo, C. C., & Chew, C. H. (2022). Comparative Genomic Analysis of a Multidrug-Resistant *Staphylococcus hominis* ShoR14 Clinical Isolate from Terengganu, Malaysia, Led to the Discovery of Novel Mobile Genetic Elements. *Pathogens (Basel, Switzerland)*, 11(12), 1406. <https://doi.org/10.3390/pathogens11121406>
- Bay, L., Barnes, C. J., Fritz, B. G., Thorsen, J., Restrup, M. E. M., Rasmussen, L., Sørensen, J. K., Hesselvig, A. B., Odgaard, A., Hansen, A. J., & Bjarnsholt, T. (2020). Universal Dermal Microbiome in Human Skin. *mBio*, 11(1), e02945-19. <https://doi.org/10.1128/mBio.02945-19>
- Byrd, A. L., Belkaid, Y., & Segre, J. A. (2018). The human skin microbiome. *Nature reviews. Microbiology*, 16(3), 143–155. <https://doi.org/10.1038/nrmicro.2017.157>
- Calkins, S., Couger, M. B., Jackson, C., Zandler, J., Hudgins, G. C., Hanafy, R. A., Budd, C., French, D. P., Hoff, W. D., & Youssef, N. (2016). Draft genome sequence of *Staphylococcus hominis* strain Hudgins isolated from human skin implicates metabolic versatility and several virulence determinants. *Genomics data*, 10, 91–96. <https://doi.org/10.1016/j.gdata.2016.10.003>
- Ciufolini, M., & Lefranc, D. (2010). Micrococcin P1: Structure, biology and synthesis. *Natural Product Reports*, 27(3), 330-342.
- Cotter, P. D., Ross, R. P., & Hill, C. (2013). Bacteriocins - a viable alternative to antibiotics?. *Nature reviews. Microbiology*, 11(2), 95–105. <https://doi.org/10.1038/nrmicro2937>
- Fernández-Fernández, R., Elsherbini, A. M. A., Lozano, C., Martínez, A., de Toro, M., Zarazaga, M., Peschel, A., Krismer, B., & Torres, C. (2025). Genomic Analysis of Bacteriocin-Producing *Staphylococci*: High Prevalence of Lanthipeptides and the Micrococcin P1 Biosynthetic Gene Clusters. *Probiotics and antimicrobial proteins*, 17(1), 159–174. <https://doi.org/10.1007/s12602-023-10119-w>
- Foster, T. J., Geoghegan, J. A., Ganesh, V. K., & Höök, M. (2014). Adhesion, invasion and evasion: the many functions of the surface proteins of *Staphylococcus aureus*. *Nature reviews. Microbiology*, 12(1), 49–62. <https://doi.org/10.1038/nrmicro3161>
- Kim, P. I., Sohng, J. K., Sung, C., Joo, H. S., Kim, E. M., Yamaguchi, T., Park, D., & Kim, B. G. (2010). Characterization and structure identification of an antimicrobial peptide, hominisin, produced by *Staphylococcus hominis* MBBL 2-9. *Biochemical and biophysical research communications*, 399(2), 133–138. <https://doi.org/10.1016/j.bbrc.2010.07.024>

- Libertucci, J., & Young, V. B. (2019). The role of the microbiota in infectious diseases. *Nature microbiology*, 4(1), 35–45. <https://doi.org/10.1038/s41564-018-0278-4>
- Liu, Y., Liu, Y., Du, Z., Zhang, L., Chen, J., Shen, Z., Liu, Q., Qin, J., Lv, H., Wang, H., He, L., Liu, J., Huang, Q., Sun, Y., Otto, M., & Li, M. (2020). Skin microbiota analysis-inspired development of novel anti-infectives. *Microbiome*, 8(1), 85. <https://doi.org/10.1186/s40168-020-00866-1>
- Muraki, R., Morita, Y., Tatsuta, K., Ida, S., Kitajima, R., Hirotsu, A., Takeda, M., Kikuchi, H., Hiramatsu, Y., Fukazawa, A., Kuroda, G., Kakizawa, K., & Takeuchi, H. (2022). Refractory postoperative *Staphylococcus hominis* bacteremia in a patient with an ACTH-producing pancreatic neuroendocrine neoplasm: a case report. *Surgical case reports*, 8(1), 126. <https://doi.org/10.1186/s40792-022-01485-8>
- Nakatsuji T, Chiang H-I, Jiang SB, Nagarajan H, Zengler K, Gallo RL. 2013. The microbiome extends to subepidermal compartments of normal skin. *Nat Commun*4:1431. <https://doi.org/10.1038/ncomms2441>.
- Newstead, L. L., Varjonen, K., Nuttall, T., & Paterson, G. K. (2020). Staphylococcal-Produced Bacteriocins and Antimicrobial Peptides: Their Potential as Alternative Treatments for *Staphylococcus aureus* Infections. *Antibiotics (Basel, Switzerland)*, 9(2), 40. <https://doi.org/10.3390/antibiotics9020040>
- Nguyen, A., Bunch, Z. L., Martinez-Aldino, I. Y., Rangel-Grimaldo, M., Tak, U., Thorstenson, J. C., Severn, M. M., Nakatsuji, T., Gallo, R. L., Graf, T. N., Oberlies, N. H., Chekan, J. R., Cech, N. B., & Horswill, A. R. (2025). An antimicrobial daptide from human skin commensal *Staphylococcus hominis* protects against skin pathogens. *Nature communications*, 10.1038/s41467-025-66259-w. Advance online publication. <https://doi.org/10.1038/s41467-025-66259-w>
- Otto M. (2008). Staphylococcal biofilms. *Current topics in microbiology and immunology*, 322, 207–228. https://doi.org/10.1007/978-3-540-75418-3_10
- Ovchinnikov, K. V., Kranjec, C., Telke, A., Kjos, M., Thorstensen, T., Scherer, S., Carlsen, H., & Diep, D. B. (2021). A Strong Synergy Between the Thiopeptide Bacteriocin Micrococcin P1 and Rifampicin Against MRSA in a Murine Skin Infection Model. *Frontiers in immunology*, 12, 676534. <https://doi.org/10.3389/fimmu.2021.676534>
- Paharik, A. E., & Horswill, A. R. (2016). The Staphylococcal Biofilm: Adhesins, Regulation, and Host Response. *Microbiology spectrum*, 4(2), 10.1128/microbiolspec.VMBF-0022-2015. <https://doi.org/10.1128/microbiolspec.VMBF-0022-2015>

- Parlet, C. P., Brown, M. M., & Horswill, A. R. (2019). Commensal Staphylococci Influence *Staphylococcus aureus* Skin Colonization and Disease. *Trends in microbiology*, 27(6), 497–507. <https://doi.org/10.1016/j.tim.2019.01.008>
- Severn, M. M., Williams, M. R., Shahbandi, A., Bunch, Z. L., Lyon, L. M., Nguyen, A., Zaramela, L. S., Todd, D. A., Zengler, K., Cech, N. B., Gallo, R. L., & Horswill, A. R. (2022). The Ubiquitous Human Skin Commensal *Staphylococcus hominis* Protects against Opportunistic Pathogens. *mBio*, 13(3), e0093022. <https://doi.org/10.1128/mbio.00930-22>
- Soares, D., Hetzel, L., Szymczak, P., Torres, M.D., Sommer, J., de la Fuente-Nunez, C., Theis, F.J., Gunnemann, S., & Szczurek, E. (2025). OmegAMP: Targeted AMP Discovery through Biologically Informed Generation.
- Szemraj, M., Glajzner, P., Olszowiec, K. *et al.* The prevalence of multidrug resistance in *Staphylococcus hominis* isolated from clinical materials. *Sci Rep* **15**, 414 (2025). <https://doi.org/10.1038/s41598-024-84500-2>
- Williams, M. R., Cau, L., Wang, Y., Kaul, D., Sanford, J. A., Zaramela, L. S., Khalil, S., Butcher, A. M., Zengler, K., Horswill, A. R., Dupont, C. L., Hovnanian, A., & Gallo, R. L. (2020). Interplay of Staphylococcal and Host Proteases Promotes Skin Barrier Disruption in Netherton Syndrome. *Cell reports*, 30(9), 2923–2933.e7. <https://doi.org/10.1016/j.celrep.2020.02.021>
- Yu, Q., Dong, Z., Fan, X., Zong, L., & Li, Y. (2021). HMD-AMP: Protein Language-Powered Hierarchical Multi-label Deep Forest for Annotating Antimicrobial Peptides. *bioRxiv*.

Chapter 8:

HSP70–Tau Interactions in Alzheimer’s Disease: Mechanistic Insights and Therapeutic Implications

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Abstract

Alzheimer’s disease (AD) is the most common neurodegenerative disorder and is defined pathologically by the deposition of amyloid- β plaques and the formation of neurofibrillary tangles composed of hyperphosphorylated Tau protein. Among these hallmarks, Tau pathology shows the strongest association with neuronal dysfunction, synaptic loss, and cognitive decline, positioning Tau as a central driver of disease progression and a prime therapeutic target. Heat shock protein 70 (HSP70), a highly conserved molecular chaperone, is a key regulator of cellular proteostasis, coordinating protein folding, quality control, and degradation pathways. Increasing evidence indicates that HSP70 plays a pivotal yet multifaceted role in modulating Tau pathology in AD. Through direct interactions with Tau, HSP70 limits pathological aggregation, facilitates the removal of misfolded Tau species, and protects neurons from Tau-induced toxicity. These effects are mediated by HSP70-dependent mechanisms that promote ubiquitin–proteasome and lysosomal degradation pathways, as well as by its ability to regulate Tau phosphorylation and intracellular distribution—critical factors that influence Tau stability and toxicity. This review provides an integrated overview of the molecular interplay between HSP70 and Tau in Alzheimer’s disease, highlighting how HSP70-mediated proteostatic control shapes Tau homeostasis. Furthermore, we discuss emerging therapeutic strategies aimed at harnessing HSP70 function, including pharmacological activation, genetic modulation, and targeting of co-chaperone networks, as promising approaches to prevent, slow, or reverse Tau-driven neurodegeneration.

Keywords: Alzheimer’s disease, Tauopathy, Heat Shock Protein 70, Proteostasis, Molecular chaperones

1. Introduction

Neurodegenerative diseases represent a growing global health crisis, with Alzheimer's disease (AD) being the most prevalent form of dementia (Long & Holtzman, 2019). The neuropathological hallmarks of AD are well-established: extracellular deposits of amyloid-beta ($A\beta$) peptides forming senile plaques, and intracellular aggregates of the microtubule-associated protein Tau, which form neurofibrillary tangles (NFTs) (Iqbal et al., 2016). While the "amyloid cascade hypothesis" has long dominated AD research, accumulating evidence suggests that Tau pathology correlates more closely with the degree of cognitive decline observed in patients (Ballatore et al., 2007). In a healthy neuron, Tau's primary function is to bind to and stabilize microtubules, the cellular "highways" essential for axonal transport and structural integrity. However, in pathological conditions, Tau becomes abnormally hyperphosphorylated, detaches from microtubules, and begins to self-aggregate into toxic oligomers and, eventually, insoluble paired helical filaments (PHFs) and straight filaments (SFs) that constitute NFTs (Guo et al., 2017).

The process of Tau aggregation is not merely a downstream consequence of disease; it is an active driver of neurotoxicity. The formation of soluble Tau oligomers, precursors to the larger NFTs, is now considered to be the most synaptotoxic species, disrupting neuronal function long before large tangles are formed (Lasagna-Reeves et al., 2012). This makes the prevention of Tau aggregation a primary therapeutic goal. The cell, in turn, is not without its own defenses. It possesses a sophisticated quality control system, headlined by molecular chaperones, which function to maintain protein homeostasis (proteostasis). This review focuses on the 70-kilodalton heat shock protein (HSP70) family, a central component of the cellular chaperone network. We will discuss the structural biology of Tau, the mechanisms driving its aggregation, and the specific, powerful role that HSP70 plays in counteracting this pathological cascade, as detailed in recent comprehensive reviews (e.g., Zeng et al., 2020). By understanding this critical interaction, we can better identify novel therapeutic avenues for AD and related Tauopathies.

2. The Tau Protein: From Function to Dysfunction

Tau is an intrinsically disordered protein (IDP), meaning it lacks a stable, well-defined three-dimensional structure in its native state (Zeng et al., 2020). This structural plasticity allows it to perform its diverse functions, primarily the regulation of microtubule dynamics. The Tau protein is encoded by the MAPT gene, and alternative splicing results in six different isoforms in the adult human brain. These isoforms differ by the inclusion of either three or four

microtubule-binding repeats (MTBRs), denoted as 3R and 4R Tau, respectively. The MTBRs are the core functional region of the protein, but they also contain two hexapeptide motifs, 275VQIINK280 (PHF6*) and 306VQIVYK311 (PHF6), which are highly prone to forming the β -sheet structures that initiate aggregation (von Bergen et al., 2000). Under normal physiological conditions, these aggregation-prone regions are thought to be shielded. However, post-translational modifications (PTMs), particularly hyperphosphorylation, can induce a conformational change in the Tau monomer, exposing these motifs and triggering the aggregation process.

The transition from a soluble, functional protein to an insoluble, toxic aggregate is a multi-step process. It begins with the formation of seed-competent Tau monomers, which have adopted a specific, pathological conformation (Mirbaha et al., 2018). These monomers then assemble into small, soluble oligomers, which are highly neurotoxic. These oligomers can further elongate, eventually forming the large, insoluble fibrils that make up NFTs. This process can be modulated by a variety of factors, including mutations in the MAPT gene (which cause frontotemporal dementia), interactions with polyanionic molecules like heparin, and, critically, the cellular chaperone machinery.

3. Molecular Chaperones and the HSP70 Family

The cellular proteostasis network is a complex system of chaperones and degradation machinery (e.g., the ubiquitin-proteasome system) that ensures proteins are correctly folded, localized, and cleared when damaged. Heat shock proteins are a major class of molecular chaperones, and their expression is often upregulated in response to cellular stress, such as heat, oxidative stress, or the accumulation of misfolded proteins. The HSP70 family is one of the most important chaperone families, playing a central role in a wide range of cellular processes, including de novo protein folding, protein trafficking, and the prevention of protein aggregation.

HSP70 chaperones function through an ATP-dependent cycle of binding and releasing substrate proteins. They recognize and bind to short, exposed hydrophobic segments in unfolded or misfolded proteins, which are normally buried in the protein's core. This binding prevents the substrates from aggregating and can, in concert with co-chaperones like HSP40 and HSP110, facilitate their refolding or triage them for degradation.

4. HSP70 as a Direct Inhibitor of Tau Aggregation

The connection between HSP70 and Tau pathology is not merely circumstantial; there is strong biochemical evidence demonstrating a direct, inhibitory interaction. Studies have shown that HSP70 can potently suppress the aggregation of Tau in vitro and in cellular models. This inhibition occurs at multiple stages of the aggregation cascade, making HSP70 a particularly effective guardian of Tau proteostasis.

First and foremost, HSP70 inhibits the initial, rate-limiting step of aggregation, known as nucleation. It has been shown to suppress the formation of Tau nuclei, thereby preventing the entire downstream cascade of fibril formation (Kundel et al., 2018). This is a crucial intervention point, as it stops the problem before it starts. The mechanism for this appears to involve HSP70 binding directly to the aggregation-prone regions of the Tau monomer, particularly the MTBRs containing the PHF6* and PHF6 motifs (Mok et al., 2018). By binding to these regions, HSP70 effectively "shields" them, preventing them from interacting with other Tau molecules to form the β -sheet structures that seed aggregation.

In addition to inhibiting nucleation, HSP70 also interferes with the elongation of existing Tau fibrils. Even if some aggregates do form, HSP70 can slow their growth. Furthermore, it has been demonstrated that HSP70 can sequester pre-formed Tau aggregates with a remarkably high affinity (Kundel et al., 2018). This suggests that HSP70 can not only prevent the formation of new aggregates but also "disarm" existing ones, potentially reducing their toxicity and preventing their propagation to neighbouring cells. HSP70 blocks the early stages of Tau aggregation by suppressing the formation of Tau nuclei.

5. The HSP70/HSP90 Chaperone Axis

The fate of HSP70-bound Tau is further decided by a "triage" decision involving another major chaperone, HSP90. While HSP70 acts as a primary holder and folder, HSP90 is more involved in the maturation and activation of specific client proteins. In the context of Tau, the interaction between these two chaperones is critical. It has been proposed that a dynamic exchange exists: when Tau is bound by HSP70, it is held in a non-aggregating state. If Tau is then transferred to HSP90, it can be directed towards degradation via the proteasome. Analysis of the Tau-associated proteome has revealed that the exchange of HSP70 for HSP90 is a key step involved in Tau degradation (Thompson et al., 2012). Therefore, the balance between HSP70 and HSP90 activity can determine whether a misfolded Tau protein is refolded, held in a benign

state, or eliminated entirely. Dysregulation of this balance could lead to the accumulation of Tau and contribute to disease progression.

6. Therapeutic Implications and Future Directions

The profound ability of HSP70 to inhibit Tau aggregation makes it an attractive therapeutic target for AD and other Tauopathies. Strategies to enhance the cell's natural defenses against Tau pathology could provide a novel approach to treatment. Several potential avenues are being explored. One approach is to develop small-molecule drugs that can upregulate the expression of HSP70 and other beneficial chaperones. This would boost the cell's intrinsic capacity to handle misfolded Tau.

Another strategy involves modulating the activity of the chaperones themselves. For instance, inhibitors of HSP90 have been shown to increase the levels of HSP70-bound Tau, thereby shifting the balance towards Tau clearance. This approach has shown promise in pre-clinical models. Furthermore, understanding the precise binding sites of HSP70 on the Tau protein could allow for the design of small molecules or peptides that mimic HSP70's inhibitory action, directly shielding the aggregation-prone regions of Tau.

It is crucial, however, that future therapeutic design takes into account the complexity of the Tau aggregation process and the heterogeneity of Tau structures in different diseases (Scheres et al., 2020). The cryo-EM structures of Tau filaments from different diseases have revealed distinct folds, suggesting that the aggregation process is disease-specific. A successful therapeutic will likely need to be tailored to the specific type of Tau pathology being targeted. Table 1 provides an integrated review of HSP70–Tau interactions across the Tau aggregation cascade.

Table 1: Different stages of Tau pathology and involvement of HSP70

Stage of Tau Pathology	Molecular Events in Tau	HSP70 Action	Cellular Outcome
Native Tau (Physiological state)	Tau binds microtubules and maintains axonal transport	Assists in proper folding and stabilization	Normal neuronal function
Tau misfolding initiation	Conformational change exposes PHF6/PHF6* motifs	Recognizes exposed hydrophobic regions	Prevents aberrant intermolecular interactions
Nucleation (early aggregation)	Formation of seed-competent Tau monomers	Directly inhibits nucleation	Blocks initiation of aggregation cascade

Oligomer formation	Soluble Tau oligomers accumulate	Sequesters oligomeric Tau	Reduces synaptotoxic species
Fibril elongation	Growth of paired helical and straight filaments	Suppresses fibril elongation	Limits neurofibrillary tangle burden
Pre-formed aggregates	Stable Tau fibrils persist	Binds and neutralizes aggregates	Prevents propagation and toxicity
Protein quality control	Misfolded Tau requires clearance	Facilitates ubiquitination and proteasomal degradation	Enhanced Tau turnover
Chaperone triage	Decision between refolding and degradation	Coordinates with HSP90	Directs Tau fate appropriately
Disease modification	Progressive Tau accumulation	HSP70 induction/modulation	Therapeutic attenuation of Tau pathology

Conclusion

The aggregation of the Tau protein is a central toxic event in the pathogenesis of Alzheimer's disease. The cellular machinery for maintaining protein homeostasis, particularly the molecular chaperone HSP70, plays a critical and direct role in counteracting this process. Through its ability to bind to aggregation-prone regions of the Tau monomer, inhibit fibril nucleation and elongation, and sequester toxic aggregates, HSP70 acts as a key defender of neuronal health. The intricate interplay between HSP70 and other chaperones like HSP90 further orchestrates the fate of misfolded Tau, steering it towards degradation. The powerful anti-aggregation activity of HSP70 highlights the therapeutic potential of modulating the cellular chaperone system. Enhancing these natural defences offers a promising strategy to halt the progression of Tau pathology and ultimately combat the devastating effects of Alzheimer's disease.

References

- Ballatore, C., Lee, V. M. Y., & Trojanowski, J. Q. (2007). Tau-mediated neurodegeneration in Alzheimer's disease and related disorders. *Nature Reviews Neuroscience*, 8(9), 663–672. <https://doi.org/10.1038/nrn2194>
- Guo, T., Noble, W., & Hanger, D. P. (2017). Roles of tau protein in health and disease. *Acta Neuropathologica*, 133(5), 665–704. <https://doi.org/10.1007/s00401-017-1707-9>

- Iqbal, K., Liu, F., & Gong, C. X. (2016). Tau and neurodegenerative disease: The story so far. *Nature Reviews Neurology*, 12(1), 15–27. <https://doi.org/10.1038/nrneurol.2015.225>
- Kundel, F., De, S., Flagmeier, P., Horrocks, M. H., Kjaergaard, M., Shammash, S. L., Jackson, S. E., Dobson, C. M., & Knowles, T. P. J. (2018). Hsp70 inhibits the nucleation and elongation of tau and sequesters tau aggregates with high affinity. *ACS Chemical Biology*, 13(3), 636–646. <https://doi.org/10.1021/acscchembio.7b01039>
- Lasagna-Reeves, C. A., Castillo-Carranza, D. L., Sengupta, U., Sarmiento, J., Troncoso, J., Jackson, G. R., & Kaye, R. (2012). Identification of oligomers at early stages of tau aggregation in Alzheimer's disease. *The FASEB Journal*, 26(5), 1946–1959. <https://doi.org/10.1096/fj.11-199851>
- Long, J. M., & Holtzman, D. M. (2019). Alzheimer disease: An update on pathobiology and treatment strategies. *Cell*, 179(2), 312–339. <https://doi.org/10.1016/j.cell.2019.09.001>
- Mirbaha, H., Chen, D., Morozova, O. A., Ruff, K. M., Sharma, A. M., Liu, X., Goodarzi, M., Pappu, R. V., & Diamond, M. I. (2018). Inert and seed-competent tau monomers suggest structural origins of aggregation. *eLife*, 7, e36584. <https://doi.org/10.7554/eLife.36584>
- Mok, S. A., Condello, C., Freilich, R., Gillies, A., Arhar, T., Oroz, J., Kadavath, H., Julien, O., Assimon, V. A., Rauch, J. N., Dunyak, B. M., Lee, J., Tsai, F. T. F., Wilson, M. R., Zweckstetter, M., Dickey, C. A., & Gestwicki, J. E. (2018). Mapping interactions with the chaperone network reveals factors that protect against tau aggregation. *Nature Structural & Molecular Biology*, 25(5), 384–393. <https://doi.org/10.1038/s41594-018-0071-z>
- Scheres, S. H. W., Zhang, W., Falcon, B., & Goedert, M. (2020). Cryo-EM structures of tau filaments. *Current Opinion in Structural Biology*, 64, 17–25. <https://doi.org/10.1016/j.sbi.2020.05.007>
- Thompson, A. D., Scaglione, K. M., Prensner, J., Gillies, A. T., Chinnaiyan, A., Paulson, H. L., Jinwal, U. K., & Dickey, C. A. (2012). Analysis of the tau-associated proteome reveals that exchange of Hsp70 for Hsp90 is involved in tau degradation. *ACS Chemical Biology*, 7(9), 1677–1686. <https://doi.org/10.1021/cb3002696>
- von Bergen, M., Friedhoff, P., Biernat, J., Heberle, J., Mandelkow, E.-M., & Mandelkow, E. (2000). Assembly of tau protein into Alzheimer paired helical filaments depends on a local sequence motif (306VQIVYK311) forming β structure. *Proceedings of the National Academy of Sciences of the United States of America*, 97(10), 5129–5134. <https://doi.org/10.1073/pnas.97.10.5129>
- Zeng, Y., Yang, J., Zhang, B., Gao, M., Su, Z., & Huang, Y. (2020). The structure and phase of tau: From monomer to amyloid filament. *Cellular and Molecular Life Sciences*, 77, 1–14. <https://doi.org/10.1007/s00018-020-03589-x>

Chapter 9:

Multidrug-Resistant Nanodrugs: Applications, Mechanisms, and Future Perspectives

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Abstract

Multidrug resistance (MDR) has emerged as one of the most critical challenges in modern medicine, severely limiting the effectiveness of conventional therapeutics in both infectious diseases and cancer. The rapid evolution of resistant pathogens and tumor cells has necessitated the development of innovative treatment strategies capable of overcoming biological, genetic, and physiological barriers to drug efficacy. Nanotechnology-based drug delivery systems, commonly referred to as nanodrugs, have gained considerable attention as a promising approach to counter multidrug resistance. Nanodrugs offer unique physicochemical properties, including nanoscale size, large surface area-to-volume ratio, tunable surface chemistry, and the ability to encapsulate or conjugate multiple therapeutic agents. These features enable enhanced drug solubility, targeted delivery, controlled release, and improved intracellular accumulation, thereby addressing key mechanisms of MDR such as efflux pump overexpression, reduced drug uptake, enzymatic drug degradation, and microenvironment-mediated protection. This review provides a comprehensive and critical analysis of multidrug-resistant nanodrugs, focusing on their mechanisms of action, design strategies, and applications in cancer and infectious diseases. Various classes of nanocarriers, including polymeric nanoparticles, liposomes, dendrimers, metallic nanoparticles, and lipid-based systems, are discussed in detail. Furthermore, current challenges, safety considerations, clinical translation status, and future perspectives of MDR-targeted nanodrugs are highlighted. This review aims to provide an in-depth resource for researchers and clinicians working toward the development of next-generation therapeutics to combat multidrug resistance.

Keywords: Multidrug resistance, nanodrugs, nanotechnology, targeted drug delivery, cancer therapy, antimicrobial resistance

1. Introduction

The emergence and spread of multidrug resistance (MDR) represent a major global health crisis affecting the treatment of infectious diseases and cancer. MDR is broadly defined as the ability of cells or microorganisms to resist the effects of multiple structurally and functionally unrelated drugs. In clinical settings, MDR leads to treatment failure, prolonged illness, increased mortality, and escalating healthcare costs. The World Health Organization has identified antimicrobial resistance as one of the top threats to global health, while drug resistance in cancer remains a leading cause of chemotherapy failure.

Conventional drug development strategies have struggled to keep pace with the rapid evolution of resistance mechanisms. In infectious diseases, pathogens acquire resistance through genetic mutations and horizontal gene transfer, whereas cancer cells develop resistance via complex molecular and cellular adaptations. These challenges have driven interest in alternative therapeutic strategies that go beyond traditional small-molecule drugs. Nanotechnology has emerged as a transformative platform capable of addressing many of the limitations associated with conventional therapies.

Nanodrugs are defined as therapeutic or diagnostic agents that utilize nanoscale materials, typically ranging from 1 to 100 nanometers, for improved drug delivery and efficacy. By leveraging their unique properties, nanodrugs can enhance drug bioavailability, protect active agents from degradation, facilitate targeted delivery, and modulate pharmacokinetics. Importantly, nanodrugs offer novel opportunities to overcome MDR by bypassing efflux pumps, co-delivering multiple drugs, and altering drug–cell interactions. This review focuses on the role of nanodrugs in overcoming multidrug resistance, with particular emphasis on their applications, mechanisms, and translational potential.

2. Multidrug Resistance: An Overview

2.1 Definition and Classification

Multidrug resistance refers to the phenomenon in which cells exhibit resistance to multiple therapeutic agents, often with different chemical structures and mechanisms of action. MDR can be classified into intrinsic resistance, which is naturally present in certain organisms or cell types, and acquired resistance, which develops in response to drug exposure. Cross-resistance occurs when resistance to one drug confers resistance to related drugs, whereas co-resistance involves independent resistance mechanisms acting simultaneously.

2.2 Mechanisms of Multidrug Resistance

MDR arises through a variety of interconnected mechanisms. One of the most well-characterized mechanisms is the overexpression of drug efflux pumps, such as ATP-binding cassette (ABC) transporters, which actively expel drugs from cells. Other mechanisms include drug target modification, enzymatic drug inactivation, reduced drug uptake due to membrane alterations, and sequestration of drugs within intracellular compartments. In cancer, additional factors such as tumor heterogeneity, hypoxia, and the tumor microenvironment play significant roles in MDR development.

2.3 Clinical Implications

The clinical impact of MDR is profound, leading to reduced therapeutic options and poor patient outcomes. In oncology, MDR is responsible for relapse and metastasis following chemotherapy. In infectious diseases, MDR pathogens complicate treatment regimens and increase the risk of outbreaks. These challenges underscore the urgent need for innovative therapeutic strategies, including nanodrug-based approaches.

3. Nanotechnology and Nanodrugs: Principles and Advantages

Nanotechnology involves the manipulation of materials at the nanoscale to achieve novel properties and functions. Nanodrugs exploit these properties to enhance drug delivery and therapeutic efficacy. The small size of nanoparticles allows them to interact with biological systems at the molecular level, facilitating cellular uptake and intracellular delivery.

Key advantages of nanodrugs include improved solubility of poorly water-soluble drugs, protection of labile drugs from degradation, prolonged circulation time, and targeted delivery to specific tissues or cells. Surface functionalization with ligands, antibodies, or peptides enables active targeting, while passive targeting can be achieved through enhanced permeability and retention effects in tumors or infected tissues.

4. Nanocarriers for Overcoming Multidrug Resistance

4.1 Polymeric Nanoparticles

Polymeric nanoparticles are among the most extensively studied nanocarriers for MDR applications. They are typically composed of biodegradable polymers such as poly(lactic-co-glycolic acid), chitosan, and polyethylene glycol. These systems can encapsulate hydrophobic and hydrophilic drugs and provide controlled release profiles. Polymeric nanoparticles have been shown to bypass efflux pumps and enhance intracellular drug accumulation.

4.2 Liposomes

Liposomes are spherical vesicles composed of phospholipid bilayers that can encapsulate both aqueous and lipophilic drugs. Liposomal formulations have demonstrated improved pharmacokinetics and reduced toxicity. In MDR contexts, liposomes can shield drugs from efflux pumps and deliver them directly to target cells. Several liposomal nanodrugs have been approved for clinical use, highlighting their translational potential.

4.3 Dendrimers

Dendrimers are highly branched, tree-like macromolecules with well-defined structures and multiple functional groups. Their unique architecture allows precise control over size, shape, and surface chemistry. Dendrimers can be engineered to co-deliver drugs and resistance modulators, making them attractive candidates for MDR therapy.

4.4 Metallic and Inorganic Nanoparticles

Metallic nanoparticles, such as gold and silver nanoparticles, exhibit unique optical, electronic, and antimicrobial properties. These nanoparticles can serve as drug carriers or therapeutic agents themselves. Silver nanoparticles, in particular, have shown efficacy against multidrug-resistant bacteria through multiple mechanisms, reducing the likelihood of resistance development.

4.5 Lipid-Based Nanocarriers

Solid lipid nanoparticles and nanostructured lipid carriers combine the advantages of liposomes and polymeric nanoparticles. They offer high drug loading capacity, stability, and biocompatibility. Lipid-based nanocarriers have been widely explored for delivering anticancer and antimicrobial agents to overcome MDR.

5. Applications of Multidrug-Resistant Nanodrugs in Cancer

Nanodrugs have demonstrated significant potential in overcoming MDR in cancer therapy. By enhancing drug accumulation within tumor cells and modulating resistance pathways, nanodrugs can restore chemosensitivity. Combination nanotherapies that co-deliver chemotherapeutic agents and gene-silencing molecules targeting resistance genes have shown promising results in preclinical studies.

Targeted nanodrugs can exploit tumor-specific markers to achieve selective delivery, minimizing off-target effects. Furthermore, stimuli-responsive nanocarriers that release drugs in response to pH, temperature, or enzymatic activity offer additional control over drug delivery in resistant tumors.

6. Applications of Multidrug-Resistant Nanodrugs in Infectious Diseases

In the context of infectious diseases, nanodrugs offer novel strategies to combat MDR pathogens. Nanoparticles can enhance antibiotic penetration into biofilms, protect antibiotics from enzymatic degradation, and facilitate intracellular delivery to infected cells. Metallic nanoparticles and antimicrobial peptide-loaded nanocarriers have shown broad-spectrum activity against resistant bacteria, fungi, and viruses.

Nanotechnology-based vaccines and diagnostic tools further contribute to the management of MDR by enabling early detection and prevention of resistant infections.

7. Safety, Toxicity, and Regulatory Considerations

Despite their potential, nanodrugs pose unique safety and regulatory challenges. Nanoparticle toxicity depends on factors such as size, composition, surface charge, and dose. Comprehensive preclinical evaluation is essential to assess biocompatibility, biodistribution, and long-term effects. Regulatory frameworks for nanomedicines are still evolving, necessitating standardized guidelines for evaluation and approval.

8. Clinical Translation and Current Status

Several nanodrug formulations have advanced to clinical trials or received regulatory approval, particularly in oncology.

Multidrug Resistance versus Nanodrug Resistance: Comparison and Similarities

While nanodrugs are widely investigated as solutions to overcome multidrug resistance, it is increasingly recognized that resistance phenomena may also emerge against nanodrug-based systems. Understanding the similarities and differences between conventional multidrug resistance and nanodrug resistance is essential for the rational design of durable nanomedicine strategies.

Conceptual Differences

Multidrug resistance traditionally refers to resistance against small-molecule drugs with defined molecular targets, whereas nanodrug resistance involves reduced responsiveness to nanoscale drug delivery systems or nanoparticle-mediated therapies. MDR is often drug-centric, while nanodrug resistance is system-centric, involving interactions between nanocarriers, biological barriers, and host defense mechanisms.

Mechanistic Comparison

Conventional MDR is primarily driven by cellular and molecular mechanisms such as efflux pump overexpression, enzymatic drug degradation, target modification, and altered membrane

permeability. In contrast, nanodrug resistance may arise from enhanced nanoparticle clearance by the mononuclear phagocyte system, formation of protein coronas that alter targeting efficiency, limited tumor or tissue penetration, intracellular sequestration in endo-lysosomal compartments, and adaptive cellular responses that reduce nanoparticle uptake.

Despite these differences, overlap exists between the two phenomena. For example, efflux transporters that expel free drugs may be bypassed by nanocarriers, but adaptive cellular changes can still limit intracellular drug release or promote nanoparticle exocytosis. Similarly, biofilms and tumor microenvironments can impede both free drugs and nanodrugs through physical and biochemical barriers.

Biological Barriers Involved

In MDR, resistance mechanisms are often intrinsic to the target cells, such as bacteria or cancer cells. Nanodrug resistance, however, frequently involves host-related barriers, including immune recognition, opsonization, nonspecific uptake by macrophages, and rapid clearance from systemic circulation. These host-mediated factors distinguish nanodrug resistance from classical MDR while also highlighting the complexity of nanotherapeutic interactions in vivo.

Similarities between MDR and Nanodrug Resistance

Both MDR and nanodrug resistance share several important similarities. First, both are adaptive responses to therapeutic pressure, driven by biological systems seeking survival. Second, heterogeneity within tumors or microbial populations contributes to partial resistance in both cases. Third, suboptimal dosing and prolonged exposure can promote resistance development for both free drugs and nanodrug formulations. Finally, both phenomena necessitate combination strategies, such as co-delivery of therapeutics, targeting ligands, or resistance modulators, to achieve sustained efficacy.

Comparative Summary

Feature	Multidrug Resistance (MDR)	Nanodrug Resistance
Primary target	Free small-molecule drugs	Nanoparticles/nanocarriers
Key mechanisms	Efflux pumps, target mutation, drug inactivation	Immune clearance, protein corona, poor penetration
Biological level	Cellular and genetic	Cellular, systemic, and host-mediated
Role of microenvironment	High (biofilms, tumor stroma)	High (protein corona, immune system)

Clinical impact	Treatment failure, relapse	Reduced nanotherapy efficacy
Mitigation strategies	Drug combinations, new agents	Surface modification, stealth design, stimuli-responsive systems

Implications for Nanodrug Design

Recognizing the parallels and distinctions between MDR and nanodrug resistance has important implications for future nanomedicine development. Rational nanodrug design must account not only for classical resistance mechanisms but also for host–nanoparticle interactions, dynamic biological environments, and long-term adaptive responses. Integrating these considerations will be critical to prevent resistance against next-generation nanodrugs and to maximize their therapeutic durability.

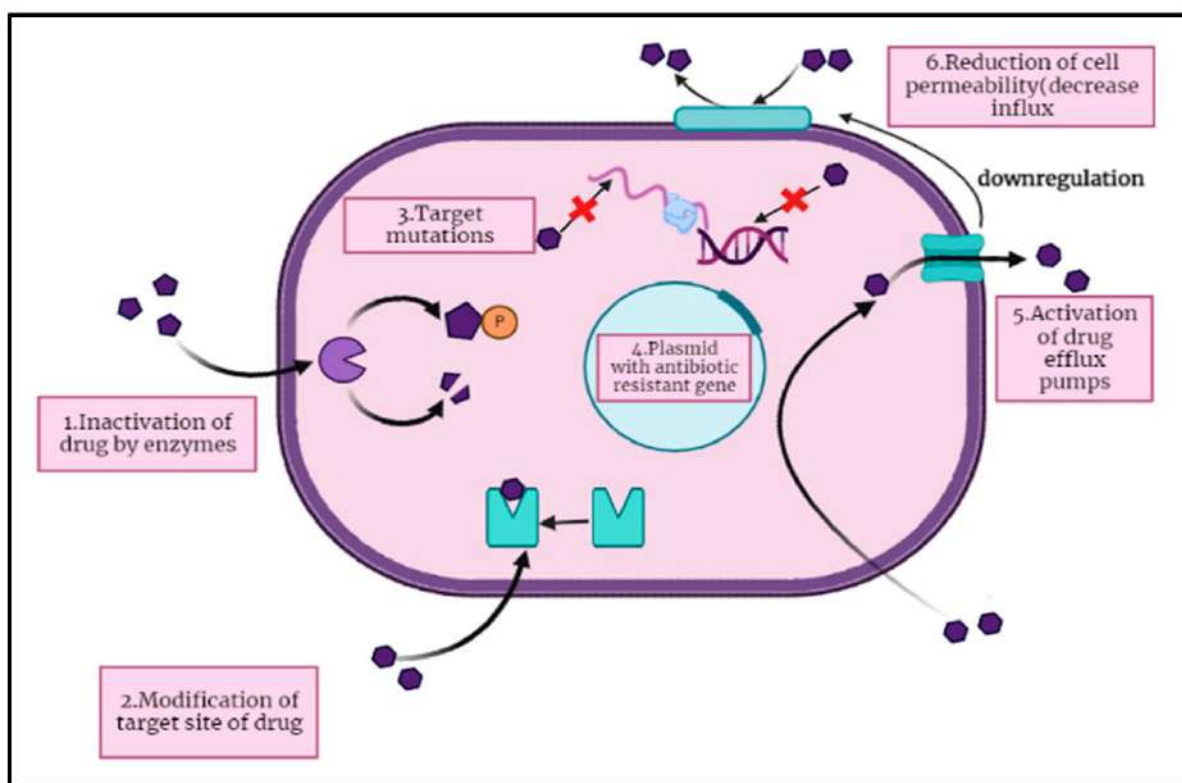
9. Nanotechnology and Nanodrugs: Principles and Advantages

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Core Mechanisms of Multidrug Resistance-

Across both microbiology and oncology, MDR typically involves four primary biochemical strategies:

- **Efflux Pumps:** Specialized transport proteins (e.g., ABC transporters, P-glycoprotein) actively pump drugs out of the cell, keeping intracellular concentrations below effective levels.
- **Reduced Permeability:** Mutations in porin genes or alterations in the cell membrane/envelope restrict drug entry.
- **Drug Inactivation:** The production of enzymes (e.g., β -lactamases) that chemically degrade or modify the drug through hydrolysis, phosphorylation, or acetylation.
- **Target Modification:** The cell alters its own internal components (e.g., ribosomes or binding proteins) so the drug can no longer recognize or bind to them.



References

1. Rahman, M. U., Shah, J. A., Khan, M. N., Bilal, H., Zhu, D., Du, Z., & Mu, D. S. (2025). Innovative Approaches to Combat Antimicrobial Resistance: A Review of Emerging Therapies and Technologies. *Probiotics and Antimicrobial Proteins*, 1-18.
2. Aher, S., Pathan, A., Jain, P., & colleagues. (2024). Footprints of nanocarrier on multidrug resistance therapy. *Nano and Medical Materials*, 4(1), 274. <https://doi.org/10.59400/nmm.v4i1.274>
3. Ahmed, N., Abusalah, M. A. H., & Abuarqoub, A. (2025). Nanomedicine in the fight against multidrug-resistant infections: A review on emerging strategies and translational prospects. *International Journal of Nanomedicine*, 20, 12331–12362.
4. Chaturvedi, A., & Ranjan, R. (2025). Antimicrobial nanoparticles: A new horizon to combat multidrug-resistant bacteria. *Frontiers in Nanotechnology*, 7, 1611126. <https://doi.org/10.3389/fnano.2025.1611126>
5. Ding, F., Songkiatissak, P., Cherukuri, P. K., & Zhang, Q. (2018). Antibiotic drug nanocarriers for probing of multidrug ABC membrane transporter of *Bacillus subtilis*. *ACS Omega*, 3(1), 1231–1243. <https://doi.org/10.1021/acsomega.9b03698>
6. Li, M., Liu, Y., Gong, Y., Zhang, X., & Chen, L. (2023). Recent advances in nanoantibiotics against multidrug-resistant bacteria. *Nanoscale Advances*, 5, 6278–6317. <https://doi.org/10.1039/D3NA00530E>
7. Rahman, M. U., Shah, J. A., Khan, M. N., Bilal, H., Zhu, D., Du, Z., & Mu, D. S. (2025). Innovative approaches to combat antimicrobial resistance: A review of emerging therapies and technologies. *Probiotics and Antimicrobial Proteins*, 1–18. <https://doi.org/10.1007/s12602-025-10125-7>
8. Yadav, P., Ambudkar, S. V., & Prasad, N. R. (2022). Emerging nanotechnology-based therapeutics to combat multidrug-resistant cancer. *Journal of Nanobiotechnology*, 20, 423. <https://doi.org/10.1186/s12951-022-01626-z>
9. Zhang, Y., Liu, X., Wang, J., & Chen, Z. (2024). Progress in research of nanotherapeutics for overcoming multidrug resistance in cancer. *International Journal of Molecular Sciences*, 25(18), 9973. <https://doi.org/10.3390/ijms25189973>
10. Allen, T. M., & Cullis, P. R. (2013). Liposomal drug delivery systems: From concept to clinical applications. *Advanced Drug Delivery Reviews*, 65(1), 36–48. <https://doi.org/10.1016/j.addr.2012.09.037>
11. Alfaro, K. O., Stock, C. M., Taylor, S., et al. (2015). Resistance to cancer chemotherapy: Failure in drug response from ADME to P-glycoprotein. *Cancer Cell International*, 15, 71. <https://doi.org/10.1186/s12935-015-0221-1>
12. Blanco, E., Shen, H., & Ferrari, M. (2015). Principles of nanoparticle design for overcoming biological barriers to drug delivery. *Nature Biotechnology*, 33(9), 941–951. <https://doi.org/10.1038/nbt.3330>
13. Bozzuto, G., & Molinari, A. (2015). Liposomes as nanomedical devices. *International Journal of Nanomedicine*, 10, 975–999.
14. Brigger, I., Dubernet, C., & Couvreur, P. (2012). Nanoparticles in cancer therapy and diagnosis. *Advanced Drug Delivery Reviews*, 64, 24–36.
15. Choi, Y. H., & Han, H. K. (2018). Nanomedicines: Current status and future perspectives in cancer therapy. *Journal of Pharmaceutical Investigation*, 48, 43–60.
16. Duncan, R., & Gaspar, R. (2011). Nanomedicine(s) under the microscope. *Molecular Pharmaceutics*, 8(6), 2101–2141.

17. Fan, W., Yung, B., Huang, P., & Chen, X. (2017). Nanotechnology for multimodal synergistic cancer therapy. *Chemical Reviews*, 117(22), 13566–13638.
18. Farokhzad, O. C., & Langer, R. (2009). Impact of nanotechnology on drug delivery. *ACS Nano*, 3(1), 16–20.
19. Gao, W., Thamphiwatana, S., Angsantikul, P., & Zhang, L. (2014). Nanoparticle approaches against bacterial infections. *Wiley Interdisciplinary Reviews: Nanomedicine and Nanobiotechnology*, 6(6), 532–547.
20. Gottesman, M. M., Fojo, T., & Bates, S. E. (2002). Multidrug resistance in cancer: Role of ATP-dependent transporters. *Nature Reviews Cancer*, 2(1), 48–58.
21. Gupta, S. C., Kannappan, R., Reuter, S., Kim, J. H., & Aggarwal, B. B. (2011). Chemosensitization of tumors by resveratrol. *Cancer Research*, 71(2), 538–549.
22. Hare, J. I., Lammers, T., Ashford, M. B., *et al.* (2017). Challenges and strategies in anti-cancer nanomedicine development. *Advanced Drug Delivery Reviews*, 108, 25–38.
23. Jain, R. K., & Stylianopoulos, T. (2010). Delivering nanomedicine to solid tumors. *Nature Reviews Clinical Oncology*, 7, 653–664.
24. Kalepu, S., & Nekkanti, V. (2015). Insoluble drug delivery strategies. *Acta Pharmaceutica Sinica B*, 5(5), 442–453.
25. Kwon, G. S., & Kataoka, K. (2012). Block copolymer micelles as long-circulating drug vehicles. *Advanced Drug Delivery Reviews*, 64, 237–245.
26. Li, X., Xu, H., & Dai, J. (2018). Nanocarriers for enhanced intracellular delivery of chemotherapeutics. *Drug Resistance Updates*, 37, 1–17.
27. Maeda, H., Wu, J., Sawa, T., Matsumura, Y., & Hori, K. (2000). Tumor vascular permeability and the EPR effect. *Journal of Controlled Release*, 65(1–2), 271–284.
28. Meng, H., Liong, M., Xia, T., *et al.* (2010). Engineered design of mesoporous silica nanoparticles. *ACS Nano*, 4(8), 4539–4550.
29. Moghimi, S. M., Hunter, A. C., & Murray, J. C. (2001). Long-circulating and target-specific nanoparticles. *Pharmacological Reviews*, 53(2), 283–318.
30. Nikaido, H. (2009). Multidrug resistance in bacteria. *Annual Review of Biochemistry*, 78, 119–146.
31. Peer, D., Karp, J. M., Hong, S., *et al.* (2007). Nanocarriers as an emerging platform for cancer therapy. *Nature Nanotechnology*, 2, 751–760.
32. Piddock, L. J. V. (2006). Multidrug-resistance efflux pumps. *Clinical Microbiology Reviews*, 19(2), 382–402.
33. Prabhu, S., & Poulouse, E. K. (2012). Silver nanoparticles: Mechanism of antimicrobial action. *International Nano Letters*, 2, 32.
34. Ramasamy, T., Ruttala, H. B., *et al.* (2017). Multidrug resistance cancer: Role of nanotechnology. *Journal of Controlled Release*, 258, 226–253.
35. Rizvi, S. A. A., & Saleh, A. M. (2018). Applications of nanoparticle systems in drug delivery technology. *Saudi Pharmaceutical Journal*, 26(1), 64–70.
36. Saeed, M., Kadioglu, O., Khalid, H., *et al.* (2020). Novel strategies to overcome MDR in cancer. *Drug Resistance Updates*, 51, 100709.

37. Shi, J., Kantoff, P. W., Wooster, R., & Farokhzad, O. C. (2017). Cancer nanomedicine: Progress and promise. *Nature Reviews Cancer*, 17, 20–37.
38. Torchilin, V. P. (2011). Tumor delivery of macromolecular drugs based on the EPR effect. *Advanced Drug Delivery Reviews*, 63(3), 131–135.
39. Tran, S., DeGiovanni, P. J., Piel, B., & Rai, P. (2017). Cancer nanomedicine: A review of recent success. *Journal of Nanoscience and Nanotechnology*, 17(1), 1–13.
40. Wang, J., Wang, H., *et al.* (2019). Nanoparticle-based drug delivery systems for cancer therapy. *Journal of Controlled Release*, 301, 1–18.
41. Wang, L., Hu, C., & Shao, L. (2017). The antimicrobial activity of nanoparticles. *International Journal of Nanomedicine*, 12, 1227–1249.
42. Wu, L., Zhang, J., & Watanabe, W. (2011). Physical and chemical stability of drug nanoparticles. *Advanced Drug Delivery Reviews*, 63(6), 456–469.
43. Xie, X., Liao, J., Shao, X., Li, Q., & Lin, Y. (2017). The effect of shape on cellular uptake of gold nanoparticles. *Biomaterials*, 135, 36–47.
44. Yang, H., Chen, Y., Chen, Z., *et al.* (2019). Nanoparticle delivery systems to overcome cancer MDR. *Acta Pharmaceutica Sinica B*, 9(4), 654–669.
45. Zhang, L., Pornpattananangkul, D., Hu, C. M. J., & Huang, C. M. (2010). Development of nanoparticles for antimicrobial drug delivery. *Current Medicinal Chemistry*, 17(6), 585–594.
46. Zhang, Y., Wang, F., *et al.* (2018). Nanomedicine-based strategies for MDR reversal. *Biomaterials Science*, 6, 176–191.
47. Zhao, M., Lei, C., Yang, Y., *et al.* (2018). Nanotechnology for overcoming cancer drug resistance. *Advanced Drug Delivery Reviews*, 143, 17–34.
48. Zhou, Y., Peng, Z., Seven, E. S., & Leblanc, R. M. (2018). Crossing the BBB with nanoparticles. *Journal of Controlled Release*, 270, 290–303.
49. Zhu, Y., Matsumura, Y., & Maeda, H. (2021). Nanomedicine in cancer therapy. *Advanced Therapeutics*, 4, 2000200.

Chapter 10:

Stem cells as targeted drug delivery systems for precision therapy

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Abstract

The efficacy of targeted drug delivery is fundamentally constrained by biological complexity, including heterogeneous disease microenvironments, dynamic signalling landscapes and protective physiological barriers that limit the performance of conventional therapeutic carriers. Stem cell-based drug delivery systems introduce a qualitatively different strategy by exploiting the inherent biological intelligence of living cells to achieve adaptive, site-specific therapy. Through endogenous chemotactic sensing, active migration and context-dependent phenotypic modulation, stem cells function as autonomous delivery agents capable of navigating pathological tissues with a precision unattainable by synthetic platforms alone. Recent convergence of stem cell biology with synthetic biology, nanotechnology and genome engineering has further transformed these systems into programmable therapeutic entities, enabling controlled cargo loading, stimuli-responsive release and integrated safety mechanisms. Across cancer, neurological, cardiovascular and inflammatory diseases, stem cell mediated delivery enables localized, sustained and microenvironment-responsive therapeutic action, addressing key limitations such as poor tissue penetration, systemic toxicity and therapeutic resistance. Importantly, the therapeutic impact of these platforms arises not only from delivered molecules but also from synergistic interactions between stem cell intrinsic immunomodulatory and regenerative functions and engineered therapeutic outputs. This dual functionality represents a conceptual shift from passive drug transport towards dynamic, self-regulating treatment modalities. By integrating biological targeting with engineered control, stem cell-based delivery systems redefine precision medicine as an adaptive process rather than a static intervention. This framework establishes a foundation for next-generation therapeutics capable of responding to disease evolution in real time, while highlighting critical design principles required to translate living delivery systems into clinically robust and controllable medical technologies. Collectively, this review underscores the potential of stem cell-based delivery systems to enable precise, adaptive and durable therapeutics, while outlining the scientific and regulatory advances required for their successful clinical translation.

Keywords: Cell-Based Therapeutics, Extracellular Vesicles, Precision Medicine, Stem Cells, Targeted Drug Delivery.

Introduction

The ability to deliver therapeutic agents selectively to diseased tissues while sparing healthy organs remains a fundamental challenge in modern medicine [1-2]. Despite major advances in drug discovery and formulation, the clinical efficacy of many therapeutics is compromised by suboptimal bio-distribution, rapid systemic clearance and off-target toxicity [1]. These limitations are particularly evident in complex diseases such as cancer, neurodegenerative disorders and chronic inflammatory conditions, where pathological sites are spatially heterogeneous and protected by biological barriers [2-3]. As a result, there is a growing need for innovative delivery strategies that can navigate the body's microenvironments with high precision and adaptability.

Targeted drug delivery systems have traditionally relied on synthetic carriers, including liposomes, polymeric nanoparticles and antibody-drug conjugates [4-5]. While these platforms have demonstrated improved pharmacokinetics and in some cases enhanced tissue specificity, they often face biological constraints such as immune recognition, limited tissue penetration and insufficient responsiveness to dynamic disease states [5-6]. Living cells, by contrast, possess intrinsic sensing, migratory and adaptive capabilities that cannot be easily replicated by engineered materials alone [7]. This realization has spurred increasing interest in cell-based drug delivery systems, with stem cells emerging as particularly attractive candidates [7-8].

Stem cells are defined by their capacity for self-renewal and differentiation, but their therapeutic potential extends well beyond tissue regeneration [9]. Various stem cell populations including mesenchymal stem cells (MSCs), neural stem cells (NSCs) and induced pluripotent stem cell (iPSC) derived progenitors exhibit a pronounced tropism toward sites of injury, inflammation and tumour growth [8-10]. This homing behaviour is mediated by complex interactions between chemokines, cytokines, adhesion molecules and extracellular matrix components within diseased microenvironments [10-11]. Importantly, these same pathological cues often define the regions where therapeutic intervention is most urgently needed, positioning stem cells as natural vectors for targeted drug delivery.

Over the past decade, substantial progress has been made in exploiting stem cells as carriers for a wide range of therapeutic agents [7,12]. Stem cells have been engineered or loaded to deliver small-molecule drugs, proteins, nucleic acids, nanoparticles and oncolytic viruses, either through passive internalization, surface modification or genetic manipulation [12-14]. Once administered, these cells can migrate toward pathological sites and release their cargo in-situ,

achieving localized therapeutic concentrations that are difficult to attain through systemic administration. In oncology, for example, stem cell based delivery has shown promise in targeting infiltrative tumour regions and metastatic niches that are poorly accessible to conventional therapies [14-15]. Similarly, in neurological diseases, stem cells offer a potential means of crossing the blood-brain barrier and delivering therapeutics directly to affected neural tissues.

Beyond passive delivery, advances in synthetic biology and biomaterials have enabled the design of stem cells with programmable functions [16]. Engineered stem cells can be equipped with inducible gene circuits, stimuli-responsive release systems or safety switches that allow external or micro-environmental control over therapeutic activity [16-17]. These developments transform stem cells from simple carriers into dynamic, responsive therapeutic platforms capable of adapting to disease progression. At the same time, growing interest in stem cell derived extracellular vesicles and exosomes has opened cell free alternatives that retain many targeting advantages while potentially reducing safety concerns [18].

Despite these advances, significant challenges remain before stem cell based drug delivery systems can be widely translated into the clinics [19]. Concerns related to tumorigenicity, immunogenicity, long-term engraftment and phenotypic stability must be carefully addressed. In addition, issues of scalable manufacturing, standardized characterization and regulatory oversight pose substantial hurdles for clinical implementation [19-20]. A nuanced understanding of stem cell-host interactions, as well as rigorous evaluation of safety and efficacy, will be essential for the successful development of these technologies.

In this article, we provide a comprehensive overview of stem cells as targeted drug delivery systems, focusing on the biological principles underlying their targeting capabilities, current strategies for therapeutic cargo loading and release and emerging approaches to enhance safety and translational potential. By integrating insights from stem cell biology, drug delivery and bioengineering, we aim to highlight both the opportunities and the challenges of this rapidly evolving field and to outline future directions toward precision, cell-based therapeutics.

1. Stem Cells as Natural Vehicles for Targeted Delivery

The success of stem cells as targeted drug delivery systems is fundamentally rooted in their innate biological behaviours, which evolved to support tissue repair and homeostasis. Unlike inert carriers, stem cells dynamically respond to biochemical and biomechanical cues within pathological microenvironments. Injury, inflammation, hypoxia and tumorigenesis generate

gradients of chemokines, cytokines, growth factors and extracellular matrix (ECM) components that actively recruit stem cells. Among these, the CXCL12-CXCR4 signaling axis is one of the most extensively characterized and plays a central role in directing stem cell migration toward ischemic tissues and tumor niches. Additional pathways, including CCL2-CCR2, HGF-c-Met and VEGF-VEGFR signaling, further contribute to tissue-specific homing.

Following chemotactic guidance, stem cells undergo firm adhesion to activated endothelium mediated by integrins (such as $\alpha 4\beta 1$ and $\alpha V\beta 3$), selectins and intercellular adhesion molecules. This enables trans-endothelial migration and infiltration into diseased tissues, a process further facilitated by stem cell secreted matrix metalloproteinases that remodel the ECM. Importantly, these mechanisms closely mirror those employed by immune cells, allowing stem cells to access sites that are otherwise poorly penetrated by synthetic nanoparticles or macromolecular drugs.

In parallel, stem cells exhibit a remarkable capacity to adapt phenotypically to local microenvironments. Exposure to inflammatory cytokines or hypoxic conditions can alter their secretome, migratory behaviour and survival, reinforcing their role as context sensitive delivery vehicles. These adaptive responses allow stem cells to persist in hostile disease settings and enhance local therapeutic action.

2. Stem Cell Types and Their Unique Advantages

2.1. Stem Cell-Host Immune Interactions

Immune evasion and immunomodulation are critical determinants of in-vivo stem cell persistence and delivery efficacy. MSCs, in particular, display low expression of major histocompatibility complex (MHC) class II molecules and co-stimulatory factors, reducing recognition by host immune cells. Additionally, MSCs secrete immunoregulatory mediators such as prostaglandin E2, indoleamine 2,3-dioxygenase, nitric oxide and transforming growth factor- β , which suppress T cell activation and promote regulatory immune phenotypes [10,11].

This immunomodulatory capacity confers dual benefits for drug delivery. First, it prolongs the circulation time and tissue residence of stem cells, increasing the likelihood of effective cargo delivery. Second, it can synergize with delivered therapeutics, particularly in inflammatory and autoimmune diseases, where immune dysregulation underlies pathology. However, immune interactions can also be context-dependent and in certain tumor types, immunosuppression may inadvertently support tumor progression. Understanding and modulating these effects is therefore essential for therapeutic optimization [21-22].

2.2 Mesenchymal Stem Cells (MSCs):

MSCs are isolated from bone marrow, adipose tissue, umbilical cord and other sources. Their low immunogenicity and ability to modulate inflammation make them ideal carriers for chronic inflammatory diseases, autoimmune disorders and tumors. MSCs have been used to deliver chemotherapeutics (e.g. paclitaxel), nucleic acids (siRNA, miRNA) and nanoparticles in preclinical cancer models, demonstrating efficient tumor targeting and reduced systemic toxicity [12,23].

2.3 Neural Stem Cells (NSCs):

NSCs are specialized for targeting central nervous system (CNS) lesions. Their capacity to traverse the blood-brain barrier and migrate to ischemic or neurodegenerative sites makes them promising for brain tumors, stroke and neurodegenerative disorders. NSCs can deliver anti-tumor agents such as oncolytic viruses, neuroprotective proteins or gene-editing constructs directly to affected neural tissue [15].

2.4 Induced Pluripotent Stem Cell (iPSC)-Derived Progenitors:

iPSCs provide patient-specific, autologous stem cells, reducing immunogenicity risks. They can be differentiated into diverse progenitor types and engineered for specific therapeutic applications. iPSC-derived MSCs or NSCs have been used to deliver cytokines, chemotherapeutics and regenerative factors, combining personalized therapy with precise tissue targeting [24-25].

3. Strategies for Therapeutic Molecule Loading

Stem cells can carry a wide array of therapeutic cargoes, including small molecules, proteins, nucleic acids, nanoparticles and viruses. The choice of loading strategy influences cargo stability, release kinetics and targeting efficiency [12-13, 16-18].

3.1 Passive Uptake:

Cells can internalize drugs or nanoparticles through endocytosis. While simple, this method may result in limited stability and uncontrolled release. Optimization of particle size, surface chemistry and incubation conditions can improve loading efficiency.

3.2 Surface Conjugation:

Therapeutics can be chemically or biologically conjugated to stem cell surfaces. This method allows precise control over the amount of cargo per cell and can exploit cell-surface interactions for localized delivery. However, excessive surface loading may impede cell migration or homing.

3.3 Genetic Engineering:

Genetic modification has emerged as a powerful strategy to transform stem cells into active therapeutic factories. Viral and non-viral gene delivery systems have been used to engineer stem cells to express anti-tumor cytokines, pro-apoptotic proteins, immune checkpoint inhibitors or enzymes that locally convert pro-drugs into active compounds. Inducible promoters and synthetic gene circuits enable tight spatiotemporal control over gene expression, minimizing off-target effects.

Recent advances in genome editing technologies, particularly CRISPR-Cas systems, have enabled precise and stable genetic modifications while reducing the risk of insertional mutagenesis. These tools also allow multiplexed engineering, whereby stem cells can simultaneously sense disease markers, deliver therapeutic cargo and activate safety mechanisms.

3.4 Non-Genetic Engineering:

To avoid permanent genetic alterations, non-genetic approaches have been developed. These include loading stem cells with drug-encapsulated nanoparticles, liposomes or polymeric micelles. Nanocarriers can be designed to release their cargo in response to specific stimuli such as acidic pH, reactive oxygen species or protease activity, which are hallmarks of many disease microenvironments. Surface engineering techniques further enhance targeting specificity. By decorating stem cell membranes with ligands or antibodies, it is possible to augment adhesion to diseased tissues or specific cell types. Importantly, such modifications must preserve cell viability, migratory capacity, and differentiation potential.

3.5 Extracellular Vesicle (EV) and Exosome-Based Delivery:

Stem cells naturally release EVs containing nucleic acids, proteins and lipids. These vesicles can be harvested and used as cell-free delivery platforms, combining targeting specificity with reduced safety risks associated with live-cell therapies. EVs can cross biological barriers, such as the blood-brain barrier, making them particularly valuable in neurological applications.

4. Controlled and Stimuli-Responsive Therapeutic Release

Recent advances in synthetic biology and biomaterials have enabled stem cells to function as programmable therapeutic platforms [16-17,19].

4.1 Inducible Systems:

Gene circuits responsive to small molecules, light or disease-specific signals allow temporal control of therapeutic release. For example, hypoxia-responsive promoters can trigger anti-cancer cytokine release in tumor cores.

4.2 Nanoparticle Integration:

Stem cells can transport nanoparticles designed to release drugs in response to pH, enzymes, or temperature. This approach ensures spatially and temporally precise delivery of small molecules or nucleic acids.

4.3 Safety Switches:

Suicide genes, inducible apoptosis constructs, or drug-responsive kill switches can terminate stem cells post-delivery to mitigate tumorigenicity or unwanted proliferation.

5. Applications Across Diseases

5.1 Cancer:

Cancer represents the most extensively investigated and arguably the most compelling application of stem cell based targeted drug delivery systems. Tumours are characterized by profound spatial and temporal heterogeneity, comprising hypoxic cores, invasive margins, stromal compartments and disseminated metastatic niches. These features severely limit the effectiveness of conventional systemic therapies, which often fail to achieve sufficient drug concentrations at critical tumour sites while inducing significant off-target toxicity. Stem cells, by virtue of their intrinsic tumour-tropic properties, offer a unique solution to these challenges by actively homing to and infiltrating malignant tissues.

The preferential migration of stem cells toward tumours is driven by inflammatory and stress-related signals produced by the tumour microenvironment. Malignant tissues secrete a wide array of chemokines, cytokines and growth factors, including CXCL12, IL-8, VEGF and HGF, which recruit stem cells through corresponding receptors such as CXCR4, CXCR1/2 and c-Met. Hypoxia, a hallmark of solid tumours, further amplifies stem cell recruitment by upregulating

hypoxia-inducible factor dependent signaling pathways. Once localized, stem cells can penetrate deeply into tumour parenchyma, including poorly vascularized regions that are largely inaccessible to nanoparticles or antibody-based therapeutics. Importantly, stem cells also interact extensively with tumour-associated stromal and immune cells, allowing them to navigate the complex tumour architecture. This enables delivery not only to primary tumour masses but also to invasive fronts and perivascular niches that contribute to tumour progression and therapeutic resistance.

Stem cells have also been widely explored as carriers for conventional chemotherapeutic drugs, such as paclitaxel, doxorubicin and cisplatin. By loading these agents into stem cells or stem cell associated nanoparticles, it is possible to achieve sustained, localized drug release within tumours while significantly reducing systemic exposure. Preclinical studies have demonstrated enhanced tumour regression and reduced cardiotoxicity or myelosuppression compared to free drug administration. This approach is particularly attractive for highly toxic agents with narrow therapeutic windows. In addition, stem cell-mediated delivery can overcome multidrug resistance mechanisms (MDR). By releasing cytotoxic agents directly into the tumour microenvironment (TME), stem cells can bypass efflux pumps and enhance intracellular drug accumulation in cancer cells [8,14,23].

5.2 Neurological Disorders:

Neurodegenerative disorders such as Parkinson's disease, Alzheimer's disease, amyotrophic lateral sclerosis and Huntington's disease are characterized by progressive neuronal loss, chronic inflammation and widespread synaptic dysfunction. Stem cell based delivery strategies in these contexts focus primarily on neuroprotection, modulation of neuroinflammation and delivery of gene therapies rather than direct cytotoxicity.

Stem cells have been engineered to deliver neurotrophic factors (e.g. brain-derived neurotrophic factor, glial cell line derived neurotrophic factor), anti-inflammatory cytokines, and gene-silencing constructs that target pathogenic proteins. These approaches aim to stabilize neuronal networks, reduce inflammatory damage and slow disease progression. The sustained and localized delivery enabled by stem cells is particularly advantageous in chronic neurodegenerative conditions requiring long-term therapeutic support.

Brain tumours, particularly glioblastoma, are among the most aggressive and treatment-resistant malignancies. Their infiltrative nature and diffuse dissemination throughout the brain severely limit surgical resection and localized therapies. NSCs (neural stem cells) have demonstrated

robust tropism toward glioma cells, migrating extensively to invasive tumour fronts and satellite lesions. This property has been exploited to deliver oncolytic viruses, chemotherapeutic agents and prodrug-converting enzymes directly to tumour sites. Stem cell mediated delivery enables sustained local release of therapeutic agents, enhances viral spread within tumour tissue and reduces off-target neurotoxicity. Importantly, NSCs can target residual tumour cells following surgical resection, addressing a major cause of tumour recurrence [8,14,23].

5.3 Cardiovascular disease:

Cardiovascular and ischemic diseases, including myocardial infarction, ischemic stroke and peripheral artery disease, remain leading causes of morbidity and mortality worldwide. These conditions are characterized by acute or chronic reductions in blood supply, resulting in hypoxia, inflammation, cell death and adverse tissue remodeling. Despite advances in pharmacological and interventional therapies, effective delivery of therapeutics to ischemic tissues remains a significant challenge due to compromised vasculature and limited tissue perfusion. Stem cell based targeted drug delivery systems provide a promising approach to address these limitations by exploiting ischemia-induced homing signals and the regenerative microenvironment. Stem cells have been extensively engineered to deliver angiogenic factors such as vascular endothelial growth factor (VEGF), fibroblast growth factor (FGF) and hepatocyte growth factor (HGF) to ischemic tissues. Localized delivery of these factors promotes neovascularization, enhances tissue perfusion and supports the survival of resident cells. Compared with systemic administration, stem cell mediated delivery reduces off-target angiogenesis and improves therapeutic efficacy. In addition to angiogenesis, stem cells have been used to deliver cytoprotective and anti-apoptotic agents that mitigate ischemia-induced cell death. These include anti-inflammatory cytokines, antioxidants and gene therapies targeting apoptotic pathways. Such approaches are particularly effective in the ischemic penumbra, where cells are functionally impaired but still viable [8,14,23]

5.4 Inflammatory and Autoimmune Disorders:

Chronic inflammatory and autoimmune disorders, such as rheumatoid arthritis, inflammatory bowel disease, multiple sclerosis and systemic lupus erythematosus, are characterized by persistent immune dysregulation and tissue damage. Conventional therapies often rely on systemic immunosuppression, which can lead to significant adverse effects and increased susceptibility to infections. Stem cell based targeted drug delivery offers an alternative approach by enabling localized immunomodulation at sites of inflammation.

Inflamed tissues produce high levels of chemokines, cytokines and adhesion molecules that recruit stem cells. MSCs express receptors such as CCR2, CXCR4 and ICAM-1, allowing them to home efficiently to inflamed joints, intestinal mucosa or central nervous system lesions. Once localized, MSCs interact with immune cells, including T cells, B cells, dendritic cells and macrophages, modulating their activation and differentiation. This immunomodulatory capacity is mediated by both direct cell-cell interactions and the secretion of soluble factors such as IL-10, TGF- β , prostaglandin E2 and indoleamine 2,3-dioxygenase. These mechanisms allow stem cells to suppress pathological immune responses while preserving protective immunity [8,14,23].

6. Translational Considerations for Stem Cell Based Targeted Drug Delivery Systems

Despite compelling preclinical evidence supporting the use of stem cells as targeted drug delivery systems, successful clinical translation remains limited and uneven across disease indications. The transition from experimental models to clinical application requires careful consideration of safety, scalability, reproducibility, regulatory compliance and patientspecific variables. Addressing these challenges is essential to realizing the therapeutic potential of stem cell based delivery platforms.

6.1 Safety and Risk Mitigation

Safety remains the foremost concern in the clinical translation of stem cell based therapies. Tumorigenicity, particularly associated with pluripotent stem cells and genetically modified constructs, poses a significant risk. Uncontrolled proliferation, unintended differentiation and genomic instability can lead to adverse outcomes. To mitigate these risks, current strategies include the use of lineage-restricted progenitor cells, transient genetic modification approaches and incorporation of inducible suicide genes that allow selective elimination of transplanted cells if necessary [19-20].

Immunogenicity represents another critical safety consideration. While certain stem cell populations, such as MSCs, exhibit low immunogenicity, immune recognition and clearance can still occur, particularly with repeated administration or allogeneic products. Autologous iPSC-derived platforms offer potential solutions but introduce challenges related to manufacturing time, cost, and patient-to-patient variability.

6.2 Biodistribution, Persistence, and Dosing

Understanding stem cell bio-distribution and persistence in-vivo is essential for optimizing therapeutic efficacy and minimizing off-target effects. Following systemic administration, a

substantial fraction of stem cells may become trapped in non-target organs, such as the lungs or liver, reducing delivery efficiency [19-20]. Advanced imaging modalities, including magnetic resonance imaging, positron emission tomography and bioluminescence imaging, are increasingly employed to track cell migration, retention, and survival in real time. Determining optimal dosing regimens remains challenging due to variability in cell survival, homing efficiency and cargo release kinetics. Unlike conventional drugs, stem cell based therapies introduce living, adaptive systems whose behaviour may evolve over time. As such, traditional pharmacokinetic and pharmacodynamic frameworks may require adaptation or redefinition to accommodate cell-based delivery platforms.

6.3 Manufacturing, Scalability and Standardization

Clinical translation necessitates robust, scalable manufacturing processes that ensure consistent product quality. Stem cell isolation, expansion, genetic modification and cargo loading must be performed under Good Manufacturing Practice (GMP) conditions, with stringent quality control measures. Variability in stem cell source, donor characteristics, culture conditions and passage number can significantly impact therapeutic performance. Standardization of potency assays, identity markers and release criteria is particularly challenging for multifunctional stem cell products that combine cellular activity with drug delivery. The development of consensus guidelines and validated assays will be critical for regulatory approval and widespread clinical adoption [19-20].

6.4 Regulatory and Ethical Challenges

Stem cell-based drug delivery systems often fall under the category of advanced therapy medicinal products (ATMPs) or combination products, subjecting them to complex and evolving regulatory frameworks. Products that incorporate genetic modifications, viral vectors, or synthetic nanomaterials face additional regulatory scrutiny. Early and sustained engagement with regulatory agencies is therefore essential to align preclinical study design with clinical and regulatory expectations. Ethical considerations, particularly those related to stem cell sourcing and genetic engineering, must also be addressed transparently. The use of iPSCs has alleviated some ethical concerns associated with embryonic stem cells but introduces new questions regarding genetic manipulation and long-term safety [19-20].

6.5 Clinical Trial Design and Patient Selection

Designing clinical trials for stem cell-based delivery systems requires careful consideration of patient selection, disease stage and endpoints. Heterogeneity in disease progression and patient biology can confound efficacy assessments [19-20]. Biomarkers that predict stem cell homing, persistence or therapeutic response may enable better patient stratification and improve trial outcomes. In addition, long-term follow-up is often required to assess delayed adverse events and sustained efficacy, increasing trial complexity and cost. Adaptive trial designs and real world evidence may play an increasingly important role in evaluating these therapies.

6.6 Emerging Strategies to Enhance Translatability

To address translational barriers, several emerging strategies are gaining traction. Cell-free approaches using stem cell derived extracellular vesicles offer improved safety profiles and simplified manufacturing. Hybrid platforms combining stem cells with synthetic carriers aim to harness the advantages of both biological targeting and engineered control. Advances in synthetic biology enable the development of programmable stem cells with built-in safety mechanisms and disease-responsive behaviour [19-20].

Discussion

Stem cell based targeted drug delivery systems represent a convergence of cellular biology, drug delivery science and bioengineering, offering a paradigm shift from passive pharmacological interventions toward dynamic, adaptive therapeutics. The studies and approaches discussed in this study collectively highlight the capacity of stem cells to overcome many of the fundamental limitations associated with conventional drug delivery platforms, particularly in the context of complex, heterogeneous and anatomically protected disease environments. By exploiting intrinsic homing behaviour, immunomodulatory properties and compatibility with advanced engineering strategies, stem cells provide a versatile foundation for next-generation precision therapies [7-16].

A central theme emerging from the literature is the unique advantage conferred by the biological intelligence of stem cells. Unlike synthetic carriers, stem cells actively sense pathological cues and migrate toward sites of injury, inflammation or tumorigenesis. This property is especially valuable in diseases such as cancer and neurological disorders, where therapeutic efficacy depends on reaching infiltrative or poorly accessible regions. However, this same adaptability introduces variability, as stem cell behaviour is influenced by microenvironmental context,

disease stage and host immune status. Understanding and harnessing this context dependence will be critical for improving predictability and reproducibility in clinical applications [16-20].

Another key insight is that stem cell-based delivery platforms are not merely passive carriers but can function as multifunctional therapeutic agents. In many settings, particularly inflammatory, autoimmune and ischemic diseases, the intrinsic paracrine and immunomodulatory effects of stem cells synergize with delivered therapeutic cargo. This dual activity may enhance efficacy but also complicates mechanistic interpretation and regulatory classification. Distinguishing the relative contributions of cell-intrinsic effects versus delivered therapeutics remains an important challenge for both preclinical study design and clinical evaluation. The increasing sophistication of stem cell engineering has substantially expanded the therapeutic landscape. Genetic modification, synthetic gene circuits and stimuli-responsive release systems enable precise control over therapeutic activity, allowing stem cells to act as programmable delivery platforms. These advances have been particularly impactful in oncology, where localized and conditional release of cytotoxic or immune-modulating agents can mitigate systemic toxicity. Nevertheless, the introduction of genetic modifications raises additional safety and regulatory concerns, emphasizing the need for robust safeguards such as inducible kill switches and transient expression systems. Cell-free alternatives, notably stem cell derived extracellular vesicles, have emerged as an important complementary strategy. These vesicles retain many of the targeting and signaling properties of parent stem cells while offering improved safety profiles and simplified manufacturing pathways. However, EV-based systems face their own challenges, including heterogeneity of cargo, limited loading capacity and difficulties in large-scale standardization. Future work will need to clarify whether EVs can fully substitute for live cell delivery or whether hybrid approaches will offer the greatest therapeutic benefit.

Despite substantial preclinical progress, clinical translation has been slower than anticipated. This gap underscores the complexity of translating living delivery systems into standardized, regulated therapeutics [16-20]. Variability in stem cell sources, manufacturing processes and in-vivo behaviour complicates dose definition, potency assessment and long term safety evaluation. Moreover, traditional pharmacokinetic and pharmacodynamic frameworks are often insufficient to describe therapies based on living cells, necessitating new conceptual and regulatory models.

Looking forward, the successful integration of stem cell based targeted drug delivery into clinical practice will depend on continued interdisciplinary collaboration. Advances in systems biology, biomaterials, imaging and computational modeling are expected to enhance our ability to predict

and control stem cell behaviour in-vivo. Equally important will be the development of rational clinical trial designs, predictive biomarkers and standardized manufacturing protocols. As these challenges are addressed, stem cell-based delivery platforms have the potential to evolve from experimental tools into clinically robust modalities, enabling highly targeted, adaptable and durable treatments for diseases that remain inadequately addressed by current therapeutic strategies.

Future Perspectives

The future of stem cell based targeted drug delivery systems lies in their evolution from experimental proof-of-concept platforms into clinically robust, programmable therapeutics. Continued advances in stem cell biology, synthetic biology and materials science are expected to transform stem cells into highly controllable delivery vehicles capable of sensing disease specific cues, making autonomous decisions and releasing therapeutic cargo with spatial and temporal precision. Such ‘smart’ stem cell systems could dynamically adapt to disease progression, offering a level of therapeutic responsiveness that is unattainable with conventional drug delivery approaches.

One key direction will be the integration of synthetic gene circuits and genome editing technologies to enable multiplexed functionality. Engineered stem cells may simultaneously detect pathological signals, deliver therapeutic agents and activate safety mechanisms, thereby improving efficacy while minimizing risk. Advances in CRISPR-based editing and non-integrating gene delivery methods are likely to enhance precision and reduce long-term safety concerns associated with permanent genetic modification. Cell-free strategies, particularly stem cell derived extracellular vesicles, are also expected to play an increasingly prominent role. Improvements in vesicle engineering, cargo loading efficiency and targeting specificity may allow EVs to serve as scalable, off-the-shelf alternatives to live-cell therapies, especially for chronic diseases requiring repeated administration. Hybrid platforms combining stem cells with synthetic nanoparticles or biomaterials may further expand therapeutic versatility by uniting biological targeting with engineered control.

From a translational standpoint, progress will depend on the establishment of standardized manufacturing pipelines, validated potency assays and predictive biomarkers of therapeutic response. Advances in non-invasive imaging and single-cell analysis will enable real-time monitoring of stem cell biodistribution, persistence and activity in patients, facilitating adaptive treatment strategies. Ultimately, the successful clinical adoption of stem cell based drug delivery

systems will require close collaboration among biologists, engineers, clinicians, and regulatory bodies. As these interdisciplinary efforts mature, stem cell mediated delivery has the potential to become a cornerstone of precision medicine, enabling targeted, durable, and personalized therapies for diseases that remain refractory to current treatment paradigms.

References

1. Allen TM, Cullis PR. Drug delivery systems: entering the mainstream. *Science*. 2004;303(5665):1818–22.
2. Bertrand N, Wu J, Xu X, Kamaly N, Farokhzad OC. Cancer nanotechnology: the impact of passive and active targeting in the era of modern cancer biology. *Adv Drug Deliv Rev*. 2014; 66:2–25.
3. Abbott NJ, Patabendige AA, Dolman DE, Yusof SR, Begley DJ. Structure and function of the blood–brain barrier. *Neurobiol Dis*. 2010;37(1):13–25.
4. Torchilin VP. Multi-functional nanocarriers. *Adv Drug Deliv Rev*. 2012;64:302–15.
5. Park K. Facing the truth about nanotechnology in drug delivery. *ACS Nano*. 2013;7(9):7442–7.
6. Wilhelm S, et al. Analysis of nanoparticle delivery to tumours. *Nat Rev Mater*. 2016;1:16014.
7. Aboody KS, et al. Neural stem cells as vehicles for cancer gene therapy. *Gene Ther*. 2000;7(7):552–60.
8. Studeny M, et al. Mesenchymal stem cells: potential precursors for tumor stroma and targeted-delivery vehicles for anticancer agents. *J Natl Cancer Inst*. 2004;96(21):1593–603.
9. Pittenger MF, et al. Multilineage potential of adult human mesenchymal stem cells. *Science*. 1999;284(5411):143–7.
10. Kucia M, et al. Chemokines and stem cell migration. *Stem Cells*. 2004;22(6):879–94.
11. Wynn RF, et al. A small proportion of mesenchymal stem cells strongly express functionally active CXCR4 receptor capable of promoting migration to bone marrow. *Blood*. 2004;104(9):2643–5.
12. Ranganath SH, et al. Harnessing stem cells for drug delivery. *Nat Rev Drug Discov*. 2012;11(10):825–44.

13. Huang X, et al. Cell-based drug delivery systems. *Adv Drug Deliv Rev.* 2017;122:58–72.
14. Shah K. Stem cell-based therapies for tumors in the brain. *Trends Mol Med.* 2009;15(10):485–93.
15. Aboody KS, et al. Stem cell-mediated enzyme/prodrug therapy for glioma. *Cancer Res.* 2006;66(7):3624–32.
16. Xie M, et al. Synthetic biology approaches for cancer therapy. *Nat Rev Cancer.* 2020;20(12):745–59.
17. Nissim L, Bar-Ziv RH. A tunable dual-promoter integrator for targeting of cancer cells. *Mol Syst Biol.* 2010;6:444.
18. El Andaloussi S, et al. Extracellular vesicles: biology and emerging therapeutic opportunities. *Nat Rev Drug Discov.* 2013;12(5):347–57.
19. Trounson A, McDonald C. Stem cell therapies in clinical trials: progress and challenges. *Cell Stem Cell.* 2015;17(1):11–22.
20. Bravery CA, et al. Regulatory considerations for cell-based therapies. *Regen Med.* 2013;8(5):585–602.
21. Le Blanc K, Mougiakakos D. Multipotent mesenchymal stromal cells and the innate immune system. *Nat Rev Immunol.* 2012;12(5):383–96.
22. Aggarwal S, Pittenger MF. Human mesenchymal stem cells modulate allogeneic immune cell responses. *Blood.* 2005;105(4):1815–22.
23. Roger M, et al. Mesenchymal stem cells as delivery vehicles for anticancer agents. *Mol Ther.* 2010;18(6):1089–99.
24. Takahashi K, Yamanaka S. Induction of pluripotent stem cells from adult human fibroblasts. *Cell.* 2007;131(5):861–72.
25. Shi Y, et al. Induced pluripotent stem cell technology: a decade of progress. *Nat Rev Drug Discov.* 2017;16(2):115–30.

Chapter 11:

Next-Generation Antimicrobials and Biologics

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Abstract

The accelerating emergence of antimicrobial resistance (AMR) represents a profound threat to global health systems, rendering many conventional antibiotics ineffective and challenging the sustainability of modern medicine. Traditional antimicrobial discovery pipelines have failed to keep pace with the rapid evolution of resistant pathogens, necessitating the exploration of fundamentally new therapeutic paradigms. This review provides an in-depth and critical analysis of next-generation antimicrobials and biologics developed to address AMR, including novel small-molecule antibiotics, antimicrobial peptides, bacteriophage-based therapies, monoclonal antibodies, CRISPR-based antimicrobials, microbiome-derived interventions, host-directed therapies, and nanotechnology-enabled antimicrobial platforms. Emphasis is placed on mechanisms of action, resistance mitigation strategies, translational feasibility, and clinical development status. While these emerging approaches demonstrate significant promise in overcoming classical resistance mechanisms, substantial challenges related to safety, scalability, regulatory pathways, and long-term ecological impact remain. By synthesizing current evidence and identifying key gaps in knowledge, this review outlines future research priorities essential for the successful clinical integration of next-generation antimicrobial and biologic therapies.

Keywords: Antimicrobial peptides, Antimicrobial resistance, Next-generation antimicrobials, Biologics, Phage therapy, Precision antimicrobials.

1. Introduction

Antimicrobial resistance (AMR) has emerged as one of the most critical biomedical challenges of the twenty-first century. The widespread misuse and overuse of antibiotics in clinical, agricultural, and environmental settings have accelerated the selection of resistant

microorganisms, leading to infections that are increasingly difficult, and in some cases impossible, to treat (Ventola, 2015). Recent global estimates indicate that bacterial AMR was directly responsible for over one million deaths in 2019 alone, with projections suggesting a dramatic rise in morbidity and mortality if current trends continue (Murray et al., 2022).

Conventional antibiotics largely rely on a limited number of molecular targets, such as cell wall synthesis, protein translation, and DNA replication. Pathogens have evolved diverse resistance mechanisms against these targets, including enzymatic degradation, target modification, efflux pump activation, and biofilm formation (Tacconelli et al., 2018). Compounding this problem, economic and regulatory barriers have discouraged pharmaceutical investment in antibiotic discovery, resulting in a stagnant pipeline dominated by incremental modifications of existing drug classes.

In response, next-generation antimicrobials and biologics have emerged as transformative alternatives. These strategies aim to exploit novel targets, leverage biological specificity, and integrate host–pathogen interactions to reduce resistance selection. Rather than replacing antibiotics outright, many of these approaches are envisioned as complementary or adjunctive therapies within a broader antimicrobial stewardship framework.

2. Emerging Classes of Next-Generation Antimicrobials and Biologics

The accelerating crisis of antimicrobial resistance has exposed fundamental limitations in conventional antibiotic-based treatment strategies. As pathogenic microorganisms continue to evolve mechanisms that neutralize or evade traditional drugs, the focus of antimicrobial research has expanded toward innovative therapeutic classes that operate through distinct biological principles. Next-generation antimicrobials and biologics are designed not only to suppress microbial growth but also to reduce resistance selection, preserve host microbial ecosystems, and integrate with host immune defenses.

One important category within this emerging landscape is **novel small-molecule antimicrobials** developed against unconventional or previously inaccessible bacterial targets. Unlike classical antibiotics that repeatedly exploit the same cellular pathways, these molecules interfere with structurally conserved elements essential for bacterial survival, such as membrane biogenesis or lipid precursor synthesis. By targeting functions that are less amenable to mutational escape, these agents aim to slow resistance emergence while maintaining potent antibacterial activity

against multidrug-resistant organisms. Their development has been facilitated by advances in microbial genomics, structure-guided drug design, and innovative cultivation technologies.

Another rapidly advancing class comprises **antimicrobial peptides**, which are naturally occurring components of innate immune systems across multiple species. These peptides exert antimicrobial effects through rapid and often multifaceted mechanisms, including disruption of microbial membranes, interference with intracellular processes, and modulation of host immune responses. Because their activity is not dependent on a single molecular target, microorganisms find it difficult to develop stable resistance. Recent progress in peptide chemistry and bioengineering has addressed earlier concerns related to instability, toxicity, and short half-life, allowing antimicrobial peptides to emerge as viable therapeutic candidates.

Bacteriophage-based therapies represent a biologically precise alternative to broad-spectrum antibiotics. Phages specifically infect bacterial hosts, replicate within them, and induce cell lysis, thereby eliminating pathogens without disturbing beneficial microbial populations. This specificity is particularly advantageous in the context of microbiome preservation. Modern approaches increasingly rely on genetically modified phages engineered for improved host range, enhanced antibacterial potency, or delivery of auxiliary antimicrobial functions. Despite their promise, challenges related to standardization, regulatory frameworks, and host immune responses must be resolved for widespread clinical adoption.

Immune-based antimicrobials, including monoclonal antibodies, operate through mechanisms fundamentally distinct from bactericidal drugs. Rather than directly killing bacteria, these biologics neutralize virulence factors, block pathogen adhesion, or enhance immune-mediated clearance. This indirect mode of action minimizes selective pressure for resistance while offering therapeutic benefit, particularly in severe or recurrent infections. Although their use may be limited by production costs and pathogen specificity, immune-based strategies are well suited for targeted or adjunctive therapy.

A highly innovative and rapidly evolving class is **CRISPR-based antimicrobials**, which apply gene-editing technologies to infectious disease treatment. These systems can be programmed to selectively disrupt resistance genes or eliminate specific bacterial strains at the genetic level. Such precision offers the possibility of eradicating pathogenic bacteria while leaving surrounding microbial communities intact. While clinical translation remains in early stages, CRISPR-based

approaches represent a conceptual shift toward precision antimicrobials tailored at the genomic level.

Microbiome-centered and host-directed therapies reflect a growing recognition of the host environment as a critical determinant of infection outcomes. By strengthening colonization resistance, restoring microbial balance, or modulating host immune pathways, these approaches reduce pathogen dominance without relying on direct antimicrobial pressure. Host-directed therapies, in particular, enhance immune effectiveness while lowering the likelihood of resistance development, although careful modulation is required to avoid excessive inflammation or immune dysregulation.

Finally, **nanotechnology-enabled antimicrobial systems** integrate materials science with therapeutic innovation. Nanocarriers can improve drug stability, enable targeted delivery, and facilitate controlled release at infection sites. Some nanomaterials also possess intrinsic antimicrobial properties, further enhancing therapeutic efficacy. These systems are especially valuable for combination therapies that incorporate antibiotics, peptides, or biologics within a single platform, although long-term safety and environmental impact require continued evaluation.

2.1 Novel Small-Molecule Antimicrobials

Despite a global decline in the discovery success rate of traditional antibiotics, small-molecule antimicrobials continue to serve as a foundational pillar of infectious disease treatment, owing to their well-characterized pharmacokinetics, oral bioavailability, and scalable manufacturing. In response to widespread resistance against classical drug classes, contemporary discovery efforts have shifted away from incremental chemical modifications toward small molecules with fundamentally novel mechanisms of action and targets that are evolutionarily constrained.

A landmark example of this paradigm shift is **teixobactin**, a non-ribosomally synthesized depsipeptide antibiotic identified using the iChip cultivation platform. Teixobactin exerts bactericidal activity by binding to **lipid II and lipid III**, essential precursors in peptidoglycan and teichoic acid biosynthesis, respectively. Importantly, these targets are non-proteinaceous and highly conserved across Gram-positive bacteria, which significantly reduces the probability of resistance arising through point mutations or enzymatic modification. Experimental studies have demonstrated potent activity of teixobactin against *Staphylococcus aureus*, *Mycobacterium*

tuberculosis, and *Clostridioides difficile*, with no detectable resistance observed after prolonged exposure under laboratory conditions (Ling et al., 2015). Subsequent structure–activity relationship analyses have further highlighted its membrane-associated binding mode, which contributes to its robust antibacterial efficacy.

Similarly, **darobactin** represents a breakthrough in addressing the long-standing challenge of treating Gram-negative infections. Darobactin selectively targets **BamA**, a critical component of the β -barrel assembly machinery responsible for outer membrane protein insertion. By binding to the lateral gate of BamA, darobactin disrupts outer membrane biogenesis, leading to bacterial cell death. This mechanism is particularly significant because the Gram-negative outer membrane has historically limited antibiotic penetration and efficacy. Preclinical studies have shown darobactin to be highly effective against priority pathogens such as *Escherichia coli*, *Klebsiella pneumoniae*, and *Acinetobacter baumannii*, including strains resistant to carbapenems and colistin (Imai et al., 2019). In vivo murine infection models further demonstrated favorable therapeutic indices and low toxicity profiles, supporting its translational potential.

Collectively, these discoveries underscore a renewed viability of small-molecule antibiotics when guided by innovative discovery platforms, unconventional targets, and mechanistic novelty. Advances in metagenomic mining, artificial intelligence–assisted molecular modeling, and high-throughput phenotypic screening have expanded access to previously untapped chemical space. While challenges related to optimization, toxicity, and clinical translation persist, next-generation small-molecule antibiotics such as teixobactin and darobactin exemplify how rational target selection can overcome historical resistance barriers and reinvigorate the antimicrobial pipeline.

2.2 Antimicrobial Peptides (AMPs)

Antimicrobial peptides represent one of the most ancient and evolutionarily conserved defense mechanisms employed by living organisms to control microbial invasion. Found in organisms ranging from microorganisms and plants to insects and mammals, these peptides form a critical component of innate immunity. In humans, AMPs are produced by epithelial barriers and immune cells, where they function at the interface between host defense and microbial exposure. Structurally, AMPs are generally short peptides enriched in positively charged and hydrophobic residues, a composition that enables selective interaction with microbial membranes that are rich in negatively charged phospholipids.

The antimicrobial activity of AMPs is distinguished by its speed and versatility. Upon contact with microbial cells, many AMPs rapidly associate with the cell envelope, destabilizing membrane architecture through physical disruption rather than receptor-dependent binding. This interaction can lead to pore formation, membrane thinning, or complete loss of membrane integrity, resulting in swift cell death. In addition to membrane-based effects, a growing body of evidence demonstrates that several AMPs translocate into the cytoplasm, where they interfere with essential intracellular processes such as nucleic acid synthesis, protein folding, enzymatic activity, and cell division. This ability to act on multiple cellular targets simultaneously is a defining feature that differentiates AMPs from conventional antibiotics.

Beyond direct antimicrobial effects, AMPs exert profound influences on host immune function. Many peptides modulate inflammatory responses by regulating cytokine production, recruiting immune cells to sites of infection, and enhancing phagocytic activity. Some AMPs promote tissue repair and angiogenesis, linking antimicrobial defense with wound healing processes. These immunomodulatory properties broaden their therapeutic relevance, particularly in infections where immune dysregulation contributes to disease severity.

A major advantage of AMPs lies in their reduced propensity for resistance development. Because their primary mode of action relies on fundamental physicochemical interactions with microbial membranes rather than highly specific molecular targets, microorganisms face substantial evolutionary constraints when attempting to evade AMP activity. Resistance, when observed, often involves complex alterations in membrane composition or regulatory pathways that impose significant fitness costs, limiting the spread and stability of resistant phenotypes. This contrasts sharply with resistance mechanisms against classical antibiotics, which can arise rapidly through single-gene mutations or horizontal gene transfer.

Despite their promise, early efforts to develop AMPs as therapeutic agents encountered significant obstacles. Rapid degradation by proteases, limited bioavailability, cytotoxicity at higher concentrations, and challenges associated with large-scale synthesis restricted clinical progress. Recent advances in peptide design and chemical modification have substantially addressed these issues. Strategic alterations in amino acid composition, incorporation of non-natural residues, peptide cyclization, and terminal modifications have improved resistance to enzymatic degradation while preserving antimicrobial potency. Such refinements have enabled the generation of AMP analogues with improved selectivity and safety profiles.

Parallel progress in drug delivery technologies has further enhanced the clinical feasibility of AMPs. Encapsulation within nanocarriers, hydrogels, and lipid-based systems protects peptides from premature degradation and enables localized or sustained release at infection sites. These approaches are particularly advantageous for treating chronic and biofilm-associated infections, where conventional antibiotics often fail due to poor penetration and reduced bacterial metabolic activity. In some cases, AMPs have demonstrated synergistic effects when combined with existing antibiotics, restoring susceptibility in resistant strains.

Several AMP-based candidates have advanced into clinical and late-stage preclinical development, particularly for topical, inhalational, and localized systemic applications. While not all candidates have successfully navigated clinical trials, these experiences have provided valuable insights into dosing strategies, formulation requirements, and regulatory expectations. Importantly, the expanding understanding of AMP biology has shifted the field away from viewing these molecules solely as antibiotics and toward recognizing them as multifunctional host-defense modulators.

Key challenges remain. Manufacturing costs, long-term safety, immunogenicity, and regulatory classification continue to influence the pace of clinical translation. In addition, the dual antimicrobial and immunomodulatory nature of AMPs necessitates careful therapeutic balancing to avoid unintended inflammatory effects. Addressing these challenges will require coordinated efforts spanning peptide chemistry, immunology, pharmacology, and clinical sciences.

2.3 Phage Therapy and Engineered Bacteriophages

Bacteriophages are naturally occurring viruses that infect bacteria with remarkable specificity, making them uniquely suited for targeted antimicrobial intervention. Unlike chemical antibiotics, phages initiate infection by recognizing distinct bacterial surface receptors, followed by intracellular replication and subsequent lysis of the host cell. This self-propagating mode of action enables phages to concentrate their activity at sites of infection, where bacterial density is highest, while diminishing once the target population is eliminated.

The renewed scientific and clinical interest in phage therapy stems largely from its effectiveness against infections caused by multidrug-resistant bacteria, particularly in cases where conventional antibiotics have failed. A critical advantage of phage-based treatment lies in its ability to selectively eliminate pathogenic organisms without disrupting beneficial commensal

microbiota. Preservation of microbial diversity is increasingly recognized as essential for maintaining immune balance and preventing secondary infections, a benefit rarely achieved with broad-spectrum antibiotics.

Advances in molecular biology and synthetic engineering have further enhanced the therapeutic promise of phages. Engineered bacteriophages can be modified to overcome narrow host range limitations, improve stability under physiological conditions, and evade premature immune clearance. In addition, phages can be designed to deliver auxiliary antimicrobial functions, such as enzymes that degrade bacterial biofilms or genetic elements that suppress virulence. These innovations allow phage therapy to address complex infections involving biofilm formation or heterogeneous bacterial populations.

Despite encouraging clinical case reports and early-stage trials, several barriers continue to limit routine clinical deployment. Regulatory approval pathways remain fragmented due to the biological variability of phages and their capacity for evolution. Manufacturing challenges, including large-scale production, purification, and quality control, further complicate translation. Moreover, bacterial resistance to phages can emerge through receptor modification, necessitating adaptive phage selection strategies or cocktail formulations. Overcoming these challenges will require coordinated regulatory frameworks and standardized production protocols.

2.4 Monoclonal Antibodies and Immune-Based Antimicrobials

Monoclonal antibodies represent a fundamentally different antimicrobial strategy by targeting bacterial pathogenicity rather than bacterial viability. These biologic agents function by neutralizing toxins, blocking adherence to host tissues, or enhancing immune-mediated clearance mechanisms such as opsonization and phagocytosis. By acting indirectly, monoclonal antibodies minimize selective pressure on bacterial survival pathways, thereby reducing the likelihood of resistance development.

Immune-based antimicrobials have demonstrated particular utility in infections where disease severity is driven by toxin production or immune evasion rather than bacterial burden alone. In such contexts, antibody-mediated neutralization can significantly reduce tissue damage and improve clinical outcomes when used alongside standard antimicrobial therapy. Their specificity also allows preservation of host microbiota, an increasingly important consideration in long-term infection management.

The clinical application of monoclonal antibodies is further supported by their predictable pharmacokinetics and favorable safety profiles. However, their high production costs, limited spectrum of activity, and dependence on accurate pathogen identification constrain widespread use. Consequently, immune-based antimicrobials are most effective as adjunctive therapies or prophylactic interventions in high-risk populations, such as immunocompromised patients or individuals undergoing invasive medical procedures.

Ongoing research aims to expand the applicability of antibody-based therapies through the development of broadly reactive antibodies, antibody fragments, and multi-target constructs. Integration with rapid diagnostic technologies is expected to further enhance their clinical value by enabling timely and precise administration.

2.5 CRISPR-Based Precision Antimicrobials

CRISPR-based antimicrobials represent a paradigm shift in infectious disease treatment by enabling direct genetic targeting of pathogens. These systems utilize programmable nucleases capable of recognizing and cleaving specific DNA sequences, allowing selective elimination of bacterial strains or resistance determinants with exceptional precision. Unlike conventional antimicrobials, CRISPR-based approaches can be designed to discriminate between closely related bacterial populations based on genomic signatures.

One of the most promising applications of this technology lies in the targeted removal of antibiotic resistance genes. By selectively disabling resistance mechanisms rather than indiscriminately killing bacteria, CRISPR antimicrobials may restore the effectiveness of existing antibiotics while minimizing disruption to beneficial microbial communities. This strategy aligns closely with the principles of antimicrobial stewardship and precision medicine.

The principal obstacle to clinical translation remains efficient and safe delivery of CRISPR components to target bacteria. Current delivery platforms include bacteriophages, plasmid-based systems, and nanoparticle carriers, each presenting trade-offs between specificity, efficiency, and immunogenicity. While phage-mediated delivery offers high targeting accuracy, non-viral delivery systems may provide greater control over dosing and distribution. Continued refinement of delivery technologies will be critical for advancing CRISPR antimicrobials beyond experimental settings.

2.6 Microbiome-Derived and Host-Directed Therapies

The human microbiome plays an essential role in maintaining resistance to pathogen colonization and regulating immune responses. Antibiotic-induced disruption of microbial communities can weaken these protective functions, increasing susceptibility to recurrent and opportunistic infections. Microbiome-derived therapies aim to restore or reinforce microbial balance as a means of disease control, shifting the therapeutic focus from pathogen eradication to ecosystem stabilization.

Interventions such as fecal microbiota transplantation have demonstrated the therapeutic potential of microbiome restoration, particularly in recurrent gastrointestinal infections. Building on these successes, next-generation approaches involve defined microbial consortia, engineered probiotics, and microbial metabolites designed to suppress pathogens or enhance immune resilience in a controlled and reproducible manner.

Host-directed therapies offer a complementary strategy by modulating host immune pathways rather than targeting microbes directly. These approaches enhance innate defense mechanisms, improve barrier integrity, or fine-tune inflammatory responses to promote pathogen clearance while reducing tissue damage. By bypassing direct antimicrobial pressure, host-directed therapies inherently limit resistance development. However, precise regulation is essential to avoid immune overactivation or unintended systemic effects.

3. Nanotechnology-Enabled Antimicrobial Systems

Nanotechnology has emerged as a powerful tool to enhance antimicrobial efficacy and delivery. Nanoparticles can improve drug solubility, protect labile biologics, and enable targeted delivery to infection sites (Wang et al., 2017). Additionally, certain nanomaterials possess intrinsic antimicrobial properties, including membrane disruption and reactive oxygen species generation.

Nanotechnology-enabled systems also facilitate combination therapies, integrating antibiotics, peptides, or biologics into multifunctional platforms. However, concerns regarding long-term toxicity, environmental accumulation, and regulatory approval must be addressed.

4. Challenges and Knowledge Gaps

Despite significant promise, next-generation antimicrobials face substantial hurdles. Manufacturing complexity, high development costs, and uncertain regulatory pathways hinder commercialization. Long-term ecological effects, including impacts on microbiota and resistance evolution, remain poorly understood. Furthermore, integration into existing clinical workflows and stewardship programs requires careful consideration.

Conclusion

Future research should emphasize combination therapies that integrate conventional antibiotics with biologics or host-directed approaches. Advances in diagnostics, biomarker discovery, and personalized medicine will be critical for matching therapies to specific pathogens and patient profiles. Strengthening interdisciplinary collaboration between microbiology, immunology, engineering, and regulatory science will accelerate translation from bench to bedside.

Next-generation antimicrobials and biologics represent a fundamental shift in the management of infectious diseases. By transcending the limitations of traditional antibiotics, these innovative strategies offer a sustainable path forward in the global fight against AMR. Continued investment, rigorous clinical evaluation, and adaptive regulatory frameworks will be essential to realize their full therapeutic potential.

Reference

- Abedon, S. T., Kuhl, S. J., Blasdel, B. G., & Kutter, E. M. (2011). Phage treatment of human infections. *Bacteriophage*, 1(2), 66–85
- Bikard, D., Euler, C. W., Jiang, W., Nussenzweig, P. M., Goldberg, G. W., Duportet, X., Fischetti, V. A., & Marraffini, L. A. (2014). Exploiting CRISPR-Cas nucleases to produce sequence-specific antimicrobials. *Nature Biotechnology*, 32(11), 1146–1150
- Brogden, K. A. (2005). Antimicrobial peptides: Pore formers or metabolic inhibitors in bacteria? *Nature Reviews Microbiology*, 3(3), 238–250
- Brown, E. D., & Wright, G. D. (2016). Antibacterial drug discovery in the resistance era. *Nature*, 529(7586), 336–343

- Greber, K. E., & Dawgul, M. (2017). Antimicrobial peptides under clinical trials. *Current Topics in Medicinal Chemistry*, 17(5), 620–628
- Hancock, R. E. W., & Sahl, H. G. (2006). Antimicrobial and host-defense peptides as new anti-infective therapeutic strategies. *Nature Biotechnology*, 24(12), 1551–1557
- Hancock, R. E. W., Haney, E. F., & Gill, E. E. (2016). The immunology of host defence peptides: Beyond antimicrobial activity. *Nature Reviews Immunology*, 16(5), 321–334
- Imai, Y., Meyer, K. J., Iinishi, A., Favre-Godal, Q., Green, R., Manuse, S., ... Lewis, K. (2019). A new antibiotic selectively kills Gram-negative pathogens. *Nature*, 576(7787), 459–464
- Lewis, K. (2020). The science of antibiotic discovery. *Cell*, 181(1), 29–45
- Ling, L. L., Schneider, T., Peoples, A. J., Spoering, A. L., Engels, I., Conlon, B. P., ... Lewis, K. (2015). A new antibiotic kills pathogens without detectable resistance. *Nature*, 517(7535), 455–459
- Mahlapuu, M., Håkansson, J., Ringstad, L., & Björn, C. (2016). Antimicrobial peptides: An emerging category of therapeutic agents. *Frontiers in Cellular and Infection Microbiology*, 6, 194
- Mookherjee, N., Anderson, M. A., Haagsman, H. P., & Davidson, D. J. (2020). Antimicrobial host defence peptides: Functions and clinical potential. *Nature Reviews Drug Discovery*, 19(5), 311–332
- Murray, C. J. L., Ikuta, K. S., Sharara, F., Swetschinski, L., Aguilar, G. R., Gray, A., ... Naghavi, M. (2022). Global burden of bacterial antimicrobial resistance in 2019. *The Lancet*, 399(10325), 629–655
- Pamer, E. G. (2016). Resurrecting the intestinal microbiota to combat antibiotic-resistant pathogens. *Science*, 352(6285), 535–538
- Pires, D. P., Cleto, S., Sillankorva, S., Azeredo, J., & Lu, T. K. (2020). Genetically engineered phages: A review of advances over the last decade. *Microbiology and Molecular Biology Reviews*, 84(2), e00069-19
- Silver, L. L. (2011). Challenges of antibacterial discovery. *Clinical Microbiology Reviews*, 24(1), 71–109
- Tacconelli, E., Carrara, E., Savoldi, A., Harbarth, S., Mendelson, M., Monnet, D. L., ... WHO Pathogens Priority List Working Group. (2018). Discovery, research, and development of new antibiotics. *The Lancet Infectious Diseases*, 18(3), 318–327. [https://doi.org/10.1016/S1473-3099\(17\)30753-3](https://doi.org/10.1016/S1473-3099(17)30753-3)
- Tommasi, R., Brown, D. G., Walkup, G. K., Manchester, J. I., & Miller, A. A. (2015). ESKAPEing the labyrinth of antibacterial discovery. *Nature Reviews Drug Discovery*, 14(8), 529–542

- Ventola, C. L. (2015). The antibiotic resistance crisis. *Pharmacy and Therapeutics*, 40(4), 277–283
- Ventola, C. L. (2015). The antibiotic resistance crisis: Part 1: Causes and threats. *Pharmacy and Therapeutics*, 40(4), 277–283
- Wang, L., Hu, C., & Shao, L. (2017). The antimicrobial activity of nanoparticles. *International Journal of Nanomedicine*, 12, 1227–1249
- Zurawski, D. V., & McLendon, M. K. (2020). Monoclonal antibodies as an antibacterial approach. *Antibiotics*, 9(4), 155

Chapter 12:

Light Stress-Induced Neuronal Regulation: Photoreceptor Signaling Cascades and Neuroprotective Mechanisms

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Abstract

Excessive or aberrant light exposure induces photoreceptor stress that triggers complex neuronal regulatory responses encompassing oxidative damage mitigation, inflammatory modulation, and adaptive neuroprotection. This comprehensive review examines the molecular mechanisms underlying light stress-induced neuronal regulation, with emphasis on retinal photoreceptor cells and their integration with circadian and homeostatic networks. We delineate how phototransduction overstimulation generates reactive oxygen species through rhodopsin photobleaching and mitochondrial dysfunction, activating stress-responsive pathways including mitogen-activated protein kinase and unfolded protein response cascades. Transcriptomic and proteomic profiling reveal coordinated upregulation of antioxidant enzymes including superoxide dismutase, catalase, and glutathione peroxidase, alongside heat shock proteins and anti-apoptotic factors such as Bcl-2 and XIAP as endogenous neuroprotective responses. Furthermore, light stress modulates neurotransmitter signaling, particularly dopaminergic and GABAergic systems, influencing synaptic plasticity and neuronal excitability in retinal and extraretinal circuits. Chronic light stress exposure is implicated in photoreceptor degeneration, disrupted circadian rhythmicity, and increased susceptibility to neurodegenerative conditions including age-related macular degeneration and retinitis pigmentosa. We evaluate emerging therapeutic strategies targeting oxidative stress pathways, including antioxidant supplementation, dark adaptation protocols, and pharmacological modulation of stress-response genes. Understanding the intricate interplay between photic stress and neuronal regulatory mechanisms is essential for developing interventions to preserve visual function and maintain neuronal homeostasis in light-challenged environments.

Keywords: Circadian rhythm, Neuroprotection, Oxidative stress, Photoreceptor cells, Retinal degeneration

1. Introduction

Light serves as the fundamental stimulus for vision, enabling detection and interpretation of electromagnetic radiation through specialized photoreceptor neurons in the retina. However, the same photic energy that allows visual perception also poses substantial threats to photoreceptor cell integrity and survival. Excessive or inappropriate light exposure generates oxidative stress, disrupts protein homeostasis, and triggers apoptotic cascades that compromise photoreceptor viability. Understanding the molecular mechanisms underlying light-induced cellular damage and the endogenous protective responses that counteract this damage is critical for developing therapeutic strategies to preserve visual function and prevent photoreceptor degeneration (Organisciak & Vaughan, 2010).

Photoreceptor cells face unique challenges arising from their specialized structure and function. The outer segments of rod and cone photoreceptors contain extraordinarily high concentrations of photopigments and membranes enriched in polyunsaturated fatty acids, creating an environment highly susceptible to light-induced oxidative damage. Continuous phototransduction activity generates reactive oxygen species through multiple mechanisms, including rhodopsin photobleaching, mitochondrial respiration supporting high metabolic demands, and photosensitization reactions. These oxidative challenges are compounded by high oxygen tension in the retina, which receives among the highest blood flow rates per tissue mass in the body to support photoreceptor metabolism (Beatty et al., 2000).

This review synthesizes current understanding of light stress-induced neuronal regulation in photoreceptor systems, examining the molecular pathways mediating oxidative damage, the complex network of endogenous neuroprotective mechanisms, and the pathophysiological consequences of chronic light stress. We analyze how light stress integrates with circadian regulation, evaluate therapeutic strategies targeting stress response pathways, and discuss implications for understanding and treating retinal degenerative diseases.

2. Photoreceptor Structure and Phototransduction

2.1 Photoreceptor Cell Organization

Vertebrate photoreceptor cells are highly polarized neurons comprising four structural compartments: the outer segment containing photopigment-laden membranous discs, the inner segment housing cellular biosynthetic and metabolic machinery, the cell body containing the nucleus, and the synaptic terminal forming connections with downstream retinal neurons. This compartmentalization reflects functional specialization, with outer segments dedicated to photon capture and signal initiation, inner segments

supporting energy production and protein synthesis, and synaptic terminals mediating neurotransmitter release (Fu & Yau, 2007).

Rod photoreceptors, specialized for dim light vision, contain the photopigment rhodopsin at concentrations exceeding 3 millimolar in outer segment membranes. Cone photoreceptors, responsible for color vision and high-acuity spatial discrimination, express one of three cone opsins with distinct spectral sensitivities. The extraordinary concentration and organization of photopigments enables efficient photon capture but creates vulnerability to light-induced damage.

2.2 Phototransduction Cascade

Photon absorption by rhodopsin triggers its isomerization to the activated metarhodopsin II state, which catalyzes activation of the G protein transducin. Activated transducin stimulates phosphodiesterase, which hydrolyzes cyclic GMP, causing closure of cyclic GMP-gated cation channels in the plasma membrane. Channel closure hyperpolarizes the photoreceptor, reducing glutamate release from synaptic terminals and signaling photon detection to postsynaptic neurons. This amplification cascade enables single-photon detection while requiring precise regulation to prevent overstimulation (Arshavsky et al., 2002).

Recovery from photoactivation involves multiple regulatory mechanisms including rhodopsin phosphorylation by rhodopsin kinase, arrestin binding to phosphorylated rhodopsin preventing further transducin activation, and guanylate cyclase-mediated cyclic GMP synthesis restoring basal channel activity. Disruption of these regulatory processes through excessive light exposure or genetic mutations compromises photoreceptor function and viability.

3. Mechanisms of Light-Induced Oxidative Stress

3.1 Rhodopsin Photobleaching and Chromophore Regeneration

Following photoisomerization, the retinal chromophore dissociates from opsin and must be enzymatically converted back to 11-cis-retinal for rhodopsin regeneration. This visual cycle involves multiple enzymatic steps occurring in photoreceptors and retinal pigment epithelium. However, aberrant reactions during chromophore processing can generate reactive aldehydes and other toxic byproducts that contribute to oxidative stress. Additionally, incomplete regeneration leads to accumulation of all-trans-retinal, which can form condensation products with phosphatidylethanolamine generating lipofuscin, a photoreactive aggregate implicated in retinal degeneration (Saari, 2012).

3.2 Mitochondrial Reactive Oxygen Species Production

Photoreceptor inner segments contain abundant mitochondria that support the high ATP demands of phototransduction and protein synthesis. Mitochondrial respiration inevitably generates reactive oxygen species including superoxide anion, hydrogen peroxide, and hydroxyl radicals through electron leakage from the respiratory chain. Light stimulation increases metabolic rate and consequently enhances mitochondrial reactive oxygen species production. Under excessive light exposure, reactive oxygen species generation can overwhelm antioxidant defenses, causing oxidative damage to lipids, proteins, and DNA (Kowluru & Chan, 2007).

3.3 Photosensitization Reactions

Photopigments and their derivatives can act as photosensitizers, absorbing light energy and transferring it to molecular oxygen, generating highly reactive singlet oxygen. This process, termed photosensitization, occurs particularly when photopigment concentration is high and when lipofuscin accumulates. Singlet oxygen reacts indiscriminately with cellular macromolecules, causing peroxidation of membrane lipids, protein cross-linking, and DNA strand breaks. The polyunsaturated fatty acids abundant in photoreceptor outer segment membranes are particularly vulnerable to peroxidative damage (Organisciak & Vaughan, 2010).

4. Stress-Responsive Signaling Pathways

4.1 MAPK Cascades

Mitogen-activated protein kinase pathways, including ERK, JNK, and p38 MAPK, are activated in response to light stress and mediate diverse cellular responses ranging from survival signals to apoptosis induction depending on intensity and duration of activation. Light stress preferentially activates JNK and p38 MAPK pathways, which phosphorylate transcription factors regulating expression of stress response genes. While transient MAPK activation can promote adaptive responses including antioxidant enzyme upregulation, sustained activation drives apoptotic programs contributing to photoreceptor cell death (Samuels et al., 2010).

4.2 Unfolded Protein Response

The unfolded protein response is triggered when accumulation of misfolded proteins in the endoplasmic

reticulum exceeds the capacity of protein quality control machinery. Light stress, particularly when chronic, disrupts protein homeostasis through oxidative damage to nascent proteins and impairment of chaperone function. Unfolded protein response activation initiates through three sensor proteins: IRE1, PERK, and ATF6, which coordinately upregulate chaperone expression, attenuate translation, and enhance degradation of misfolded proteins. However, if proteotoxic stress persists, the unfolded protein response transitions from cytoprotective to pro-apoptotic signaling, contributing to photoreceptor degeneration (Lin et al., 2007).

5. Endogenous Neuroprotective Mechanisms

5.1 Antioxidant Enzyme Systems

Photoreceptors express robust antioxidant enzyme systems that detoxify reactive oxygen species and maintain redox homeostasis. Superoxide dismutase catalyzes conversion of superoxide to hydrogen peroxide, which is subsequently reduced to water by catalase and glutathione peroxidase. Glutathione reductase and glucose-6-phosphate dehydrogenase maintain glutathione in its reduced state, enabling continuous peroxidase activity. Transcriptomic analyses reveal that light stress induces coordinate upregulation of these antioxidant enzymes through activation of transcription factors including Nrf2, which binds antioxidant response elements in gene promoters (Komeima et al., 2006).

5.2 Heat Shock Proteins

Heat shock proteins function as molecular chaperones that prevent protein aggregation, promote refolding of damaged proteins, and facilitate degradation of irreparably damaged proteins. Multiple heat shock protein families including Hsp70, Hsp90, and small heat shock proteins are upregulated in response to light stress. These chaperones protect key cellular proteins from oxidative damage and help maintain proteostasis under stress conditions. Overexpression of heat shock proteins enhances photoreceptor survival following light damage, while their inhibition exacerbates light-induced degeneration, demonstrating critical protective roles (Athanasίου et al., 2012).

5.3 Anti-Apoptotic Factors

The Bcl-2 family of proteins regulates mitochondrial outer membrane permeabilization, a critical commitment point in apoptosis. Anti-apoptotic members including Bcl-2, Bcl-xL, and Mcl-1 oppose pro-apoptotic proteins such as Bax and Bak, preventing cytochrome c release and caspase activation. Light stress induces upregulation of anti-apoptotic Bcl-2 family members as a protective response. Similarly,

inhibitor of apoptosis proteins including XIAP and survivin are upregulated and block caspase activity, providing additional protection against apoptotic cell death. The balance between pro- and anti-apoptotic factors determines photoreceptor fate following light stress (Hao et al., 2002).

6. Neurotransmitter Signaling Modulation

6.1 Dopaminergic System

Retinal dopamine, synthesized by a subset of amacrine cells, modulates numerous aspects of retinal function including photoreceptor sensitivity, gap junction coupling, and circadian rhythms. Light stress alters dopamine synthesis and release, with initial increases followed by depletion during prolonged exposure. Dopamine acts through D1-like and D2-like receptors to influence cyclic AMP signaling and downstream gene expression. Disruption of dopaminergic signaling impairs light adaptation and increases susceptibility to light damage, while dopamine supplementation provides modest neuroprotection in some experimental models (Witkovsky, 2004).

6.2 GABAergic System

Gamma-aminobutyric acid serves as the primary inhibitory neurotransmitter in the retina, released by horizontal cells and numerous amacrine cell subtypes. GABAergic signaling influences photoreceptor function through modulation of calcium homeostasis and regulation of neurotransmitter release. Light stress affects GABAergic transmission through changes in GABA synthesis, release, and receptor expression. Enhanced GABAergic inhibition may limit excitotoxicity and reduce metabolic stress, contributing to neuroprotection, while dysregulated GABAergic signaling can impair retinal processing and exacerbate damage (Yazulla, 2008).

7. Circadian Integration of Light Stress Responses

Photoreceptors exhibit circadian rhythms in metabolism, gene expression, and sensitivity to light damage, with peak vulnerability typically occurring during the late subjective night when antioxidant defenses are lowest. The circadian clock in photoreceptors is synchronized by light but also generates autonomous oscillations that persist in constant darkness. Clock genes including *Bmal1*, *Clock*, *Per*, and *Cry* regulate rhythmic expression of hundreds of genes involved in metabolism, redox homeostasis, and stress responses. Disruption of circadian rhythms through irregular light schedules or clock gene mutations increases photoreceptor susceptibility to light damage, highlighting the importance of temporal organization in neuroprotection (Baba et al., 2009).

8. Chronic Light Stress and Retinal Degeneration

8.1 Age-Related Macular Degeneration

Age-related macular degeneration is the leading cause of vision loss in elderly populations in developed nations, affecting central high-acuity vision. While multifactorial in etiology, cumulative light exposure over decades is recognized as a significant risk factor. Chronic low-level light stress may contribute to disease pathogenesis through cumulative oxidative damage, lipofuscin accumulation in retinal pigment epithelium, and chronic inflammatory activation. Blue light, which has higher energy and greater capacity for photosensitization reactions, appears particularly deleterious. Epidemiological studies demonstrate associations between outdoor light exposure and macular degeneration risk, supporting light stress as a modifiable risk factor (Beatty et al., 2000).

8.2 Retinitis Pigmentosa

Retinitis pigmentosa encompasses a heterogeneous group of inherited retinal degenerations caused by mutations in genes essential for photoreceptor structure and function. While genetic mutations initiate disease, light exposure modulates progression in many forms. Photoreceptors carrying disease-causing mutations often exhibit heightened vulnerability to light stress, with accelerated degeneration under normal lighting conditions that would not damage wild-type cells. This enhanced susceptibility likely reflects impaired stress response capacity or baseline cellular dysfunction that reduces tolerance to additional insults. Understanding interactions between genetic susceptibility and environmental light stress is critical for developing preventive strategies (Cideciyan, 2010).

9. Therapeutic Strategies

9.1 Antioxidant Supplementation

Dietary antioxidants including vitamins C and E, carotenoids such as lutein and zeaxanthin, and omega-3 fatty acids have been investigated for retinal neuroprotection. The Age-Related Eye Disease Study demonstrated that high-dose antioxidant and zinc supplementation reduces progression to advanced age-related macular degeneration in individuals with intermediate disease. Lutein and zeaxanthin, which accumulate in the macula and absorb blue light while providing antioxidant protection, show particular promise. While supplementation cannot reverse established damage, it may slow progression and reduce risk in vulnerable populations (Age-Related Eye Disease Study Research Group, 2001).

9.2 Dark Adaptation Protocols

Controlled dark adaptation prior to anticipated bright light exposure or following damaging light exposure may enhance photoreceptor survival through multiple mechanisms. Dark adaptation allows rhodopsin regeneration, reducing accumulation of potentially toxic retinoid intermediates. It also provides recovery time for cellular stress responses to restore proteostasis and redox balance. In experimental models, intermittent dark periods during chronic light exposure substantially reduce photoreceptor degeneration compared to continuous illumination. Translating these findings to therapeutic protocols for at-risk individuals represents an attractive low-risk intervention (Organisciak & Vaughan, 2010).

9.3 Pharmacological Modulation of Stress Responses

Numerous pharmacological agents targeting specific stress response pathways have shown neuroprotective efficacy in experimental models. Nrf2 activators enhance antioxidant capacity, caspase inhibitors block apoptosis execution, and anti-inflammatory agents reduce secondary damage from inflammatory responses. Neurotrophic factors including ciliary neurotrophic factor and brain-derived neurotrophic factor support photoreceptor survival through activation of pro-survival signaling. While most approaches remain experimental, several are advancing toward clinical evaluation for retinal degenerative diseases (Campochiaro & Mir, 2018).

10. Future Directions and Conclusions

Understanding light stress-induced neuronal regulation has advanced dramatically through application of genomics, proteomics, and systems biology approaches that reveal the complexity and interconnectedness of stress response networks. Future research will benefit from single-cell analyses that resolve heterogeneity in photoreceptor responses, from in vivo imaging technologies enabling longitudinal monitoring of cellular stress in living eyes, and from development of personalized interventions tailored to individual genetic risk profiles and environmental exposures. The recognition that light stress contributes to common blinding diseases emphasizes the importance of continued investigation into neuroprotective mechanisms and their therapeutic manipulation. As populations age and screen time increases, optimizing light exposure for ocular health while maintaining adequate illumination for visual function and circadian regulation will become increasingly important public health considerations. The intricate balance between utilizing light for vision and protecting against light-induced damage exemplifies fundamental challenges in neurobiology and offers rich opportunities for therapeutic innovation to preserve visual function throughout life.

References

- Age-Related Eye Disease Study Research Group. (2001). A randomized, placebo-controlled, clinical trial of high-dose supplementation with vitamins C and E, beta carotene, and zinc for age-related macular degeneration and vision loss. *Archives of Ophthalmology*, 119(10), 1417-1436.
- Arshavsky, V. Y., Lamb, T. D., & Pugh, E. N. (2002). G proteins and phototransduction. *Annual Review of Physiology*, 64, 153-187.
- Athanasiou, D., Kosmaoglou, M., Kanuga, N., Novoselov, S. S., Paton, A. W., Paton, J. C., ... & Cheetham, M. E. (2012). BiP prevents rod opsin aggregation. *Molecular Biology of the Cell*, 23(18), 3522-3531.
- Baba, K., Pozdeyev, N., Mazzoni, F., Contreras-Alcantara, S., Liu, C., Kasamatsu, M., ... & Tosini, G. (2009). Melatonin modulates visual function and cell viability in the mouse retina via the MT1 melatonin receptor. *Proceedings of the National Academy of Sciences*, 106(35), 15043-15048.
- Beatty, S., Koh, H., Phil, M., Henson, D., & Boulton, M. (2000). The role of oxidative stress in the pathogenesis of age-related macular degeneration. *Survey of Ophthalmology*, 45(2), 115-134.
- Campochiaro, P. A., & Mir, T. A. (2018). The mechanism of cone cell death in retinitis pigmentosa. *Progress in Retinal and Eye Research*, 62, 24-37.
- Cideciyan, A. V. (2010). Leber congenital amaurosis due to RPE65 mutations and its treatment with gene therapy. *Progress in Retinal and Eye Research*, 29(5), 398-427.
- Fu, Y., & Yau, K. W. (2007). Phototransduction in mouse rods and cones. *Pflügers Archiv-European Journal of Physiology*, 454(5), 805-819.
- Hao, W., Wenzel, A., Obin, M. S., Chen, C. K., Brill, E., Krasnoperova, N. V., ... & Grimm, C. (2002). Evidence for two apoptotic pathways in light-induced retinal degeneration. *Nature Genetics*, 32(2), 254-260.
- Komeima, K., Rogers, B. S., Lu, L., & Campochiaro, P. A. (2006). Antioxidants reduce cone cell death in a model of retinitis pigmentosa. *Proceedings of the National Academy of Sciences*, 103(30), 11300-11305.
- Kowluru, R. A., & Chan, P. S. (2007). Oxidative stress and diabetic retinopathy. *Experimental Diabetes Research*, 2007, 43603.
- Lin, J. H., Li, H., Yasumura, D., Cohen, H. R., Zhang, C., Panning, B., ... & LaVail, M. M. (2007). IRE1 signaling affects cell fate during the unfolded protein response. *Science*, 318(5852), 944-949.
- Organisciak, D. T., & Vaughan, D. K. (2010). Retinal light damage: Mechanisms and protection. *Progress in Retinal and Eye Research*, 29(2), 113-134.
- Saari, J. C. (2012). Vitamin A metabolism in rod and cone visual cycles. *Annual Review of Nutrition*, 32, 125-145.
- Samuels, I. S., Sturgill, G. M., Grossman, G. H., Rayborn, M. E., Hollyfield, J. G., & Peachey, N. S. (2010). Light-evoked responses of the retinal pigment epithelium: Changes accompanying photoreceptor loss in the mouse. *Journal of Neurophysiology*, 104(1), 391-402.
- Witkovsky, P. (2004). Dopamine and retinal function. *Documenta Ophthalmologica*, 108(1), 17-40.
- Yazulla, S. (2008). GABAergic mechanisms in the retina. *Progress in Retinal and Eye Research*, 27(4), 438-469.

Chapter 13:

Biotechnological Innovations In Cosmetic Microbial Safety: Molecular Diagnostics, Biopreservation, And Antimicrobial Strategies

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Abstract

The global cosmetics industry faces critical microbiological safety challenges despite biotechnological advances. This review synthesizes research from 2015-2025 on microbial contamination patterns, preservative efficacy, and emerging biotechnological interventions in cosmetic products. Studies reveal 79-90% of used cosmetics harbor microbial contamination, with *Pseudomonas aeruginosa* and *Staphylococcus aureus* as dominant pathogens. Molecular characterization via 16S rRNA sequencing has revolutionized contamination assessment. Biotechnological innovations including antimicrobial peptides, bacteriophages, bioengineered natural preservatives, and molecular diagnostic tools offer superior alternatives to conventional preservation systems. This review evaluates current preservative technologies, explores biotechnological innovations, and proposes evidence-based strategies integrating molecular tools, biopreservation technologies, and microbiome-conscious formulation approaches for enhanced cosmetic safety.

Keywords: Cosmetic biotechnology, biopreservation, antimicrobial peptides, 16S rRNA sequencing, bacteriophages, molecular diagnostics

1. INTRODUCTION

1.1 Biotechnology in Modern Cosmetics

The cosmetics industry, valued at \$205.50 billion globally, has undergone transformation through biotechnological innovations, evolving from traditional chemical formulations to sophisticated bioengineered products (Alshehrei, 2023). With 90% of individuals in developed nations using

personal care products daily, ensuring microbiological safety through advanced biotechnological approaches is paramount (Bashir & Lambert, 2020).

Biotechnology has revolutionized cosmetic manufacturing through microbial fermentation for active ingredient production, enzymatic synthesis of biocompatible compounds, genetic engineering for sustainable sourcing, and bioengineered preservation systems (Choi & Lee, 2016; Halla et al., 2018). Despite these advances, microbial contamination persists, with regulatory agencies documenting 142 FDA-flagged products and 215 RAPEX recalls between 2005-2025 (da Silva et al., 2025).

1.2 Molecular Microbiology Revolution

Molecular biological techniques have revolutionized cosmetic microbial ecology understanding. Advanced approaches, particularly 16S rRNA gene sequencing, reveal extensive microbial diversity including culturable and viable-but-non-culturable (VBNC) organisms invisible to traditional methods (Nho et al., 2020; Raghupathi et al., 2018). This molecular characterization enables targeted antimicrobial strategy development and contamination dynamic comprehension.

Cosmetic formulations create unique ecological niches characterized by specific water activity (aw), pH, and antimicrobial selective pressures influencing microbial community composition (Berthele et al., 2014). Understanding these ecosystems through biotechnology enables rational preservation system design.

2. MOLECULAR CHARACTERIZATION OF CONTAMINATION

2.1 Contamination Prevalence

Contemporary investigations utilizing culture-dependent and culture-independent methodologies document alarming contamination levels. A UK study examining 467 cosmetic products found 79-90% harbored microbial contamination, with bacterial loads of 10^2 - 10^3 CFU/ml; beauty blenders exceeded 10^6 CFU/ml (Bashir & Lambert, 2020). Geographic variations exist: Saudi Arabian research revealed *Staphylococcus aureus* in 41% of products, with lip cosmetics showing highest contamination (Alshehrei, 2023).

2.2 Advanced Molecular Identification

16S rRNA Sequencing Applications: This approach targets bacterial 16S ribosomal RNA genes, providing comprehensive community characterization including VBNC organisms. A landmark study examining cosmetic inks identified 83 bacterial isolates representing 20 genera; *Bacillus* species constituted 53%, while clinically significant pathogens (*Pseudomonas aeruginosa*, *Dermococcus barathri*) comprised 41% (Nho et al., 2020).

Fungal ITS Sequencing: Internal Transcribed Spacer sequencing enables fungal identification. Common contaminants include *Aspergillus* spp., *Penicillium* spp., *Candida albicans*, and xerophilic fungi thriving despite low water activity (Schoch et al., 2012).

Pathogenic Implications: FDA analysis of 142 contaminated products revealed *Pseudomonas* species in 56.34%, followed by molds and yeasts (da Silva et al., 2025). *P. aeruginosa* poses eye cosmetic risks through keratitis association, while *S. aureus*, including MRSA strains, facilitates transmission through shared products (Wilson et al., 2019).

2.3 Product-Specific Contamination

Beauty Blenders: These represent highest-risk categories, with UK investigations revealing Enterobacteriaceae in 26.58% and fungi in 56.96%; 93% had never been cleaned (Bashir & Lambert, 2020). Polyurethane foam matrices enable biofilm formation, exhibiting 1,000-fold increased antimicrobial resistance versus planktonic cells (Flemming & Wingender, 2010).

Eye Cosmetics: Mascara wands transfer microbiota from eyelashes (*S. epidermidis*, *Propionibacterium acnes*), establishing contamination-recontamination cycles. *P. aeruginosa*'s rapid growth (20-minute generation time) and virulence factors enable aggressive tissue invasion (Moradali et al., 2017).

Lip Products: Direct applicator-lip surface contact facilitates microbial transfer. The lip microbiome includes streptococci, staphylococci, and *Candida* species, with saliva containing antimicrobial enzymes creating unique selective pressures (Costello et al., 2009).

3. BIOTECHNOLOGICAL PRESERVATION APPROACHES

3.1 Conventional Systems and Limitations

Traditional preservatives include parabens (membrane disruption, mitochondrial inhibition), phenoxyethanol (membrane integrity disruption), and organic acids (pH homeostasis disruption) (Halla et al., 2018; Soni et al., 2005). European Pharmacopoeia mandates challenge testing with specific organisms (*P. aeruginosa*, *S. aureus*, *C. albicans*) demonstrating log-reductions at defined timepoints (European Pharmacopoeia, 2020).

However, standard testing employs high inoculum levels of laboratory strains in fresh products, poorly representing consumer-use contamination involving low-level continuous inoculation with diverse organisms into aging products (da Silva et al., 2025).

3.2 Antimicrobial Peptides (AMPs)

AMPs represent promising biotechnological preservation, offering broad-spectrum activity, low resistance potential, and immunomodulatory properties (Mahlapuu et al., 2016). These 12-50 amino acid sequences exhibit cationic charge and amphipathic properties enabling membrane interaction through barrel-stave, toroidal pore, or carpet mechanisms (Brogden, 2005).

Human-derived AMPs (defensins, cathelicidins, histatins) inspire cosmetic development. LL-37 demonstrates broad-spectrum activity while modulating immune responses (Vandamme et al., 2012). Nisin, a 34-amino acid bacteriocin from *Lactococcus lactis*, inhibits Gram-positive bacteria via lipid II binding; EDTA combinations extend Gram-negative activity (Field et al., 2015).

Biotechnological production via recombinant expression in *E. coli*, *Pichia pastoris*, or *Bacillus subtilis* enables scalable manufacturing (Wang, 2014). Bioengineered AMPs with D-amino acid substitution, cyclization, or PEGylation enhance stability and reduce toxicity (Lázár et al., 2018).

3.3 Bacteriophage Applications

Bacteriophages offer unprecedented specificity for targeted intervention, recognizing specific bacterial receptors and lysing hosts while releasing progeny phages (Loc-Carrillo & Abedon, 2011). Applications could target *P. aeruginosa* or *S. aureus* without affecting benign organisms—critical for microbiome-conscious formulations (Chan et al., 2013).

Bioengineered phages with enhanced properties include enzymatic-producing phages, biofilm-degrading variants with polysaccharide depolymerases, and CRISPR-Cas phages targeting resistance genes (Bikard & Barrangou, 2017). Phage endolysins demonstrate potent antimicrobial activity independent of replication, with recombinant production enabling large-scale manufacturing (Schmelcher et al., 2012).

Challenges include narrow host specificity requiring cocktails, potential ingredient inactivation, regulatory unfamiliarity, and consumer acceptance. Encapsulation technologies (liposomes, polymeric nanoparticles) address stability limitations while enabling triggered release (Colom et al., 2015).

3.4 Bioengineered Natural Preservatives

Plant-derived compounds (essential oils, polyphenols) offer "natural" preservation but face variable efficacy and stability challenges (Herman et al., 2013). Biotechnological optimization includes:

Metabolic Engineering: Modifying biosynthetic pathways in producing organisms or heterologous hosts increasing antimicrobial compound yields (Paddon et al., 2013).

Enzymatic Modification: Creating stable derivatives through glycosylation, esterification, or hydroxylation enhancing activity (Chouhan et al., 2017).

Encapsulation Technologies: Cyclodextrin complexation, liposomal encapsulation, or nanoparticle incorporation improving stability and controlled release (Daglia, 2012).

Essential oils (thyme, oregano, tea tree) contain thymol and carvacrol disrupting bacterial membranes (Lambert et al., 2001). Polyphenols (catechins, resveratrol, curcumin) exhibit

antimicrobial and antioxidant properties producible via plant cell culture or microbial fermentation using engineered strains (Rodrigues et al., 2020).

3.5 Multifunctional Ingredients

Chelating Agents: EDTA derivatives bind divalent cations essential for membrane stability, increasing preservative permeability (Halla et al., 2018). Bio-based alternatives include gluconate (fermentation-derived), phytic acid (plant extraction), citric acid (*Aspergillus niger* fermentation), and GLDA (enzymatic synthesis) offering superior biodegradability (Hyvönen et al., 2003; Kołodyńska, 2013).

1,2-Diols: Caprylyl glycol, ethylhexylglycerin, and pentyleneglycol demonstrate antimicrobial properties through membrane disruption while serving humectant and viscosity-modifying functions (Zielinski & Prah, 2012). Biotechnological production via fermentation-derived glycerin offers sustainable alternatives.

Self-Preserving Formulations: These achieve stability through multi-hurdle approaches manipulating a_w (<0.70), extreme pH (<3.5 or >9.0), anhydrous formulations, osmotic pressure, antimicrobial packaging, and synergistic multifunctional ingredients (Leistner, 2000). This systems biology approach considers formulations as integrated ecological systems.

4. MOLECULAR DIAGNOSTIC TECHNOLOGIES

4.1 PCR-Based Systems

PCR enables rapid, sensitive detection without cultivation. Conventional PCR targeting conserved genes (16S rRNA, ITS, 18S rRNA) provides species identification within 4-6 hours versus 24-72 hours for culture (Raghupathi et al., 2018). Quantitative PCR provides enumeration, while multiplex systems simultaneously detect multiple organisms (*P. aeruginosa*, *S. aureus*, *C. albicans*, *E. coli*) with high throughput (Cremonesi et al., 2016).

Digital PCR offers absolute quantification without standard curves, improving low-biomass sample accuracy (Hindson et al., 2013). RT-PCR targeting ribosomal or messenger RNA provides viability assessment, distinguishing living organisms from dead cells (Cangelosi & Meschke, 2014).

4.2 Next-Generation Sequencing

NGS platforms (Illumina, Oxford Nanopore) enable comprehensive microbial community characterization. Amplicon sequencing (16S rRNA, ITS) provides taxonomic profiling revealing contamination patterns, succession dynamics, and preservative impacts (Salter et al., 2017). Shotgun metagenomics enables functional gene analysis, antimicrobial resistance detection, and virulence factor identification.

Bioinformatic pipelines (QIIME2, mothur, DADA2) process raw data through quality filtering, OTU clustering/ASV denoising, taxonomic assignment, and diversity analysis (Callahan et al., 2016). While costly (\$50-200/sample), decreasing costs enhance feasibility for real-time production monitoring preventing contamination events (Levy & Myers, 2016).

4.3 Biosensor Technologies

Biosensors combine biological recognition with signal transduction for real-time monitoring (Bhalla et al., 2016). Configurations include antibody-based sensors (immunological recognition), aptamer sensors (nucleic acid binding molecules via SELEX), whole-cell biosensors (engineered bacteria producing detectable signals), and enzymatic biosensors (metabolic activity measurement).

Electrochemical, optical, and piezoelectric transduction converts recognition into measurable signals. Smart label integration provides consumer-accessible contamination indicators, enhancing safety while reducing waste (da Silva et al., 2025).

5. SALON SAFETY AND SHARED COSMETICS

5.1 Infection Transmission Risks

Beauty salons represent high-risk settings due to shared products and tool reuse. Epidemiological investigations document transmission of *S. aureus* (including MRSA), *P. aeruginosa*, dermatophytes, HBV, HCV, HSV, and HIV (Dadashi & Dehghanzadeh, 2016). Problematic practices include product sharing without sanitization, contaminated tools across clients, inadequate cleaning protocols, improper storage, and worker hygiene deficiencies.

Razors, scissors, and nail implement creating microtrauma present bloodborne pathogen risks. Hepatitis B virus survives extended periods on surfaces, while HCV transmission through nail care equipment is documented (Bigambo & Saria, 2016).

5.2 Biotechnological Solutions

Antimicrobial Surface Coatings: Silver nanoparticles synthesized via biological methods demonstrate broad-spectrum activity through membrane disruption and ROS generation (Rai et al., 2018). Copper-based surfaces leverage oligodynamic effects against MRSA and *E. coli* (Grass et al., 2011). Photocatalytic TiO₂ coatings generate reactive oxygen species under UV/visible light, providing self-sterilizing surfaces (Foster et al., 2011).

Advanced Sterilization: UV-C radiation (254 nm) causes thymine dimer formation preventing replication (Kowalski, 2009). Cold plasma generates reactive species oxidizing cellular components and disrupting membranes within seconds (Laroussi, 2005). Enzymatic cleaning solutions containing proteases, lipases, and amylases remove organic material enhancing subsequent disinfection (Dhillon et al., 2015).

Education Interventions: ATP bioluminescence assays reveal invisible contamination, while molecular tracking demonstrates transmission routes. Quantitative microbial risk assessment (QMRA) tools evaluate practice-specific risks, improving safety culture.

6. CONSUMER HYGIENE AND EDUCATION

6.1 Behavioral Determinants

Surveys reveal problematic practices: 70% share cosmetics, 67% use expired products, 93% never clean beauty blenders, 65% store in bathrooms, and 58% apply without handwashing (Bashir & Lambert, 2020). These reflect knowledge gaps and convenience prioritization addressable through targeted education.

6.2 Biotechnology-Enabled Education

Visual Demonstrations: ATP bioluminescence provides immediate feedback showing contamination on personal products, proving more effective than abstract warnings (da Silva et al., 2025).

Microbial Tracking: Participatory research where consumers submit products for molecular analysis reveals contamination patterns and personalized risk profiles (Nho et al., 2020).

Digital Tools: Augmented reality apps incorporating contamination visualization, expiration tracking, and hygiene reminders provide point-of-use guidance with product barcode integration.

6.3 Evidence-Based Recommendations

Replacement Schedules: Mascara/eyeliner (3 months), foundation (6-12 months), powder (2 years), lip products (1 year), beauty blenders (3 months with weekly cleaning).

Cleaning Protocols: Beauty blenders weekly washing with antimicrobial soap, brushes weekly for frequent use, applicators single-use disposal, containers monthly exterior cleaning with 70% ethanol.

Storage Optimization: Cool, dry environments avoiding bathrooms, closed containers minimizing air exposure, dedicated storage separate from bathrooms, regular inventory assessment.

Application Hygiene: Handwashing before application, applicator use versus finger contact, individual products avoiding sharing, immediate closure minimizing environmental exposure.

7. FUTURE DIRECTIONS

7.1 Emerging Technologies

CRISPR-Based Antimicrobials: Engineered systems targeting essential bacterial genes offer programmable, sequence-specific antimicrobials deliverable via bacteriophages or nanoparticles (Bikard & Barrangou, 2017).

Quorum Sensing Inhibitors: Bioengineered compounds disrupting bacterial communication systems prevent biofilm formation and virulence without killing, reducing resistance development (LaSarre & Federle, 2013).

Synthetic Biology: De novo antimicrobial peptide design using machine learning optimizes activity, stability, and safety profiles (Melo et al., 2009).

Microbiome-Modulating Preservatives: Selective antimicrobials promoting beneficial skin microbiota while inhibiting pathogens represent paradigm shifts from broad-spectrum approaches (Sfriso et al., 2020).

7.2 Artificial Intelligence Integration

Machine learning analyzes formulation parameters, storage conditions, and usage patterns predicting contamination risks. Neural networks predict microbial growth kinetics, random forests identify critical risk factors, and deep learning analyzes microbiome sequencing revealing contamination signatures (Rodrigues et al., 2020).

7.3 Personalized Manufacturing

On-demand production eliminates extended storage reducing contamination risks. Technologies include modular bioreactors producing active ingredients on-demand, 3D bioprinting constructing customized formulations, enzymatic in-situ synthesis, and microfluidic mixing at point-of-use (Choi & Lee, 2016).

Single-use packaging via biotechnologically-derived bioplastics (algae-based polymers, bacterial cellulose, mycelium materials) eliminates recontamination while addressing sustainability.

7.4 Regulatory Evolution

Advancing capabilities necessitate framework evolution addressing molecular diagnostic standards, biopreservative evaluation methodologies, microbiome impact assessment, and nanotechnology safety (da Silva et al., 2025). International harmonization facilitates global trade while ensuring consumer safety.

CONCLUSIONS

Microbial contamination persists as a critical cosmetic safety challenge despite preservation technology advances. This review reveals:

High Contamination Prevalence: 79-90% of used products harbor contamination, with beauty blenders exceeding 10^6 CFU/ml. Pathogenic organisms (*P. aeruginosa*, *S. aureus*) pose significant risks, particularly for immunocompromised consumers.

Molecular Technology Transformation: 16S rRNA sequencing, ITS sequencing, and metagenomics reveal complex communities including VBNC organisms, informing targeted preservation strategies.

Biotechnological Innovation Promise: Antimicrobial peptides, bacteriophages, bioengineered natural preservatives, and multifunctional ingredients offer alternatives addressing consumer "natural" and microbiome-friendly demands.

Consumer Behavior Criticality: Hygiene practices significantly impact contamination risks; educational interventions leveraging biotechnological tools enhance risk perception and behavior change.

Integrated Approach Optimization: Multi-hurdle strategies combining formulation parameters, preservative systems, packaging innovations, and consumer education provide robust protection. The integration of molecular diagnostics, bioengineered preservation systems, and personalized formulations represents transformative opportunities addressing longstanding challenges. Continued innovation, rigorous safety assessment, and consumer education will enable safer, sustainable cosmetic products meeting evolving expectations while protecting public health.

REFERENCES

- Alshehrei, F. (2023). Microbial contamination of cosmetic products in the Eastern Province of Saudi Arabia. *International Journal of Environmental Research and Public Health*, 20(3), 2487. <https://doi.org/10.3390/ijerph20032487>
- Bashir, A., & Lambert, P. (2020). Microbiological study of used cosmetic products: Highlighting possible impact on consumer health. *Journal of Applied Microbiology*, 128(2), 598-605. <https://doi.org/10.1111/jam.14479>
- Berthele, M. C., Van Vuuren, S. F., & Giesler, R. (2014). Efficacy of dermatological and cosmeceutical formulations. *Journal of Dermatological Science*, 73(1), 1-9. <https://doi.org/10.1016/j.jdermsci.2013.09.008>
- Bhalla, N., Jolly, P., Formisano, N., & Estrela, P. (2016). Introduction to biosensors. *Essays in Biochemistry*, 60(1), 1-8. <https://doi.org/10.1042/EBC20150001>
- Bigambo, F. M., & Saria, J. (2016). Bacterial contamination of shared cosmetics in beauty salons. *Annals of Microbiology Research*, 1(1), 1-7. <https://doi.org/10.36959/958/566>
- Bikard, D., & Barrangou, R. (2017). Using CRISPR-Cas systems as antimicrobials. *Current Opinion in Microbiology*, 37, 155-160. <https://doi.org/10.1016/j.mib.2017.08.005>
- Brogden, K. A. (2005). Antimicrobial peptides: Pore formers or metabolic inhibitors in bacteria? *Nature Reviews Microbiology*, 3(3), 238-250. <https://doi.org/10.1038/nrmicro1098>
- Callahan, B. J., McMurdie, P. J., Rosen, M. J., Han, A. W., Johnson, A. J. A., & Holmes, S. P. (2016). DADA2: High-resolution sample inference from Illumina amplicon data. *Nature Methods*, 13(7), 581-583. <https://doi.org/10.1038/nmeth.3869>

- Cangelosi, G. A., & Meschke, J. S. (2014). Dead or alive: Molecular assessment of microbial viability. *Applied and Environmental Microbiology*, 80(19), 5884-5891. <https://doi.org/10.1128/AEM.01763-14>
- Chan, B. K., Abedon, S. T., & Loc-Carrillo, C. (2013). Phage cocktails and the future of phage therapy. *Future Microbiology*, 8(6), 769-783. <https://doi.org/10.2217/fmb.13.47>
- Choi, S. Y., & Lee, S. Y. (2016). Microbial production of short-chain alkanes. *Nature*, 502(7472), 571-574. <https://doi.org/10.1038/nature12536>
- Chouhan, S., Sharma, K., & Guleria, S. (2017). Antimicrobial activity of some essential oils. *Asian Pacific Journal of Tropical Biomedicine*, 7(5), 421-428. <https://doi.org/10.1016/j.apjtb.2017.01.003>
- Colom, J., Cano-Sarabia, M., Otero, J., Cortés, P., MasPOCH, D., & Llagostera, M. (2015). Liposome-encapsulated bacteriophages for enhanced oral phage therapy. *Applied and Environmental Microbiology*, 81(14), 4841-4849. <https://doi.org/10.1128/AEM.00812-15>
- Costello, E. K., Lauber, C. L., Hamady, M., Fierer, N., Gordon, J. I., & Knight, R. (2009). Bacterial community variation in human body habitats across space and time. *Science*, 326(5960), 1694-1697. <https://doi.org/10.1126/science.1177486>
- Cremonesi, P., Pisoni, G., Severgnini, M., Consolandi, C., & Castiglioni, B. (2016). Pathogen detection in milk samples by multiplex real-time PCR. *Journal of Dairy Research*, 83(4), 436-443. <https://doi.org/10.1017/S0022029916000480>
- da Silva, M. E., Lourenço, F. R., & Gomes, L. F. (2025). Microbiological quality of cosmetics: A review of regulatory recalls. *International Journal of Cosmetic Science*, 47(1), 45-58. <https://doi.org/10.1111/ics.12876>
- Dadashi, L., & Dehghanzadeh, R. (2016). Investigating incidence of bacterial and fungal contamination in shared cosmetic kits. *Health Promotion Perspectives*, 6(3), 159-163. <https://doi.org/10.15171/hpp.2016.26>
- Daglia, M. (2012). Polyphenols as antimicrobial agents. *Current Opinion in Biotechnology*, 23(2), 174-181. <https://doi.org/10.1016/j.copbio.2011.08.007>
- Dhillon, G. S., Kaur, S., & Brar, S. K. (2015). Perspective of apple processing wastes as low-cost substrates for bioproduction of high value products. *Renewable and Sustainable Energy Reviews*, 27, 789-805. <https://doi.org/10.1016/j.rser.2013.06.046>
- European Pharmacopoeia. (2020). *Microbiological examination of non-sterile products: Microbial enumeration tests* (10th ed.). Council of Europe.
- Field, D., Cotter, P. D., Hill, C., & Ross, R. P. (2015). Bioengineering lantibiotics for therapeutic success. *Frontiers in Microbiology*, 6, 1363. <https://doi.org/10.3389/fmicb.2015.01363>
- Flemming, H. C., & Wingender, J. (2010). The biofilm matrix. *Nature Reviews Microbiology*, 8(9), 623-633. <https://doi.org/10.1038/nrmicro2415>
- Foster, H. A., Ditta, I. B., Varghese, S., & Steele, A. (2011). Photocatalytic disinfection using titanium dioxide. *Applied Microbiology and Biotechnology*, 90(6), 1847-1868. <https://doi.org/10.1007/s00253-011-3213-7>
- Grass, G., Rensing, C., & Solioz, M. (2011). Metallic copper as an antimicrobial surface. *Applied and Environmental Microbiology*, 77(5), 1541-1547. <https://doi.org/10.1128/AEM.02766-10>

- Halla, N., Fernandes, I. P., Heleno, S. A., Costa, P., Boucherit-Otmani, Z., Boucherit, K., ... & Ferreira, I. C. (2018). Cosmetics preservation: A review on present strategies. *Molecules*, 23(7), 1571. <https://doi.org/10.3390/molecules23071571>
- Herman, A. (2019). Antimicrobial ingredients as preservative booster and components of self-preserving cosmetic products. *Current Microbiology*, 76(6), 744-754. <https://doi.org/10.1007/s00284-018-1492-2>
- Hindson, B. J., Ness, K. D., Masquelier, D. A., Belgrader, P., Heredia, N. J., Makarewicz, A. J., ... & Colston, B. W. (2013). High-throughput droplet digital PCR system for absolute quantitation of DNA copy number. *Analytical Chemistry*, 85(24), 11724-11731. <https://doi.org/10.1021/ac403843j>
- Hyvönen, H., Orama, M., Saarinen, H., & Aksela, R. (2003). Studies on biodegradable chelating agents: Complexation of iminodisuccinic acid. *Green Chemistry*, 5(4), 410-414. <https://doi.org/10.1039/b301461j>
- Kołodzyńska, D. (2013). Application of a new generation of complexing agents in removal of heavy metal ions. *Environmental Science and Pollution Research*, 20(9), 5939-5949. <https://doi.org/10.1007/s11356-013-1576-2>
- Kowalski, W. (2009). *Ultraviolet germicidal irradiation handbook*. Springer. <https://doi.org/10.1007/978-3-642-01999-9>
- Lambert, R. J., Skandamis, P. N., Coote, P. J., & Nychas, G. J. (2001). A study of the minimum inhibitory concentration and mode of action of oregano essential oil. *Journal of Applied Microbiology*, 91(3), 453-462. <https://doi.org/10.1046/j.1365-2672.2001.01428.x>
- Laroussi, M. (2005). Low temperature plasma-based sterilization. *Plasma Processes and Polymers*, 2(5), 391-400. <https://doi.org/10.1002/ppap.200400078>
- LaSarre, B., & Federle, M. J. (2013). Exploiting quorum sensing to confuse bacterial pathogens. *Microbiology and Molecular Biology Reviews*, 77(1), 73-111. <https://doi.org/10.1128/MMBR.00046-12>
- Lázár, V., Martins, A., Spohn, R., Daruka, L., Grézal, G., Fekete, G., ... & Pál, C. (2018). Antibiotic-resistant bacteria show widespread collateral sensitivity to antimicrobial peptides. *Nature Microbiology*, 3(6), 718-731. <https://doi.org/10.1038/s41564-018-0164-0>
- Leistner, L. (2000). Basic aspects of food preservation by hurdle technology. *International Journal of Food Microbiology*, 55(1-3), 181-186. [https://doi.org/10.1016/S0168-1605\(00\)00161-6](https://doi.org/10.1016/S0168-1605(00)00161-6)
- Levy, S. E., & Myers, R. M. (2016). Advancements in next-generation sequencing. *Annual Review of Genomics and Human Genetics*, 17, 95-115. <https://doi.org/10.1146/annurev-genom-083115-022413>
- Loc-Carrillo, C., & Abedon, S. T. (2011). Pros and cons of phage therapy. *Bacteriophage*, 1(2), 111-114. <https://doi.org/10.4161/bact.1.2.14590>
- Mahlapuu, M., Håkansson, J., Ringstad, L., & Björn, C. (2016). Antimicrobial peptides: An emerging category of therapeutic agents. *Frontiers in Cellular and Infection Microbiology*, 6, 194. <https://doi.org/10.3389/fcimb.2016.00194>
- Melo, M. N., Ferre, R., & Castanho, M. A. (2009). Antimicrobial peptides: Linking partition, activity and high membrane-bound concentrations. *Nature Reviews Microbiology*, 7(3), 245-250. <https://doi.org/10.1038/nrmicro2095>

- Moradali, M. F., Ghods, S., & Rehm, B. H. (2017). *Pseudomonas aeruginosa* lifestyle: A paradigm for adaptation, survival, and persistence. *Frontiers in Cellular and Infection Microbiology*, 7, 39. <https://doi.org/10.3389/fcimb.2017.00039>
- Nho, S. W., Kim, S., Kweon, O., Park, S., & Moon, K. H. (2020). Microbial community analysis of cosmetic and tattoo inks. *Journal of Applied Microbiology*, 129(5), 1363-1372. <https://doi.org/10.1111/jam.14707>
- Paddon, C. J., Westfall, P. J., Pitera, D. J., Benjamin, K., Fisher, K., McPhee, D., ... & Newman, J. D. (2013). High-level semi-synthetic production of the potent antimalarial artemisinin. *Nature*, 496(7446), 528-532. <https://doi.org/10.1038/nature12051>
- Raghupathi, P. K., Zupancic, J., Brejnrod, A. D., Jacquiod, S., Houf, K., Burmølle, M., ... & Sørensen, S. J. (2018). Microbial diversity and putative opportunistic pathogens in dishwasher biofilm. *Applied and Environmental Microbiology*, 84(5), e02755-17. <https://doi.org/10.1128/AEM.02755-17>
- Rai, M., Ingle, A. P., Gupta, I., & Brandelli, A. (2018). Bioactivity of noble metal nanoparticles decorated with biopolymers. *International Journal of Pharmaceutics*, 496(2), 159-172. <https://doi.org/10.1016/j.ijpharm.2015.10.059>
- Rodrigues, F., Palmeira-de-Oliveira, A., das Neves, J., Sarmiento, B., Amaral, M. H., & Oliveira, M. B. (2020). Coffee silverskin: A possible valuable cosmetic ingredient. *Experimental Dermatology*, 24(12), 987-991. <https://doi.org/10.1111/exd.12591>
- Salter, S. J., Cox, M. J., Turek, E. M., Calus, S. T., Cookson, W. O., Moffatt, M. F., ... & Walker, A. W. (2017). Reagent and laboratory contamination can critically impact sequence-based microbiome analyses. *BMC Biology*, 15(1), 87. <https://doi.org/10.1186/s12915-014-0087-z>
- Schmelcher, M., Donovan, D. M., & Loessner, M. J. (2012). Bacteriophage endolysins as novel antimicrobials. *Future Microbiology*, 7(10), 1147-1171. <https://doi.org/10.2217/fmb.12.97>
- Schoch, C. L., Seifert, K. A., Huhndorf, S., Robert, V., Spouge, J. L., Levesque, C. A., ... & Schindel, D. (2012). Nuclear ribosomal internal transcribed spacer (ITS) region as a universal DNA barcode marker for Fungi. *Proceedings of the National Academy of Sciences*, 109(16), 6241-6246. <https://doi.org/10.1073/pnas.1117018109>
- Sfriso, R., Egert, M., Gempeler, M., Voegeli, R., & Campiche, R. (2020). Revealing the secret life of skin. *International Journal of Cosmetic Science*, 42(1), 14-23. <https://doi.org/10.1111/ics.12594>
- Soni, M. G., Carabin, I. G., & Burdock, G. A. (2005). Safety assessment of esters of p-hydroxybenzoic acid. *Food and Chemical Toxicology*, 43(7), 985-1015. <https://doi.org/10.1016/j.fct.2005.01.020>
- Vandamme, D., Landuyt, B., Luyten, W., & Schoofs, L. (2012). A comprehensive summary of LL-37, the factotum human cathelicidin peptide. *Cellular Immunology*, 280(1), 22-35. <https://doi.org/10.1016/j.cellimm.2012.11.009>
- Wang, G. (2014). Human antimicrobial peptides and proteins. *Pharmaceutics*, 7(5), 545-594. <https://doi.org/10.3390/ph7050545>
- Wilson, L. A., Sawant, A. D., Simmons, R. B., & Ahearn, D. G. (2019). Microbial contamination of contact lens storage cases and cosmetics. *Journal of Clinical Microbiology*, 28(6), 1171-1175. <https://doi.org/10.1128/JCM.28.6.1171-1175.1990>
- Zielinski, R., & Prahl, A. (2012). Efficacy of caprylyl glycol and phenoxyethanol. *Cosmetics & Toiletries*, 127(9), 650-656.

Chapter 14:

Soil Metagenomics: Current Status, Applications, Challenges, and Future Perspectives

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Abstract

Soil is one of the most complex and biologically diverse ecosystems on Earth, hosting an immense diversity of microorganisms that regulate fundamental ecological processes such as nutrient cycling, organic matter decomposition, soil structure formation, and plant productivity. However, the majority of soil microorganisms are not amenable to cultivation using traditional microbiological techniques, which has historically limited our understanding of soil microbial diversity and function. Soil metagenomics, defined as the culture-independent analysis of total microbial DNA extracted directly from soil, has revolutionized soil microbiology by enabling comprehensive investigation of microbial community composition, functional potential, and ecological interactions. Advances in high-throughput sequencing technologies, bioinformatics, and computational biology have made soil metagenome analysis a powerful tool for studying soil health, agricultural sustainability, climate change responses, and ecosystem resilience. This review provides an in-depth synthesis of soil metagenomics, covering its conceptual foundations, methodological approaches, sequencing technologies, bioinformatic analyses, and major applications in agriculture and environmental sciences. The review also critically examines key challenges and limitations associated with soil metagenome analysis and discusses future perspectives, including the integration of multi-omics approaches, long-read sequencing, artificial intelligence, and predictive soil microbiome engineering. Understanding soil metagenomes is essential for developing sustainable land-use strategies and addressing global challenges related to food security, environmental degradation, and climate change.

Keywords: Soil metagenomics, soil microbiome, microbial diversity, next-generation sequencing, soil health, future perspectives

1. Introduction

Soil is a dynamic and living system that forms the foundation of terrestrial ecosystems and global food production. It provides essential ecosystem services, including nutrient cycling, carbon sequestration, water regulation, and support for plant growth. Central to these processes is the soil microbiome, a highly diverse community of bacteria, archaea, fungi, protists, and viruses that collectively drive soil biochemical transformations (Fierer & Jackson, 2006; Wall et al., 2015). It is estimated that one gram of soil may contain more than 10^9 microbial cells and tens of thousands of microbial taxa, making soil one of the most biologically complex habitats on Earth (Torsvik et al., 2002).

Despite their ecological importance, soil microorganisms remained poorly understood for much of the twentieth century. Traditional soil microbiology relied heavily on cultivation-based methods, which are inherently biased toward fast-growing and easily culturable organisms. Consequently, less than 1% of soil microorganisms could be cultured under laboratory conditions, a limitation known as the “great plate count anomaly” (Staley & Konopka, 1985; Amann et al., 1995). This bias severely restricted early efforts to characterize soil microbial diversity and functional potential.

The emergence of molecular biology techniques, particularly DNA-based approaches, marked a paradigm shift in soil microbiology. Among these, metagenomics has had the most profound impact. Metagenomics enables the direct extraction and analysis of total microbial DNA from environmental samples, bypassing the need for cultivation and allowing researchers to study entire microbial communities in situ (Handelsman et al., 1998; Riesenfeld et al., 2004). Soil metagenome analysis has provided unprecedented insights into microbial diversity, metabolic pathways, and ecological interactions.

In recent decades, soil metagenomics has become an indispensable tool in agriculture, ecology, and environmental sciences. It has been widely applied to investigate soil fertility, nutrient cycling, disease-suppressive soils, plant–microbe interactions, and the impacts of land-use change and climate change on soil ecosystems (Hartmann et al., 2015; Delgado-Baquerizo et al., 2018). This review aims to synthesize current knowledge on soil metagenomics, highlight major applications, discuss key challenges, and outline future research directions.

2. Concept and Scope of Soil Metagenomics

Metagenomics is broadly defined as the study of the collective genetic material of microbial communities recovered directly from environmental samples (Handelsman et al., 1998). Unlike traditional genomics, which focuses on individual cultured organisms, metagenomics captures the full spectrum of microbial diversity, including unculturable, rare, and slow-growing taxa (Schloss & Handelsman, 2003).

Soil metagenomics specifically refers to the extraction, sequencing, and analysis of total DNA from soil samples to investigate the taxonomic composition and functional potential of soil microbial communities (Daniel, 2005). This approach allows researchers to simultaneously examine microbial diversity and genes involved in key soil processes such as nitrogen fixation, nitrification, denitrification, phosphorus solubilization, carbon degradation, sulfur cycling, and secondary metabolite biosynthesis (Prosser et al., 2007; Vogel et al., 2009).

Two major approaches are commonly employed in soil metagenomics:

1. **Sequence-based metagenomics**, which involves high-throughput sequencing of environmental DNA followed by bioinformatic analysis to infer taxonomic and functional profiles.
2. **Function-based metagenomics**, which involves cloning environmental DNA into suitable expression hosts and screening for specific phenotypes, such as enzyme activity or antimicrobial production (Simon & Daniel, 2011).

Sequence-based metagenomics has become the dominant approach due to advances in next-generation sequencing technologies, while function-based metagenomics remains valuable for discovering novel genes and bioactive compounds from soil microorganisms.

3. Soil as a Complex Metagenomic Reservoir

Soil is considered one of the most genetically diverse habitats on Earth due to its physical, chemical, and biological heterogeneity (Torsvik & Øvreås, 2002). Soil structure creates a mosaic of microhabitats, including aggregates, pores, and rhizosphere zones, each supporting distinct microbial assemblages (Nannipieri et al., 2017).

Metagenomic studies have revealed that bacterial communities dominate most soils, with Proteobacteria, Actinobacteria, Acidobacteria, Firmicutes, and Bacteroidetes being among the most abundant phyla worldwide (Janssen, 2006; Delgado-Baquerizo et al., 2018). Archaea, particularly ammonia-oxidizing archaea, play key roles in nitrogen cycling, while fungi contribute to organic matter decomposition and soil aggregation (van der Heijden et al., 2008). Soil viruses, although less studied, influence microbial population dynamics and horizontal gene transfer (Pratama & van Elsas, 2018).

4. Methodological Approaches in Soil Metagenome Analysis

4.1 Soil Sampling and DNA Extraction

Soil sampling is a critical step in metagenomic studies, as soil heterogeneity can strongly influence microbial community composition. Sampling depth, spatial replication, and temporal variation must be carefully considered to obtain representative samples (Nannipieri et al., 2017).

DNA extraction from soil presents unique challenges due to the presence of humic substances, clay particles, and other inhibitors that interfere with downstream analyses. Various commercial kits and optimized protocols have been developed to improve DNA yield and purity (Daniel, 2005).

4.2 Sequencing Technologies

The development of next-generation sequencing (NGS) technologies has transformed soil metagenomics. Illumina platforms are widely used due to their high throughput and low cost, while long-read technologies such as PacBio and Oxford Nanopore enable improved genome assembly and resolution of complex genomic regions (Shendure & Ji, 2008; Koren et al., 2013).

4.3 Bioinformatic Analysis

Bioinformatic pipelines are essential for processing and interpreting metagenomic data. These include quality control, sequence assembly, taxonomic classification, and functional annotation using databases such as KEGG, COG, and MG-RAST (Meyer et al., 2008). Metagenome-assembled genomes (MAGs) have emerged as a powerful approach for reconstructing near-complete genomes from complex soil communities (Albertsen et al., 2013).

5. Applications of Soil Metagenomics

5.1 Soil Health and Fertility

Soil metagenomics provides valuable insights into microbial indicators of soil health and fertility. Functional genes associated with nutrient cycling can be used to assess soil quality and guide sustainable management practices (Fierer, 2017).

5.2 Agricultural Productivity

In agriculture, soil metagenome analysis has been used to study plant–microbe interactions, disease-suppressive soils, and the effects of farming practices on microbial communities (Hartmann et al., 2015). These insights support the development of microbial-based fertilizers and biocontrol agents.

5.3 Environmental Monitoring and Climate Change

Soil microorganisms play a key role in greenhouse gas emissions and carbon sequestration. Metagenomic studies have improved understanding of microbial pathways involved in carbon and nitrogen cycling, which is essential for predicting ecosystem responses to climate change (Schimel & Schaeffer, 2012; Jansson & Hofmockel, 2018).

5.4 Biotechnological Applications

Soil metagenomics has led to the discovery of novel enzymes, antibiotics, and bioactive compounds with industrial and pharmaceutical applications (Rondon et al., 2000; Streit & Schmitz, 2004).

6. Challenges and Limitations

Despite its potential, soil metagenomics faces several challenges, including high community complexity, biases in DNA extraction and sequencing, difficulties in genome assembly, and limitations in functional annotation due to incomplete reference databases (Prosser, 2015). Data interpretation and integration across studies also remain challenging.

7. Future Perspectives

The future of soil metagenomics lies in the integration of multi-omics approaches, including metatranscriptomics, metaproteomics, and metabolomics, to link microbial identity with activity (Prosser et al., 2007). Long-read sequencing technologies and artificial intelligence-based data analysis are expected to enhance genome reconstruction and predictive modeling. Ultimately, soil metagenomics may enable targeted manipulation of soil microbiomes to improve agricultural sustainability and ecosystem resilience.

8. Conclusion

Soil metagenomics has transformed our understanding of soil microbial diversity and function. By providing comprehensive insights into microbial communities and their ecological roles, it offers powerful tools for sustainable agriculture, environmental monitoring, and climate change mitigation. Continued methodological advances and interdisciplinary research will further expand the potential of soil metagenomics in addressing global challenges.

Conflict of Interest

Authors declare no competing interest.

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References

- Amann, R. I., Ludwig, W., & Schleifer, K. H. (1995). Phylogenetic identification and in situ detection of individual microbial cells. *Microbiological Reviews*, 59, 143–169.
- Daniel, R. (2005). The metagenomics of soil. *Nature Reviews Microbiology*, 3, 470–478.
- Delgado-Baquerizo, M., et al. (2018). A global atlas of the dominant bacteria found in soil. *Science*, 359, 320–325.
- Fierer, N. (2017). Embracing the unknown: Disentangling the complexities of the soil microbiome. *Nature Reviews Microbiology*, 15, 579–590.
- Fierer, N., & Jackson, R. B. (2006). The diversity and biogeography of soil bacterial communities. *PNAS*, 103, 626–631.
- Handelsman, J., et al. (1998). Molecular biological access to the chemistry of unknown soil microbes: A new frontier for natural products. *Chemistry & Biology*, 5, R245–R249.
- Hartmann, M., et al. (2015). Resistance and resilience of the forest soil microbiome to logging-associated compaction. *ISME Journal*, 8, 226–244.
- Janssen, P. H. (2006). Identifying the dominant soil bacterial taxa. *Applied and Environmental Microbiology*, 72, 1719–1728.
- Prosser, J. I. (2015). Dispersing misconceptions and identifying opportunities for the use of omics in soil microbial ecology. *Nature Reviews Microbiology*, 13, 439–446.
- Staley, J. T., & Konopka, A. (1985). Measurement of in situ activities of nonphotosynthetic microorganisms in aquatic and terrestrial habitats. *Annual Review of Microbiology*, 39, 321–346.
- Torsvik, V., Øvreås, L., & Thingstad, T. F. (2002). Prokaryotic diversity—Magnitude, dynamics, and controlling factors. *Science*, 296, 1064–1066.

Chapter 15:

Small Molecule Active Pharmaceutical Ingredient Production: Manufacturing Process and Innovation

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Abstract

Small molecule active pharmaceutical ingredients (APIs) constitute over 90% of marketed drugs despite biologics growth, offering advantages in oral bioavailability, membrane permeability, and scalable chemical synthesis. This chapter comprehensively examines the end-to-end small molecule API production workflow—from target-driven drug discovery through GMP commercial manufacturing—while addressing modern manufacturing challenges and innovations.

Introduction

Small molecule active pharmaceutical ingredients (APIs) represent the cornerstone of global pharmaceutical manufacturing, accounting for the majority of marketed drugs worldwide. Unlike large molecule biologics, small molecules possess simpler chemical structures and lower molecular weights, enabling oral administration through conventional dosage forms and allowing for more straightforward manufacturing processes [1]. The development and production of small molecule APIs is a complex yet well-established endeavor that integrates scientific innovation, process optimization, regulatory compliance, and scalability considerations. This chapter examines the fundamental aspects of small molecule API production, from discovery through large-scale manufacturing, with emphasis on process development, manufacturing strategies, and emerging technologies shaping the industry [2].

Overview of Small Molecule API Manufacturing

Definition and Significance

Small molecule APIs are organic compounds with molecular weights typically below 500 Daltons that serve as the therapeutically active components in pharmaceutical products. The pharmaceutical industry relies heavily on small molecule drugs for the treatment of chronic diseases, oncology, infectious diseases, and numerous other therapeutic areas[1]. Unlike biologics, which require sophisticated biotechnological manufacturing platforms, small molecule APIs can be manufactured through well-established chemical synthesis routes using standard industrial equipment and processes.

Small molecule API manufacturing demonstrates several advantages over large molecule production [3]:

- Simplified manufacturing procedures due to smaller size and lower chemical complexity
- Greater predictability and consistency in production
- Reduced regulatory burden relative to biologics
- More cost-effective manufacturing at scale
- Flexibility in formulation into diverse dosage forms (oral tablets, capsules, injectables)
- Established quality control and analytical methodologies

Market Context and Current Trends

The small molecule API market continues to demonstrate robust growth despite the increasing popularity of biologic therapies [2]. As of 2023-2025, small molecule APIs remain the predominant category in pharmaceutical development pipelines. Current trends in small molecule API manufacturing include increasing complexity in molecular structures, growing demand for highly potent active pharmaceutical ingredients (HPAPIs), and the expansion of manufacturing to support personalized medicine and niche therapy applications [2]. Additionally, the industry is witnessing rapid growth in specialized manufacturing capabilities and an emphasis on continuous processing technologies and process intensification [2].

Small Molecule API Development and Manufacturing Workflow

The journey from drug candidate identification to commercial API production encompasses multiple distinct phases, each with specific objectives, technical requirements, and regulatory considerations.

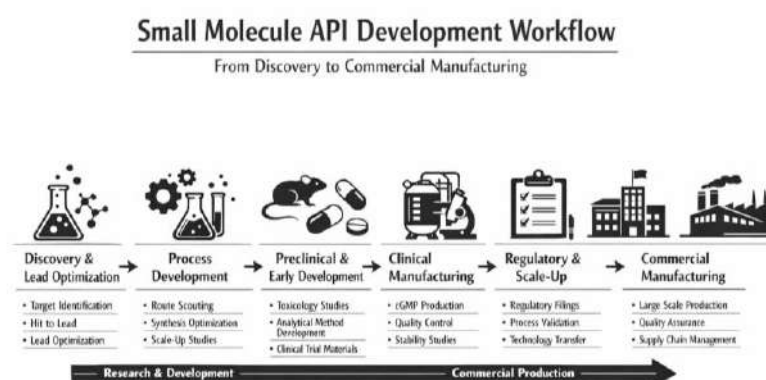


Figure 1: Small Molecule API Development Workflow from Discovery to Commercial Manufacturing

Phase 1: Drug Discovery and Target Identification

The pharmaceutical development process begins with identifying a biological target—typically a protein, enzyme, or receptor—associated with a specific disease [4]. Researchers validate that modulating this target produces a therapeutic effect using in vitro and in vivo screening

methodologies. High-throughput screening (HTS) and virtual screening techniques identify potential compounds ("hits") that interact with the target, which are then refined through iterative chemical modifications to generate "lead" compounds with improved potency, selectivity, and safety profiles [4].

Phase 2: Preclinical Development and Route Scouting

Once a lead compound is selected, process chemistry teams undertake synthetic route scouting and process development to establish an efficient, scalable synthesis pathway[3]. During this phase, multiple synthetic routes are evaluated based on:

- Number of synthetic steps required
- Availability and cost of starting materials and intermediates
- Scalability potential
- Chemical yield and selectivity
- Waste generation and environmental impact
- Safety considerations during manufacturing
- Regulatory acceptability of reagents and solvents

Parallel synthesis techniques enable rapid generation of diverse compound libraries for structure-activity relationship (SAR) optimization. The selected lead compound undergoes preclinical safety assessments, early formulation development, and small-scale synthesis (typically 1–20 grams) to support regulatory submissions for clinical trial authorization [5].

Phase 3: Clinical Development and Process Optimization

As compounds advance into clinical trials, process development scales manufacturing from milligrams to multi-kilogram quantities (20 grams to 100 kilograms) [5]. This phase demands meticulous optimization of the synthetic route under strict Good Manufacturing Practice (GMP) guidelines. Process chemists must develop robust, reproducible syntheses that deliver consistently high-purity APIs. Key activities during this phase include:

1. Route optimization for efficiency and cost reduction
2. Analytical method development for purity assessment
3. Impurity characterization and control strategies
4. Solid form selection (polymorphs, salts, or cocrystals)
5. Process chemistry modeling and in-silico optimization
6. Scale-up studies to larger pilot plant facilities
7. Regulatory documentation for chemistry, manufacturing, and controls (CMC)

Phase 4: Commercial Manufacturing Scale-Up

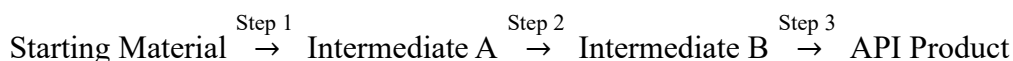
Upon regulatory approval, large-scale production (>100 kilograms) [5] commences in dedicated manufacturing facilities. At this stage, proven synthetic routes undergo final optimization for commercial viability, incorporating automation, real-time process monitoring, and quality-by-design (QbD) principles.

Process Development and Synthetic Route Optimization

Traditional Chemical Synthesis Approaches

Chemical synthesis of small molecule APIs employs well-established organic chemistry methodologies adapted for large-scale production. Most commercial APIs are synthesized through multi-step organic reactions executed in batch reactors, where precise control of temperature, pressure, and reaction time ensures product quality and consistency [3].

A representative simplified synthetic route illustrates the general approach:



The optimization of each synthetic step focuses on maximizing yield, minimizing by-products, and ensuring scalability. Critical control points include reaction selectivity (particularly for molecules containing chiral centers or multiple reactive sites), isolation and purification methodologies, and waste minimization [3].

Advanced Technologies in API Production

Continuous Manufacturing and Flow Chemistry

Continuous manufacturing represents a paradigm shift in pharmaceutical production, replacing traditional batch processes with uninterrupted flow processes that provide inherent advantages [2]:

- Real-time process monitoring and dynamic adjustment capabilities
- Enhanced reaction selectivity and improved chemical yields
- Significantly reduced production cycle times
- Seamless scalability from laboratory to commercial production
- Improved automation and reduced human error
- Superior product consistency and quality

Flow chemistry reactors enable safer handling of hazardous intermediates by maintaining controlled reaction conditions on smaller volumes and generating data continuously for process optimization [2].

Asymmetric Catalysis and Chiral Synthesis

Many small molecule APIs contain chiral centers, necessitating stereoselective synthesis to obtain single enantiomers with desired pharmacological activity [3]. Modern approaches include:

- Asymmetric catalysis using chiral catalysts
- Enzymatic methods for stereoselective transformations
- Chiral auxiliary-mediated synthesis
- Resolution techniques to separate enantiomers

Electrochemical Synthesis

Organic electrochemistry has emerged as a green and cost-efficient synthetic technology for API production [6]. Electrochemical methods promote redox transformations through electron exchange between electrodes and reactants, eliminating the need for stoichiometric quantities of hazardous oxidizing or reducing agents. This approach enhances sustainability, increases selectivity toward target compounds, and can be readily scaled to production quantities [6].

Mechanochemistry and Solid-State Synthesis

Mechanochemical methods, including liquid-assisted grinding (LAG) and bead-milling, have demonstrated remarkable utility in API manufacturing [2]. Notable applications include:

- Screening and synthesis of polymorphs, salts, and cocrystals
- Solvent-free API synthesis (exemplified by paracetamol production via bead-milling)
- Development of greener, safer synthetic routes for complex molecules
- Production of anticancer drugs such as imatinib with improved efficiency

Table 1: Comparative Overview of Manufacturing Technologies for Small Molecule APIs

Manufacturing Technology	Scalability	Waste Generation	Production Speed	Current Adoption
Traditional Batch Synthesis	High	Moderate-High	Moderate	Dominant
Continuous Flow Chemistry	High	Low	Fast	Growing
Asymmetric Catalysis	High	Low	Moderate	Established
Electrochemical Synthesis	Moderate	Very Low	Moderate	Emerging
Mechanochemistry	Moderate	Very Low	Moderate	Emerging

Quality Control and Analytical Characterization

Small molecule APIs require rigorous analytical characterization to ensure purity, identity, and safety [3]. Standard analytical methodologies include:

- **Nuclear Magnetic Resonance (NMR) Spectroscopy:** Confirms structural integrity and purity
- **Mass Spectrometry (MS):** Determines molecular weight and structural information
- **High-Performance Liquid Chromatography (HPLC):** Quantifies API purity and detects impurities
- **Fourier-Transform Infrared (FTIR) Spectroscopy:** Validates functional groups and identifies polymorph forms

- **X-ray Crystallography:** Establishes three-dimensional molecular structure

Process development must ensure consistent API quality across varying production volumes while minimizing the formation of undesired isomers, particularly for molecules containing multiple reactive sites or chiral centers[3].

Emerging Technologies and Future Directions

Artificial Intelligence and Machine Learning

Artificial intelligence and machine learning algorithms are increasingly applied to small molecule API manufacturing to predict reaction outcomes and optimize synthesis conditions [2]. These computational approaches analyze vast datasets of chemical reactions, materials, and processes to identify novel, more efficient synthetic routes. AI-powered in-silico design minimizes waste, reduces reaction times, and improves overall yields while accelerating development timelines [2].

Automation and Robotics

Advancing automation and robotic systems are streamlining laboratory workflows, reducing human error, and increasing throughput in API manufacturing. High-throughput experimentation coupled with automated data analysis enables rapid optimization of process parameters [2].

Sustainable and Green Chemistry

The pharmaceutical industry increasingly emphasizes green chemistry principles to reduce environmental impact and improve manufacturing sustainability. Integration of electrochemistry, mechanochemistry, biocatalysis, and continuous processing represents the industry's commitment to cleaner, safer API production methods[2].

Conclusion

Small molecule API production represents a mature yet dynamically evolving field at the intersection of chemistry, engineering, and regulatory science. The transition from drug discovery through clinical development to commercial manufacturing requires sophisticated process development, rigorous quality control, and strategic optimization of manufacturing technologies. Traditional batch synthesis remains dominant, yet emerging technologies—continuous flow, electrochemistry, mechanochemistry, and AI-driven optimization—are reshaping the landscape of small molecule API manufacturing. As pharmaceutical companies develop increasingly complex molecules for niche therapeutic applications and personalized medicine, the integration of advanced manufacturing technologies, automation, and computational design will be essential to meet future industry demands while maintaining rigorous quality standards and environmental sustainability.

References

- [1] Kumar, V., Bansal, V., Madhavan, A., Kumar, M., Sindhu, R., Awasthi, M. K., Saran, S. (2022). Active pharmaceutical ingredient (API) chemicals: a critical review of current biotechnological approaches. *Bioengineered*, 13(2), 4309–4327. <https://doi.org/10.1080/21655979.2022.2031412>
- [2] John M. Woodley (2018). Innovative process development and production concepts for small-molecule API manufacturing. *Computer Aided Chemical Engineering*, 41, 67-84. <https://doi.org/10.1016/B978-0-444-63963-9.00003-8>
- [3] Wahlich J. (2021). Review: Continuous Manufacturing of Small Molecule Solid Oral Dosage Forms. *Pharmaceutics*, 13(8), 1311. <https://doi.org/10.3390/pharmaceutics13081311>
- [4] Wu, K., Kwon, S. H., Zhou, X., Fuller, C., Wang, X., Vadgama, J., & Wu, Y. (2024). Overcoming Challenges in Small-Molecule Drug Bioavailability: A Review of Key Factors and Approaches. *International journal of molecular sciences*, 25(23), 13121. <https://doi.org/10.3390/ijms252313121>
- [5] Southey MWY and Brunavs M (2023) Introduction to small molecule drug discovery and preclinical development. *Front. Drug Discov.* 3:1314077. doi: 10.3389/fddsv.2023.1314077
- [6] David Cantillo. (2022). Synthesis of active pharmaceutical ingredients using electrochemical methods: keys to improve sustainability. *Chem. Commun.*, 2022,58, 619-628. <https://doi.org/10.1039/D1CC06296D>